

Global Water Research Coalition

N_2O and CH_4 emission from wastewater
collection and treatment systems
Technical Report

Report of the GWRC Research
Strategy Workshop

N₂O AND CH₄ EMISSION FROM WASTEWATER COLLECTION
AND TREATMENT SYSTEMS
TECHNICAL REPORT

REPORT

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GLOBAL WATER RESEARCH COALITION

Global cooperation for the exchange and generation of water knowledge

In 2002 twelve leading research organisations have established an international water research alliance: the Global Water Research Coalition (GWRC). GWRC is a non-profit organization that serves as a collaborative mechanism for water research. The benefits that the GWRC offers its members are water research information and knowledge. The Coalition focuses on water supply and wastewater issues and renewable water resources: the urban water cycle.

The members of the GWRC are:

KWR – Watercycle Research Institute (Netherlands), PUB – Public Utilities Board (Singapore), STOWA – Foundation for Applied Water Research (Netherlands), SUEZ Environnement – CIRSEE (France), TZW – German Water Center (Germany), UK Water Industry Research (UK), Veolia Environnement VERI (France), Water Environment Research Foundation (US), Water Quality Research Australia (Australia), Water Research Commission (South Africa), Water Research Foundation (USA), and the Water Services Association of Australia.

The US Environmental Protection Agency has been a formal partner of the GWRC since 2003. The Global Water Research Coalition is affiliated with the International Water Association (IWA).

GWRC members represents the interests and needs of 500 million consumers and has access to research programs with a cumulative annual budget of more than 150 million. The research portfolio of the GWRC members spans the entire urban water cycle and covers all aspects of resource management.

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PREFACE

The Global Water Research Coalition is an international organisation that is dedicated to the exchange and generation of knowledge to support sustainable development and management of the urban water cycle. The research agenda is developed by the member organisations of the GWRC and reflects their priorities and recognises global trends and drivers that affect the urban water cycle. The present research agenda includes Climate Change as one of the priorities areas. This research area comprises topics related to the possible impact of climate change on the urban water sector as well as the possible contribution to climate change by the urban water sector via the direct and indirect emission of greenhouse gasses (GHG).

The objective of this joint effort was to collect and develop knowledge needed to understand and manage the emission of N₂O (nitrous oxide) and CH₄ (methane) by wastewater collection and treatment systems. Starting with a kick-off meeting in Vienna in September 2008, the GWRC members involved in this activity have bundled their individual research programs on this topic, aligned methodologies used and exchanged and discussed the resulting information of the programs and developed additional actions where needed. The outcomes were reviewed and discussed at a final workshop in Montreal in September 2010.

These activities has resulted in two reports: a State of the Science report which presents an overview of the current knowledge and know-how regarding the emissions of N₂O and CH₄ by wastewater collection and treatment systems and a Technical Report which includes all the details, facts and figures of the underlying studies used to develop the State of the Science report.

GWRC expresses the wish that our joint effort and resulting reports will be useful to all who are active in the field of understanding and control of greenhouse gas emissions by wastewater collection and treatment systems.

Frans Schulting
Managing Director GWRC

ACKNOWLEDGEMENT

The Global Water Research Coalition wishes to express its appreciation to STOWA - Foundation for Applied Water Research (Netherlands) for acting as the GWRC's lead organisation for this joint effort and to recognise the high quality contributions by all organisations involved in this activity including Suez Environnement – CIRSEE (France), Water Environment Research Foundation (US), Water Research Commission (South Africa), and the Water Services Association of Australia. The support of the lead agent Stowa by Royal Haskoning is gratefully acknowledged as well.

The reports could not have been completed without the input and commitment of a number of individuals of the involved members of the GWRC and their associated organisations. These were:

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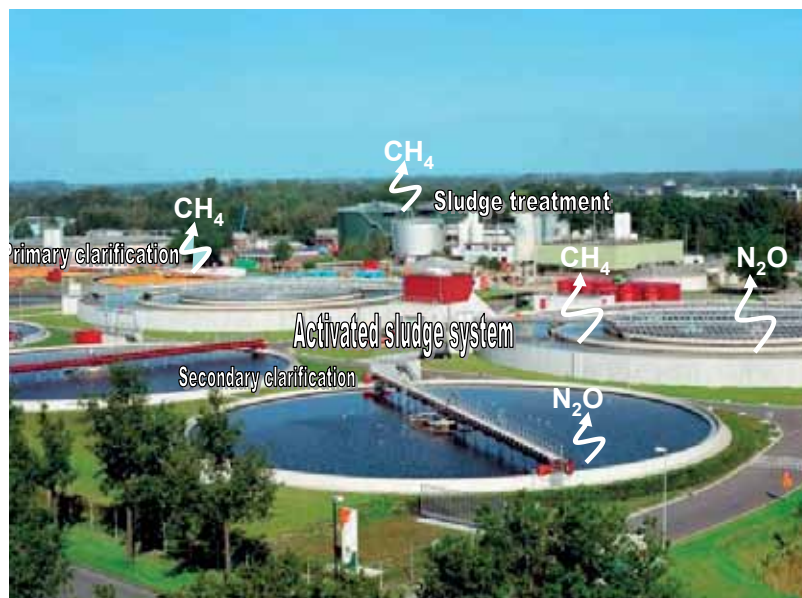
SUMMARY

BACKGROUND

In a world where there is a growing awareness on the possible effects of human activities on climate change, there is a need to identify the emission of greenhouse gases (GHG) from wastewater treatment plants (WWTPs) (See Figure i). As a result of this growing awareness, some governments started to implement regulations that force water authorities to report their GHG emissions. With these developments, there exists a strong need for adequate insight into the emissions of N₂O (nitrous oxide) and CH₄ (methane), two important greenhouse gases. With this insight water authorities would be able to estimate and finally control their emissions. However, at this point few field data were available, with the result that the emission factors used by the Intergovernmental Panel on Climate Change (IPCC) were based on limited data. The lack of available data became the driver to start extensive research programs in Australia, France, the United States of America and the Netherlands with the objective to gain information needed to estimate, understand and control the emission of N₂O and CH₄ from wastewater collection and treatment systems.

FIGURE 1

GREENHOUSE GAS EMISSION FROM WASTEWATER TREATMENT PLANTS



CURRENT KNOWLEDGE

At the start of the research programs little was known about the processes which form N₂O, in contrast with the extensive knowledge on the formation of methane. In both cases, however, very little field data were available that gave insight on the level at which these two greenhouse gases were emitted from wastewater collection and treatment systems.

This lack of data resulted in the fact that the currently used IPCC emission factor for N₂O (3.2 g N₂O·person⁻¹·year⁻¹), which is used to estimate the N₂O emission from wastewater treatment plants, is based on only one field study in which the plant was not designed to remove nitrogen. Furthermore this lack of data has led the IPCC to conclude that: "wastewater in closed underground sewers is not believed to be a significant source of methane" (IPCC, 2006 a,b).

The data that has been published prior to the start of the research programs showed a very large variation in the level of N₂O emission. This is due to the fact that the formation of N₂O is a very complex process which can be performed by both nitrifying and denitrifying bacteria and is influenced by several process parameters. Denitrification in anoxic zones was in many cases indicated as the dominant source of N₂O emission from biological nitrogen removal processes.

JOINT EFFORTS

Since the topic of greenhouse gas emission from wastewater collection and treatment collection systems is of significance for the whole sector, the GWRC members¹ decided to join their individual research program results and support collaboration between their individual research partners. These joint efforts have led to an increased level of understanding on the processes forming N₂O emission from wastewater treatment facilities, the variety therein, and the contribution of methane emission from sewers and WWTPs. This increased level of understanding can already be used by the stakeholders of the GWRC members who are directly involved in the daily operation of wastewater collection and treatment systems.

Adjacent to the joint efforts of the GWRC members and individual research partners, the International Water Association (IWA) formed a Task group on the use of water quality and process models for minimising wastewater utility greenhouse gas footprints. The IWA Task Group is also collaborating with the GWRC researchers.

OBJECTIVES

The overall objectives of the different research programs were:

- Define the origin of N₂O emission.
- Understand the formation processes of N₂O.
- Identify the level of CH₄ emissions from wastewater collection and treatment systems.
- Evaluate the use of generic emission factors to estimate the emission of N₂O from individual plants.

BOUNDARIES

The main focus was to identify the level of emission, the variation therein and improve the knowledge of N₂O formation. Definition of mitigation strategies was outside the scope of most of the research as the knowledge on formation and origin was too limited at the start of the research programs.

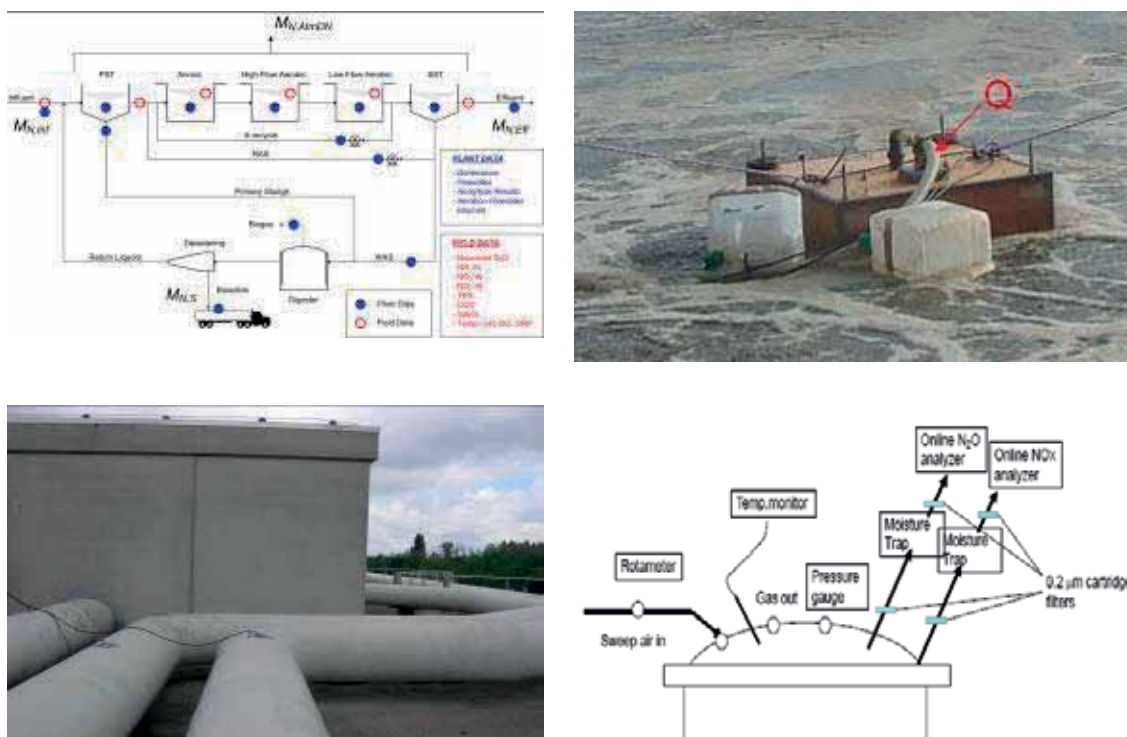
¹ GWRC members were (in brackets the partner that performed the research): WERF, USA (Columbia University, Brown and Caldwell); WSAA, Australia (The University of Queensland); STOWA, the Netherlands (Delft University of Technology; Royal Haskoning)

RESEARCH N₂O

METHODOLOGY

In all participating countries a wide range of WWTP types was selected with the expectation that differences between plant design and process conditions can help elucidate the factors influencing N₂O formation. The individual research partners used different methodologies (see Figure ii) to determine the emission of N₂O. The methodologies used in Australia, France, and the USA² were very suitable to gain insight in the formation processes of N₂O. The methodology used in the Netherlands, where the N₂O emission was measured in the total off-gas of covered WWTPs was very suitable to capture the variability of the emission. The use of different methodologies shows the complementary value of joint efforts to increase the level of knowledge on N₂O emission from WWTPs. For future work on this topic both methodologies will be required to finally estimate and control the emission of N₂O from WWTPs.

FIGURE II APPLIED METHODOLOGIES IN THE DIFFERENT RESEARCH PROGRAMS. STARTING IN THE LEFT CORNER ABOVE AND THEN CLOCKWISE: MASS BALANCE METHOD BASED ON LIQUID GRAB SAMPLES (AUSTRALIA); SAMPLING BOX FOR AERATED AREAS (FRANCE); TOTAL OFF-GAS MEASUREMENTS (THE NETHERLANDS); U.S. EPA, SURFACE EMISSION ISOLATION FLUX CHAMBER (SEIFC); (USA).



RESULTS

The emission of N₂O has been determined with different measurement protocols. For this reason it is not possible to average the emission numbers that have been derived. The results obtained in this research were suitable to increase the knowledge on N₂O formation and the variation therein, but the numbers can not be used to determine the emission from an individual plant as will be explained hereafter.

In line with earlier data, the field data in this study showed a large variety among the WWTP's

- 2 The protocol developed in the United States has been accepted by the USEPA, and is one of the most significant outputs of the research program.

sampled in the participating countries. The lowest emission that was measured was lower than 0.0001 kg N₂O-N/kg TKN_{influent}, while the highest reported emission was as high as 0.112 kg N₂O-N/kg TKN_{influent}. This led to the following conclusions:

- The N₂O emission is highly variable among different WWTPs and at the same WWTP during different seasons or throughout the day.
- The use of a generic emission factor to estimate the emission from an individual WWTP is inadequate
- The emission from an individual WWTP can only be determined based on online measurements over the operational range of the WWTP (i.e. lowest temperature, highest load etc).

On the origin of the emission results showed that:

- The emission of N₂O mainly originates from nitrification, in contrast with earlier information.

At the start of the different research studies, very little was known about the process parameters that influenced the formation of N₂O, and most of the knowledge was based on laboratory studies. The joint efforts of the GWRC members and their research partners led to an increased level of understanding of the formation of N₂O and the process parameters influencing formation. It was concluded that:

- Nitrite accumulation leads to the formation of N₂O in aerobic zones as a result of low oxygen levels, sudden changes in ammonium load, and higher temperatures.
- High ammonium concentrations can lead to the emission of N₂O if nitrification occurs.

The above conclusions could already be translated to practice, in a way that if high concentrations of nitrite, ammonium or dissolved oxygen can be avoided the risk of N₂O emission can be reduced. It was concluded that:

Systems that are not designed to remove nitrogen will have a high risk of N₂O emission if unintentional nitrification occurs.

With the present insight, it is possible to estimate the risk for N₂O emissions from a specific WWTP. This estimation can be based on the risk matrix presented in the following Table:

Parameter	Risk on N ₂ O		
	High risk	Medium risk	Low risk
Effluent total organic nitrogen (mg/l)	> 10	5 - 10	< 5
Range in N-concentration in plant	H	M	L
Load variations (daily)	H	M	L
Maximum NO ₂ concentration (mg N/l) anywhere in plant	> 0.5*	0.2 - 0.5	0.2

* Risk does not increase at higher NO₂ concentrations

Based on the above matrix and the other conclusions the major conclusion of the research performed on N₂O emission from WWTPs is:

A good effluent quality (TN < 5 mgN/l) goes hand in hand with a low risk of N₂O emission

REMAINING KNOWLEDGE GAPS AND FUTURE RESEARCH

Based on the outcomes of the research, valuable knowledge was gained to estimate and control the emission of N₂O from wastewater collection and treatment systems. The remaining knowledge gaps, their objectives and the type of research required are summarised as follows:

Knowledge gap	Objective	Future research
Insight in the variability of N ₂ O emission throughout the year at a WWTP to be able to define guidelines to design a sampling program at uncovered plants.	To obtain a good emission estimate of individual plants with minimal uncertainty.	Long term measurements in the total off-gas of WWTPs (covered ones are the most suitable to do so).
The relative contribution of autotrophic and heterotrophic processes to N ₂ O generation.	To develop mitigation strategies.	High resolution monitoring of liquid phase N ₂ O specific zones of WWTP.
Mitigation strategies.	To define measures to control emission via process design and control.	Measurements at different zones of one specific WWTP to study effect of different measures.
Emission from unknown sources like biofilm based processes and receiving aquatic environment.	To define level of N ₂ O emissions from these sources and to complete the picture of the whole urban watercycle.	Measurements at several locations that capture the variability that is expected.

RESEARCH CH₄

METHODOLOGY

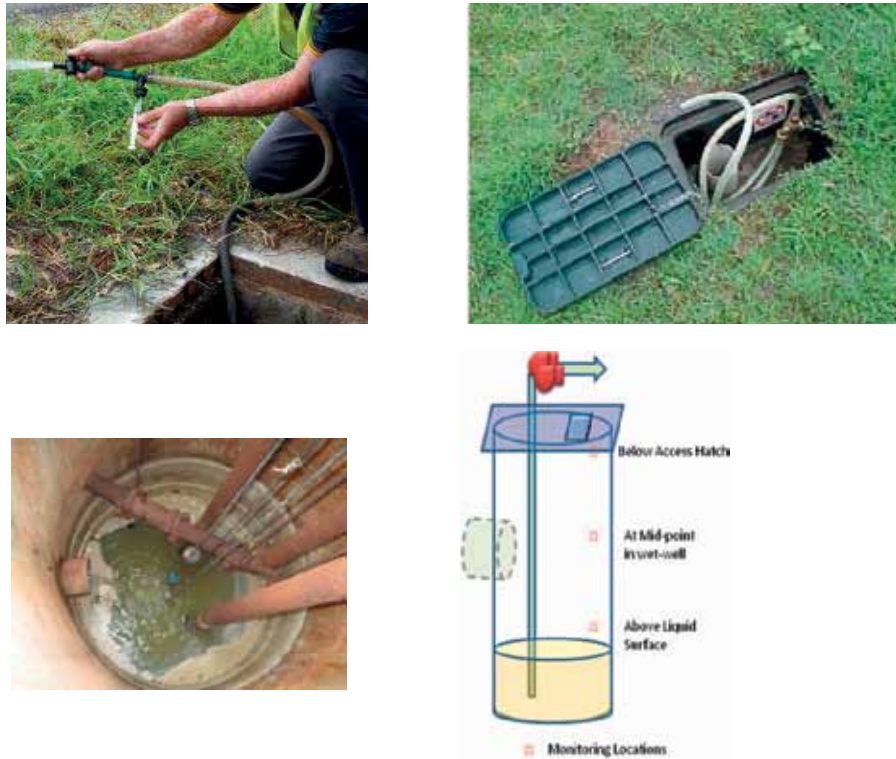
The emission of methane was determined both from wastewater collection and treatment systems. The emission from wastewater collection systems was performed in Australia and the United States of America (see Figure iii). In Australia measurements were made in the liquid and gas phase in or around raising mains. The gas phase of unventilated lift stations was analysed in a study from the United States of America. A major obstacle in finally determining the emission of CH₄ (kg/d) from sewers is the determination of the gas flow (m³/d). Developing a strategy for this obtaining flow measurement is one of the major research topics in this area. Mitigation strategies to control the emission of CH₄ from sewers were tested on laboratory and field level in Australia.

The emission of CH₄ from wastewater treatment systems was investigated in France and the Netherlands. In France, the emission of CH₄ was monitored via a gas hood that was placed at the surface of different zones in a WWTP.

The emission of CH₄ in the Netherlands was determined based on grab samples taken from the different process units. These samples were taken in the same period as the emission of N₂O was monitored. In this way the carbon footprint of a WWTP could be determined as the data of electricity and natural gas use were readily available.

FIGURE III

ABOVE: SAMPLING SYSTEM RISING MAINS (AUSTRALIA); UNDER: SAMPLING SYSTEM UNVENTILATED LIFT STATIONS (USA)



RESULTS

At the start of the research, very little was known about the level of CH₄ emission from sewers and WWTP; the emission from sewers was even neglected. The results showed that the methane concentration in the liquid and gas phase from wastewater collection and treatment can be substantial. Concentrations up to more than 30 mg/l in the liquid phase were reported and emissions from lift stations were found to be as high as ~700 kg CH₄/year, but also emissions close to zero were found. This led to the following conclusion:

- Formation and emission from wastewater collection systems can be substantial and should not be neglected.

Measurements to define the emission of CH₄ (i.e. kg/d) from sewerage systems were found to be very difficult and complicated. Development of a good strategy measurement is seen as an important research topic.

Furthermore, a start was made to find strategies that could control the emission of CH₄ from sewers. Based on these preliminary experiments it was concluded that:

- Odour mitigation strategies in sewers likely also supports reduced CH₄ formation.

The level of CH₄ emission from WWTPs varied greatly from almost zero emission (< 0.0004 kg CH₄-COD/kg COD_{influent}) to emissions as high as 0.048 kg CH₄-COD/kg COD_{influent}). In general it was concluded that:

- Emission of CH₄ from WWTPs mainly originates from CH₄ formed in sewers and from sludge handling processes.

REMAINING KNOWLEDGE GAPS AND FUTURE RESEARCH

Based on the outcomes of the research valuable knowledge was gained to estimate and control the emission CH₄ from wastewater collection and treatment systems. The knowledge gaps, their objectives and the type of research required are summarised as follows:

Knowledge gap	Objective	Future research
Strategy to determine amount of gas emitted to the air from wastewater collection systems.	To define the emission (kg/d) of CH ₄ from wastewater collection systems.	Develop a strategy based on field data.
Field data from different type of wastewater collection systems around the world.	To make a good estimate of the contribution of wastewater collection systems. To deliver data for the development, calibration and validation of CH ₄ emission models.	Field measurements both liquid and gas phase from rising mains and gravity sewers around the world.
Cost effective mitigation strategies.	To control the emission of CH ₄ from wastewater collection systems.	Experiments in practice to study the effects and costs of different mitigation strategies.
Emission from sludge treatment lagoons.	To define level of CH ₄ emissions from this source.	Measurements at several locations that capture the variability that is expected.

TOTAL CARBON FOOTPRINT

As a first indication on the possible contribution of N₂O and CH₄ emission to the total carbon footprint of a WWTP, the result in the Netherlands could be used as an example.

In the case studies in the Netherlands, the specific emissions of N₂O and CH₄ were determined at the same time. Together with the data on the related consumption of electricity and natural gas, it was possible to calculate a carbon footprint of three WWTPs. To determine the carbon footprint, all sources were converted to CO₂ equivalents³. The results in the Netherlands indicated that the emission of CH₄ and N₂O can significantly contribute to the total carbon footprint of a WWTP. This contribution can vary from 2% to almost 90% of the carbon footprint under extreme conditions for N₂O and 5 – 40% for CH₄. One should be aware that these numbers are specific for the Netherlands. In any other country, these numbers can differ greatly as there exist a great variation in the way wastewater and sludge is handled as well as the specific composition of the energy mix used. Furthermore these numbers can significantly differ depending on how the boundaries are set around the analysis. In case of the analysis performed for the three Dutch WWTPs the contribution of e.g. chemical use, and sludge incineration were not accounted for.

FUTURE ACTIVITIES

In the future the following activities will be developed by GWRC members and their researchers to further estimate and control the emission of GHG from wastewater collection and treatment systems:

- Long term measurements of both N₂O formation and process variables from one WWTP, to gain insight in N₂O formation processes and the variability throughout the year.
- Mitigation strategies to gain insight in the possibilities to control the emission via process design and control.
- Development of a predictive model on N₂O production and emission.

³ It should be noted that the conversion numbers are country specific and do depend on the used energy mix (i.e. brown coal versus wind or solar energy), which is of influence on the total carbon footprint of a WWTP.

DE STOWA IN BRIEF

The Foundation for Applied Water Research (in short, STOWA) is a research platform for Dutch water controllers. STOWA participants are all ground and surface water managers in rural and urban areas, managers of domestic wastewater treatment installations and dam inspectors.

The water controllers avail themselves of STOWA's facilities for the realisation of all kinds of applied technological, scientific, administrative legal and social scientific research activities that may be of communal importance. Research programmes are developed based on requirement reports generated by the institute's participants. Research suggestions proposed by third parties such as knowledge institutes and consultants, are more than welcome. After having received such suggestions STOWA then consults its participants in order to verify the need for such proposed research.

STOWA does not conduct any research itself, instead it commissions specialised bodies to do the required research. All the studies are supervised by supervisory boards composed of staff from the various participating organisations and, where necessary, experts are brought in.

The money required for research, development, information and other services is raised by the various participating parties. At the moment, this amounts to an annual budget of some 6,5 million euro.

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N₂O AND CH₄ EMISSION FROM WASTEWATER COLLECTION AND TREATMENT SYSTEMS

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1

INTRODUCTION

1.1 BACKGROUND

In a world where there is a growing awareness of the possible effects of human activities on climate change, there is a need to identify the emission of greenhouse gases (GHG) from wastewater treatment plants (WWTPs)⁴. As a result of this growing awareness, governments started to implement regulations that require water authorities to report their GHG emissions. With these developments there exists a strong need for adequate insight into the emissions of N₂O and CH₄. With this insight water authorities would be able to estimate and finally reduce their emissions. At the time little information was available on the formation of GHG, and the emission factors used by the IPCC are based on limited data. The limits of available data became the driver to start extensive field studies in Australia, France, the United States of America and the Netherlands with the objective to fill the knowledge gaps needed to estimate and reduce the emission of N₂O and CH₄ from wastewater collection and treatment systems.

The research programs were performed by partners⁵ of the GWRC members WERF (United States of America), WSAA (Australia), CIRSEE-Suez (France) and STOWA (the Netherlands).

1.2 OBJECTIVES

The overall objectives of the different research programs⁶ were:

- Define the origin of N₂O emission.
- Understand the formation processes of N₂O.
- Identify the level of CH₄ emissions from wastewater collection and treatment systems.
- Evaluate the use of generic emission factors to estimate the emission of N₂O from individual plants.

1.3 ACTIVITIES WITHIN THE GLOBAL WATER RESEARCH FRAMEWORK

The topic of N₂O emissions from wastewater treatment facilities is part of the research area Climate Change of the joint research agenda of the Global Water Research Coalition (GWRC). STOWA took the lead to develop and coordinate this joint activity with support of the GWRC members Anjou Recherche, Eawag, CIRSEE, UKWIR, WERF, WRC and WSAA. Representatives

- 4 The greenhouse gases associated with the activities at WWTPs are CO₂, CH₄ and N₂O. Of these gases, N₂O is the most important as it has a 300-fold stronger effect than CO₂. CH₄ is less strong than N₂O but still has a 25-fold stronger effect than CO₂. Nitrous oxide (N₂O) can be formed during the conversion of nitrogenous compounds in wastewater; methane may be emitted in the sewer system and during sludge handling. The emission of CO₂ from the biological treatment is part of short cycle (or biogenic) CO₂ and does not contribute to the carbon footprint. However, some carbon in wastewater may originate from fossil fuel.
- 5 Partners were: Columbia University, USA; Brown and Caldwell, USA; The University of Queensland, Australia; Delft University of Technology, the Netherlands, Royal Haskoning, the Netherlands.
- 6 In the technical report (GWRC, 2011) that accompanies this State of the Art Report the objectives of the individual partners are mentioned.

of the involved members have met on several occasions making use of opportunities of planned conferences and workshops like the WWC in Vienna (September 2008), the GWRC workshop in Dübendorf (February 2009), and a meeting on the occasion of the SIWW/LET 2009 in Singapore (June 2009). An inventory of members research programs was performed by STOWA and detailed information on the ongoing efforts was discussed and protocols exchanged.

In August 2009 the GWRC N₂O website was launched and involved GWRC-members (and invited experts) can use the site to exchange information and comment results.

At present the members of the GWRC have either initiated or are planning to undertake research to measure the emission of N₂O from wastewater treatment facilities. An extensive research program was set up in Australia, the Netherlands and the United States of America to quantify the emission of N₂O and CH₄ from sewers and WWTPs. In these research programs there was a focus on the emission of N₂O, the emission of CH₄ was studied in less detail. The reason for this difference in focus is the fact that N₂O is a much stronger greenhouse gas than CH₄ and that little is known about the formation processes of N₂O in WWTPs.

1.4 ONGOING ACTIVITIES OUTSIDE GWRC

Besides the activities of the GWRC members, a new IWA Task Group will focus on the use of water quality and process models for minimizing wastewater utility greenhouse gas footprints. The main objectives of this group are:

- Understand the processes that are responsible for the major contributions to GHG emissions from WWTP and sewer systems.
- Incorporate this knowledge into mathematical models that can be embedded in system/plant-wide models allowing multi-criteria optimisation.

The World Bank, with partners, has financed an ongoing project at the Rio Frio wastewater treatment plant in Columbia to reduce CH₄ and N₂O emissions. The project had several objectives, including:

- Improvements in gas separation in the anaerobic reactors and during gas engines to result in additional abatement of CH₄ emissions.
- The reduction in N loads in the receiving waters will result in a corresponding reduction in N₂O.

1.5 BOUNDARIES REPORT

The research described in this report was the first extensive research on N₂O and CH₄ emission from wastewater collection and treatment systems. The main focus was to identify the level of emission, the variation therein and improve the knowledge on N₂O formation. Definition of mitigation strategies was outside the scope of most of the research as the knowledge on formation and origin was too limited at the start of the research. For methane some mitigation strategies were investigated and are reported here.

1.6 OUTLINE REPORT

This report extensively describes the field and laboratory studies that have been performed

in Australia, France, the United States of America and the Netherlands and presents a higher level of detail than the state of the art report on the topic (GWRC, 2011).

An extensive literature review is presented in chapter 2. The local regulations as they apply in countries participating in the GWRC report are presented in chapter 3. The individual objectives of the projects are presented in chapter 4. The methodology used by the individual countries is given in chapter 5. An overview of all the results is presented in chapter 6 and this is discussed in chapter 7. Finally the conclusions and recommendations for future research are presented in chapter 8. The following reports of the individual GWRC members were used:

- WERF: Chandran, K., 2010, Greenhouse nitrogen emission from wastewater treatment operations, WERF report U4R07a.
- WSAA: Foley, J., Lant, P., 2009, Direct Methane and Nitrous oxide emissions from full-scale wastewater treatment systems, Occasional paper No.24, Water Service Association of Australia.
- STOWA: Voorthuizen van, E.M., van Leusden, M., Visser, A., Kruit, J., Kampschreur, M., Dongen van, U., Loosdrecht van, M., 2010, Emissies van broeikasgassen van rwzi (in Dutch, summary in English), STOWA report 2010-08.

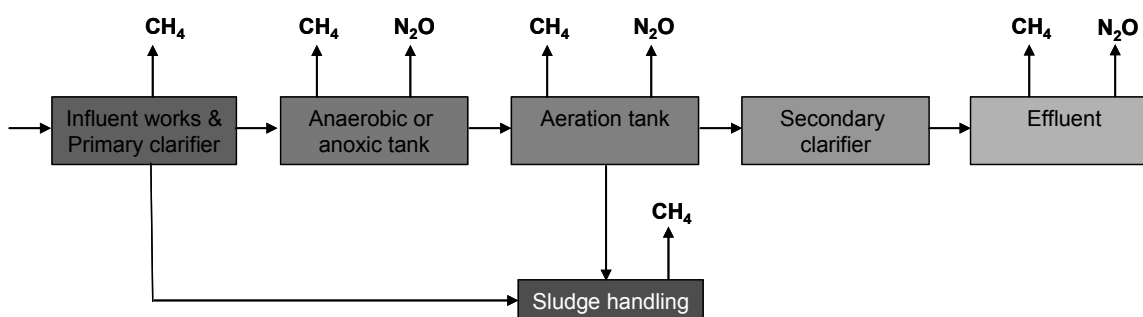
2

LITERATURE REVIEW

2.1 NON CO₂ GREENHOUSE GASES

The non CO₂ greenhouse gases that can be emitted from a domestic WWTP are nitrous oxide (N₂O) and methane (CH₄). The locations at a WWTP where these gases can be emitted are presented in Figure 1.

FIGURE 1 SCHEMATIC OVERVIEW OF A DOMESTIC WWTP AND THE LOCATIONS WHERE CH₄ AND N₂O CAN BE EMITTED



Methane that is emitted from the influent works is most likely formed in the sewer system, as the retention time of the wastewater in the influent works is too short to form CH₄. Furthermore CH₄ formation will only occur where anaerobic or anoxic conditions prevail, as in the anaerobic or anoxic tank, but then only in the biofilms at the side of tanks, and at sludge handling sites. For this reason no CH₄ formation is expected in an aeration tank. Methane that is emitted here is formed earlier (in sewer or in sludge digester) and is stripped to the gas phase in the aeration tank. Formation and emission of N₂O can only occur under anoxic or aerobic conditions in the presence of nitrate (and carbon source) and ammonium. Nitrogen that is not converted leaves the WWTP via the effluent, which can lead to the emission of N₂O from surface water.

2.2 RELEVANT PROCESSES N₂O FORMATION

Nitrous oxide can be produced during the conversion of nitrogen in WWTPs. The processes involved are nitrification and denitrification. Besides N₂O formation by biological processes in activated sludge systems, there can be N₂O generation when e.g. biogas is burned at the WWTP for electricity production.

2.2.1 NITRIFICATION

Nitrification is performed by three different groups of autotrophic microbes; ammonium-oxidizing bacteria (AOB) and ammonium-oxidizing archaea (AOA) that convert ammonia into nitrite, and nitrite-oxidizing bacteria (NOB) that convert nitrite into nitrate. The different steps involved in the nitrification are presented in Figure 2.

FIGURE 2 CONVERSION STEPS IN THE NITRIFICATION PROCESS (AS PRESENTED IN COLLIVER, 2000)

AOB / AOA			
	$\text{NH}_3 + \text{O}_2 + 2\text{H}^+ + 2\text{e}^-$	\rightarrow	$\text{NH}_2\text{OH} + \text{H}_2\text{O}$
	$\text{NH}_2\text{OH} + \text{H}_2\text{O}$	\rightarrow	$\text{NO}_2^- + 5\text{H}^+ + 4\text{e}^-$
	$0.5 \text{O}_2 + 2\text{H}^+ + 2\text{e}^-$	\rightarrow	H_2O
Total	$\text{NH}_3 + 1.5\text{O}_2$	\rightarrow	$\text{NO}_2^- + \text{H}^+ + \text{H}_2\text{O}$
NOB			
	$\text{NO}_2^- + \text{H}_2\text{O}$	\rightarrow	$\text{NO}_3^- + 2\text{H}^+ + 2\text{e}^-$
	$0.5\text{O}_2 + 2\text{H}^+ + 2\text{e}^-$	\rightarrow	H_2O
Total	$\text{NO}_2^- + 0.5\text{O}_2$	\rightarrow	NO_3^-

Even though N₂O is not present as an intermediate in the main catabolic pathway of nitrification, AOB are known to produce N₂O. This has predominantly been associated with denitrification capacity of AOB. AOB contain the enzymes to reduce NO₂⁻-N and NO with N₂O as final product. Note that these enzymes are the same as in regular denitrifying bacteria, but that in AOB denitrification is not associated with growth.

2.2.2 DENITRIFICATION

Denitrification is performed by a metabolically very diverse group of micro-organisms, bacteria as well as archaea, which couple oxidation of organic or inorganic substrates to reduction of nitrate, nitrite, NO and N₂O. As N₂O is an intermediate in the denitrification process, incomplete denitrification can lead to N₂O emission. Many denitrifying micro-organisms are facultative denitrifiers, which preferentially use oxygen as electron acceptor, due to the higher energy yield. The different steps involved in the denitrification are presented in Figure 3.

FIGURE 3 CONVERSION STEPS IN THE DENITRIFICATION PROCESS (AS PRESENTED IN OTTE, 2000)

	$2^- + 4\text{H}^+ + 4\text{e}^-$	\rightarrow	$2\text{NO}_2^- + 2\text{H}_2\text{O}$
	$2\text{NO}_2^- + 4\text{H}^+ + 2\text{e}^-$	\rightarrow	$2\text{NO} + 2\text{H}_2\text{O}$
	$2\text{NO} + 2\text{H}^+ + 2\text{e}^-$	\rightarrow	$\text{N}_2\text{O} + \text{H}_2\text{O}$
	$\text{N}_2\text{O} + 2\text{H}^+ + 2\text{e}^-$	\rightarrow	$\text{N}_2 + \text{H}_2\text{O}$
Total	$2^- + 12\text{H}^+ + 10\text{e}^-$	\rightarrow	$\text{N}_2 + 6\text{H}_2\text{O}$

2.2.3 CHEMICAL REACTIONS

Possible chemical pathways leading to N₂O formation in WWTPs are the reaction between nitrite and hydroxylamine leading to NO and N₂O and nitrite reductions with organic or inorganic compounds (Van Cleemput, 1998). In the first reaction the intermediate hydroxylamine production by AOB is required, complicating the distinction between chemical and biological N₂O production (paragraph, Kampschreur, 2009).

2.3 PROCESS PARAMETERS INFLUENCING N₂O FORMATION

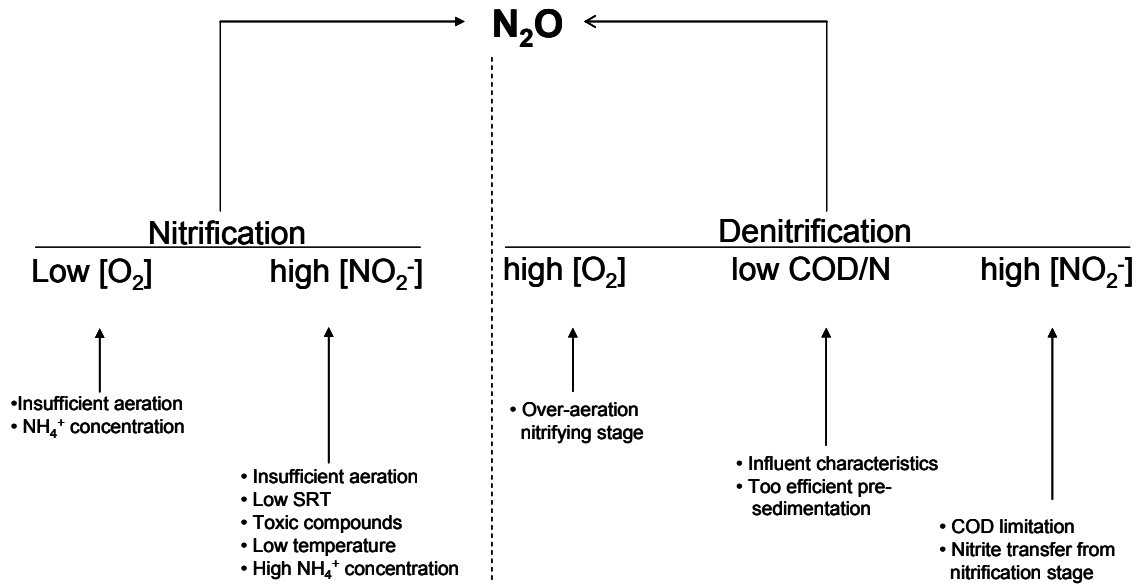
Nitrous oxide emission has been extensively studied for soil systems. Reports about the emission of N₂O from activated sludge were only reported since the early nineties. An overview of all research on the emission of N₂O from WWTPs is presented in Table 1. In the same table an overview is presented of the research performed at laboratory scale.

From Table 1 it can be observed that there is a large variation in N₂O emission among the investigated WWTPs. This variation can be understood from the fact that N₂O can be formed both during nitrification and denitrification, and that different process parameters influence

the formation of N₂O during these processes (see Figure 4⁷). The scale and frequency of changes in process conditions depend on the type of WWTP, reactor geometry, mixing intensity and aeration mode (Kampschreur, 2009).

In laboratory studies the process conditions typically change more rapidly than in large WWTPs, and this explains the higher level of variation in N₂O emission at laboratory scale as can be seen from Table 1. In addition more N₂O is stripped from laboratory scale systems than from full scale systems because aeration is often used as a means of mixing in laboratory reactors as well as for provision of oxygen.

FIGURE 4 MAIN PROCESS PARAMETERS LEADING TO N₂O FORMATION AND EMISSION



⁷ The process parameters mentioned will be discussed from page 9 onwards.

TABLE 1 OVERVIEW OF N₂O EMISSION AT FULL SCALE AND LABORATORY SCALE (FROM KAMPSCHREUR, 2009)

Type of WWTP	N ₂ O emission (% of N-load)	Sample frequency	Remarks	Reference
Full scale				
Activated sludge (11.000 p.e.)	0.035% ¹	Weekly grab sample (vented chamber) over 15 weeks		Czepiel et al., 1995
Twenty-five activated sludge plants	0 -14.6% (0.6% average)	Single grab samples per WWTP	N ₂ O emission increased with increasing nitrogen load	Wicht and Beier, 1995
Activated sludge plant (60.000 p.e.)	0.001%	Twice-weekly grab samples over a year	N ₂ O emission increased with nitrite+nitrate concentrations	Sümer, 1995
Activated sludge plant (60.000 p.e.)	0.02%	Once or twice-weekly grab samples during 1.5 years		Sommer et al., 1998
Activated sludge plant (1.000 p.e.)	0.01 – 0.08%	On-line measurements during four aeration cycles (2 h)	N ₂ O emission decreased with proportionally shorter aeration periods	Kimochi et al., 1998
Nitrification – anammox sludge water treatment	2.3% ²	On-line measurements during four days	N ₂ O emission increased with decrease in oxygen concentration (aerated stage) and increase of nitrite concentration (anoxic stage)	Kampschreur et al., 2008b
Nitrification stage (nitrogen removal stage) of activated sludge plant (620.000 p.e.)	4%	Three grab samples during one day		Kampschreur et al., 2008b
Laboratory scale				
Continuous and batch tests denitrifying activated sludge – artificial wastewater	0 – 8%	Grab samples	N ₂ O emission increased with decrease of COD/N and increasing nitrite concentration	Hanaki et al., 1992
Continuous nitrifying activated sludge – artificial wastewater	2.3 – 16%	Daily grab samples	N ₂ O emission increased with decreasing oxygen concentration and SRT	Zheng et al., 1994
Batch test (10 h) denitrifying activated sludge – real wastewater	0 – 6%	On-line measurement	N ₂ O emission increased with increasing oxygen concentration	Schulthess et al., 1994
Continuous oxic-anoxic SBR activated sludge – real high-strength wastewater (> 50 d)	1-35%	Grab samples	N ₂ O emission higher with longer aeration period in one SBR cycle, probably linked to increased nitrite levels	Osada et al., 1995
Batch tests denitrifying activated sludge – real wastewater	1.2%	Grab samples	N ₂ O emission increased with decrease of pH	Thoern and Soerensson, 1996
Continuous oxic-anoxic SBR activated sludge – artificial wastewater (380 days)	5-95%	Grab samples	N ₂ O emission decreased over time – increased N ₂ O emission possibly a start-up phenomenon	van Benthum et al., 1998
Batch tests denitrifying activated sludge – artificial wastewater	0.005 -0.5%	Grab samples	N ₂ O emission increased with decrease of COD/N	Chung and Chung, 2000
Continuous nitrifying and denitrifying activated sludge – real wastewater	0.2-4.5%	Grab samples	N ₂ O emission decreased upon methanol addition for higher COD/N ratio	Park et al., 2000

Type of WWTP	N ₂ O emission (% of N-load)	Sample frequency	Remarks	Reference
Continuous oxic-anoxic SBR activated sludge – artificial high-strength wastewater	0.5 – >20%	On-line measurement	N ₂ O emission increased with decrease of COD/N	Itokawa et al., 2001
Continuous nitrifying activated sludge – artificial wastewater	0.08 – 1.17%	On-line measurement	N ₂ O emission increased with increasing ammonia shock loads and nitrite concentration	Burgess et al., 2002
Continuous activated sludge – real wastewater (50 days)	0.2 – 1.5%	Weekly grab samples	N ₂ O emission increased with decreasing oxygen concentration and decreasing SRT	Noda et al., 2003
Continuous anaerobic-anoxic SBR activated sludge – artificial wastewater	90%	On-line measurement	N ₂ O emission increased with increasing nitrite concentration – N ₂ O might be major product of denitrification by GAO ³	Zeng et al., 2003
Continuous oxic-anoxic SBR activated sludge – artificial wastewater	>50%	On-line measurement	High N ₂ O emission (during oxic stage) was hypothesized to be caused by low diversity of population due to single carbon source	Lemaire et al., 2006
Continuous nitrifying activated sludge – artificial wastewater (30-300 days)	0.2 – 0.5%	Daily grab samples	N ₂ O emission ratio increased with increasing salt concentration	Tsuneda et al., 2005
Continuous denitrifying activated sludge – artificial wastewater (30-300 days)	0.005 – 0.02%	Daily grab samples	N ₂ O emission independent of salt concentration	Tsuneda et al., 2005
Continuous oxic-anoxic activated sludge – artificial wastewater (30-300 days)	0.7 -13%	Daily grab samples	N ₂ O emission increased with increasing salt concentration	Tsuneda et al., 2005
Batch test (5 h) nitrifying activated sludge- real wastewater	0.1 – 0.4% ⁴	On-line measurement	N ₂ O emission is largest at 1.0 mg O ₂ /L and lower above and below this oxygen concentration – emission increases with nitrite concentration	Tallec et al., 2006a
Batch test (1 day) nitrifying biofilter – real wastewater	0.2 – 1% (0.4% average)	On-line measurement	N ₂ O emission ratio increased with decreasing oxygen concentration ⁵	Tallec et al., 2006b
Batch test (1 day) denitrifying biofilter – real wastewater + methanol	% -1.3% (0.2% average)	On-line measurement	N ₂ O emission ratio increased with decreasing methanol addition	Tallec et al., 2006b
Batch test (5 h) nitrifying activated sludge- real wastewater	0.4%	On-line measurement	N ₂ O emission is largest at 0.3 mg O ₂ /L and lower above and below this oxygen concentration	Tallec et al., 2008
Continuous nitrifying SBR activated sludge – artificial wastewater	2.8%	On-line measurement	N ₂ O emission increased with decreasing oxygen and increasing nitrite concentration	Kampschreur et al., 2008a

¹ Based on the assumption of 100 g protein/person/day and 0.16 gN/gprotein (FAO-statistics, see also IPCC (2006)), the value of 3.2 gN₂O/person/year is converted into a ratio. This study is the basis for IPCC calculations.

² Sum of load based N₂O emission of nitrification reactor (1.7%) and anammox reactor (0.6%)

³ GAO Glycogen Accumulating Organisms

⁴ Ratio is expressed per N-ammonia converted instead of N-load, because nitrate is present in the wastewater which is not converted here.

⁵ The overall N₂O emission decreased with decreasing oxygen concentration because of the lower ammonia oxidation rate due to oxygen limitation

OXYGEN CONCENTRATION

A low dissolved oxygen concentration can lead to the formation of N₂O during the nitrification stage. Under these conditions autotrophic ammonia oxidizers use nitrite as the terminal electron acceptor to save oxygen for the oxygenation reaction of ammonia to hydroxylamine (Kampschreur, 2009). This process is referred to as the nitrifier denitrification. A low oxygen concentration can occur in the nitrification stage due to insufficient aeration or as a result of a high ammonium peak. The effect of a high ammonium peak was studied by Burgess et al. (2002) and Gjelsberg (1998), both found a decrease in the oxygen concentration followed by an increase in the nitrite concentration and N₂O emission.

Excessive aeration rates in the nitrification tank can lead to a substantial oxygen concentration in the denitrification tank. The consequence of this is that oxygen inhibits both synthesis and activity of denitrification enzymes. One of these, N₂O reductase is more sensitive to oxygen than the other enzymes, leading to N₂O emission during denitrification when oxygen is present in low amounts (Otte et al. 1996).

NITRITE

High nitrite concentrations in both nitrification and denitrification stages lead to an increase in N₂O emission. During nitrification increased nitrite concentrations lead to increased denitrification (effectively nitrite reduction to N₂O) by AOB provided ammonium is present (Colliver and Stephenson, 2000, Kampschreur, 2009). High nitrite concentrations during denitrification leads to a lower denitrification rate and accumulation of NO and N₂O (Schulthess et al., 1995). The nitrite concentration is affected by numerous operational parameters. These parameters are (Kampschreur, 2009):

- short SRT;
- toxic compounds (like sulphide)
- low/(high) temperatures
- high salinity
- sudden increase in ammonium load

COD/N

The COD/N ratio only affects the formation of N₂O during denitrification. A limited availability of biodegradable organic carbon is reported to increase the emission of N₂O (Kampschreur, 2009). One study showed the impact of various COD/N ratios (1.5; 2.5; 3.5 and 4.5); at the lowest COD/N ratio up to 10% of the nitrogen load was emitted as N₂O (Hanaki, 1992).

RAPIDLY CHANGING PROCESS CONDITIONS

High concentrations of some of the above process parameters such as O₂ or NO₂-N can be the result of rapidly changing process conditions. These changed process conditions can be caused by changes in environmental conditions or at the transition between anoxic and aerobic zones. Under these conditions it might be possible that the metabolism of bacteria needs time to respond to the changes in environmental conditions, resulting in substantial peak emissions of N₂O. Transient responses of activated sludge upon substrate availability were shown to be in the range of minutes (Vanrolleghem et al., 2004).

Bacterial populations subjected to continuously changing conditions can reduce their N₂O emission by adaptation; *Alcaligenes faecalis* was observed to reduce N₂O emission from 86% to 28% of nitrite converted after ten cycles of dynamic conditions (Schalk-Otte et al., 2000). Similar adaptation behaviour by mixed cultures was observed for N₂O emission upon exposure

to toxic concentrations of formaldehyde (Garrido et al., 1998) and for N₂O emission during start-up of denitrification in a biofilm airlift reactor (van Benthum et al., 1998; Kampschreur, 2009).

2.4 EMISSION OF N₂O

From the previous paragraphs it becomes clear that N₂O can be formed during the nitrification or denitrification stage. This however does not mean that N₂O is emitted directly. This depends on the location where N₂O is formed and the solubility of N₂O. Nitrous oxide formed in anoxic zones can not be stripped there; on the other hand N₂O that is formed in aerobic zones (and not stripped immediately) can be converted in anoxic zones. The solubility of N₂O is relatively high (Henry coefficient is 24 mM/atm, while the Henry coefficient for oxygen is 1.3 mM/atm (Dean, 1992), and stripping is therefore not very fast. Dissolved N₂O in effluent can therefore lead to emission from receiving rivers and estuaries. Sommer et al. (1998) stated that during a winter period the N₂O dissolved in the effluent leaving a specific WWTP was five times higher than the N₂O emitted by air stripping due to the high solubility.

2.5 LOCATIONS CH₄ EMISSION AT WWTP

In contradiction to the formation processes of N₂O, the formation processes of CH₄ are very well known. However little information is available on the amount of CH₄ that is emitted from a domestic WWTP. Up till now one study performed by Czepiel et al. (1993) measured the emission of CH₄ from WWTPs.

Methane can only be formed under anaerobic conditions. In this respect emission of CH₄ can be expected from:

- sewerage systems
- influent works (formation in sewerage system, but emitted here);
- anaerobic / anoxic tanks as part of activated sludge systems;
- sludge digestion and handling;

Methane that is emitted from the influent works is formed in the sewer system. Little information is known about the amount of CH₄ formed in sewers. This is in contrast with H₂S. In most cases the influent headworks is totally covered and emission of CH₄ occurs after the air treatment. Due to the anaerobic conditions in anaerobic and anoxic tanks CH₄ might be formed. However, in the presence of aerobic zones methanogens will not survive.

At WWTPs that are equipped with an anaerobic sludge digester, CH₄ can be emitted from different locations related to the digester. Methane can be emitted during the different processes of biogas combustion (leakages, incomplete combustion) or during storage of digested sludge. At WWTPs without sludge digesters CH₄ formation is possible if the excess sludge is stored.

The amount of CH₄ emitted from these storage facilities will depend on the sludge retention time applied in the activated sludge system, the temperature and the level of dissolved CH₄, which in turn depends on the type of transport system prior to the WWTP.

Despite the presence of oxygen in the aeration tanks CH₄ can be emitted from these tanks. This is most likely CH₄ that has been formed earlier in the sewer or originates from rejection water from sludge handling facilities. The presence of methanogens in activated sludge has

been proven by different authors (Lens et al., 1995, Gray et al., 2002). In these studies it was proven that the contribution of the CH₄ production by methanogens was very limited. Gray et al. (2002) reported 0.01 – 0.02% of the amount of carbon removed.

2.6 EMISSION FACTORS

The United Nations Framework Convention on Climate Change (UNFCCC) is the globally recognised basis for collective action on the reduction of anthropogenic greenhouse gas emissions (UNFCCC, 2007). One of the key obligations for signatory countries under the UNFCCC is the compilation of an annual national greenhouse gas (GHG) inventory, covering four general sectors (energy; industrial processes; agriculture, forestry and other land use; and waste). Emissions of CH₄ and N₂O from wastewater treatment and discharge are reported under the waste sector (IPCC, 2006b). However, GHG emissions are not usually measured directly, but instead are estimated through the application of formulas that link emissions to data on generally reported parameters (Foley and Lant, 2009).

2.6.1 NITROUS OXIDE

The revised 1996 IPCC Guidelines for National Greenhouse Gas Inventories (IPCC, 1997) estimation methodology for N₂O emissions from wastewater handling assumed minimal nitrogen removal occurs during treatment, and hence all influent nitrogen is discharged into rivers and/or estuaries, where it is mineralised, nitrified and denitrified under natural environmental processes. During these transformations, some of the discharged nitrogen will be emitted to the atmosphere as N₂O, at a default factor of 0.01 kgN₂O-N/kgN_{discharged} (uncertainty range: 0.002 – 0.12) (IPCC, 1997). The 2006 IPCC Guidelines for National Greenhouse Inventories subsequently revised this default emission factor to 0.005 kgN₂O-N/kgN discharged (uncertainty range: 0.0005 – 0.25). The assumption that minimal nitrogen removal occurs in wastewater treatment plants (WWTPs) is incorrect for many countries. Recognising this, the 2006 IPCC Guidelines also updated the N₂O estimation methodology to include direct emissions from WWTPs with “controlled nitrification and denitrification steps” (IPCC, 2006a). The proposed default emission factor was 0.0032 kgN₂O·person⁻¹·yr⁻¹ (uncertainty range: of 0.002 – 0.008), based on one full-scale study by Czepiel et al (1995) on a basic secondary treatment plant (without nitrogen removal) in New Hampshire, USA. Assuming a wastewater nitrogen loading of 16 g·person⁻¹·d⁻¹ for developed countries (i.e. high protein intake) (Tchobanoglous et al., 2003; IPCC, 2006a; DCC, 2008b), this equates to approximately 0.00035 kg N₂O-N/kg N. (Paragraph from Foley and Lant, 2009).

The above mentioned IPCC guidelines are used by most countries to estimate the emission of N₂O from domestic WWTP for their national inventory reports (NIR). Andrews et al (2009) examined ten country-specific NIRs, six countries used the IPCC default procedures. The other four can be summarised as follows (Andrews et al, 2009):

- **Denmark** uses a country specific emission factor of 10.8 gN₂O·capita⁻¹·y⁻¹, with a quoted uncertainty of ±30%, which is just under 0.0024 kgN₂O-N/kg N load on secondary treatment, assuming the contribution to load on secondary treatment is 8 gN·person⁻¹·day⁻¹.
- **Japan** uses a country specific emission factor of 160 mgN₂O-N/m³ flow to secondary treatment plus 0.6 mgN₂O-N/m³ from the sludge which, using reasonable assumptions is equivalent to an emission factor of 0.004 kgN₂O-N/kg N load on secondary treatment, with an uncertainty of 146%.
- The **USA** uses the Czepiel et al. (1995) value of 3.2 gN₂O·capita⁻¹·y⁻¹ (0.0007 kg N₂O-N/kg N load on secondary treatment) for plants that only nitrify and 7 g N₂O·capita⁻¹·y⁻¹ (0.0015 kgN₂O-N/kg N load on secondary treatment) for plants that also denitrify.

- The UK (UKWIR Carbon Accounting Workbook (CAW)) uses a country specific emission factor of 0.002 kgN₂O-N emitted/kg N in sewage works load to estimate N₂O emissions from wastewater treatment. It has a high level of uncertainty, in the range of 30 – 300%.

2.6.2 METHANE (CH₄)

The IPCC (IPCC, 2006a) prescribes the following generalised approach for estimating CH₄ emissions from wastewater treatment systems:

$$\text{CH}_4 \text{ Emissions}_w (\text{kg}) = \Delta \text{COD}_w \times EF \times \text{MCF}_w - R_{\text{CH}_4} \quad (1)$$

Similarity, for CH₄ emissions from sludge treatment systems:

$$\text{CH}_4 \text{ Emissions}_{sl} (\text{kg}) = \Delta \text{COD}_{sl} \times EF \times \text{MCF}_{sl} - R_{\text{CH}_4} \quad (2)$$

where

$\Delta \text{COD}_w, \Delta \text{COD}_{sl}$ = Mass of chemical oxygen demand (COD) consumed / removed over the wastewater and sludge treatment processes, respectively (kg), determined by a simplified COD mass balance.

EF = Maximum CH₄ production / emission factor (0.25 kg CH₄ per kg COD).

$\text{MCF}_w, \text{MCF}_{sl}$ = CH₄ correction factor for the type of process employed for wastewater treatment and sludge treatment, respectively (see Table 2)

R_{CH_4} = Mass of CH₄ captured for combustion and/or flaring on the plant, or transfer out of the plant (kg).

In essence, this approach is a reconciliation of the estimated mass of CH₄ produced in the treatment process (i.e. $\Delta \text{COD} \cdot EF \cdot \text{MCF}$), with the measured mass of CH₄ captured in the associated biogas system (i.e. R_{CH_4}). Any difference in these figures is assumed to be a loss of CH₄ to the atmosphere. For treatment systems that are uncovered, R_{CH_4} is zero. Notwithstanding the practical imprecision of measuring COD, biogas flow rates and biogas composition, it is clear that the accuracy of this estimation methodology is dependent upon the factor MCF.

TABLE 2 DEFAULT CH₄ CORRECTION FACTORS FOR DOMESTIC WASTEWATER (IPCC, 2006A; AFTER TABLE 6.3)

Type of treatment	Comments	MCF	Range	CH ₄ production ($EF \times MCF$)
Centralised aerobic treatment plant	Well managed	0.0	0.0 – 0.1	0.00 kgCH ₄ ·kg COD ⁻¹
	Over-loaded	0.3	0.2 – 0.4	0.08 kgCH ₄ ·kg COD ⁻¹
Anaerobic digester or reactor	Does not include CH ₄ recovery	0.8	0.8 – 1.0	0.20 kgCH ₄ ·kg COD ⁻¹
Shallow anaerobic lagoon	Depth < 2m	0.2	0.0 – 0.3	0.05 kgCH ₄ ·kg COD ⁻¹
Deep anaerobic lagoon	Depth > 2m	0.8	0.8 – 1.0	0.20 kgCH ₄ ·kg COD ⁻¹

(Information from Foley and Lant, 2009)

The above mentioned IPCC guidelines are used by most countries to estimate the emission of CH₄ from domestic WWTP for their national inventory reports (NIR). However, these guidelines neglect the emission of CH₄ that has been formed in the sewer and is stripped at the WWTP as the IPCC states that “*wastewater in closed underground sewers is not believed to be a significant source of methane*”. (IPCC, 2006a,b).

Andrews et al (2009) examined 12 country-specific NIRs, eight countries used the IPCC default procedures (the 2006 Guidelines have not been applied yet for the preparation of NIRs) or IPCC procedures with the development of country-specific EFs. Sweden indicated that all secondary treatment was aerobic with no CH₄ emissions, and all sludge treatment had energy recovery and was dealt with under the energy sector. The UK methodology amounts to 20.8 kg CH₄/tonne raw DS for emissions from sewage treatment and onsite sludge treatment including digestion. Making reasonable assumptions this amounts to 0.6 kg CH₄·capita⁻¹·year⁻¹. Canada uses an EF, based on IPCC (1997) of 4.015 kg·capita⁻¹·year⁻¹. Japan uses country specific values of 528.7 mgCH₄/m³ of influent from wastewater treatment and 348.0 mgCH₄/m³ from sludge treatment. This amounts to approximately 0.056 kgCH₄·capita⁻¹·year⁻¹. An overview of all applied emission factors is presented in Appendix 1.

3

LOCAL REGULATION AROUND GHG EMISSION FROM WWTP

3.1 AUSTRALIA

3.1.1 REPORTING REGULATIONS

In Australia, the National Greenhouse and Energy Reporting System (NGERS) provides a single national framework for reporting greenhouse gas emissions, energy use and energy production by corporations. It is also intended to underpin a Federal Government emissions trading scheme at some stage in the future.

Under NGERS, any “facility” using more than 100 terajoules (TJ) of energy, or emitting more than 25,000 tonnes of Scope 1 and Scope 2 carbon dioxide equivalents (CO₂-e) per annum is required to report to the Federal Department of Climate Change and Energy Efficiency (DCCEE)⁸. Similarly, any “corporation” using more than 350 TJ of energy or emitting more than 87,500 tonnes CO₂-e in 2009/10 is required to report to the DCCEE. These corporate thresholds will decrease to 200 TJ and 50,000 tonnes CO₂-e for the 2010/11 reporting year. Changes in thresholds beyond 2011/12 are presently unknown.

The following Act and Regulations define the legislative requirements for greenhouse gas reporting within Australia:

- *The National Greenhouse and Energy Reporting Act 2007*, including amendments; and
- *The National Greenhouse and Energy Reporting Regulations 2008*, including amendments;
- The Regulations provide detailed requirements for reporting under the Act, including definitions of operational control, facilities, the requirements for registration and the types of greenhouse gas emissions and energy consumption/production that have to be reported.

Further, the following documents provide methods and criteria for calculating greenhouse gas emissions and energy data under the *National Greenhouse and Energy Reporting Act 2007*

- The National Greenhouse and Energy Reporting (Measurement) Determination (2008);
- The *National Greenhouse and Energy Reporting (Measurement) Determination 2008* has been amended by the *National Greenhouse and Energy Reporting (Measurement) Amendment Determination 2009* (the Amendment Determination). The Amendment Determination is in general applicable to the 2009-10 reporting year; and
- *The National Greenhouse and Energy Reporting Technical Guidelines 2009* (or later revisions, if available).

⁸ Under NGERS, reporting of Scope 3 emissions is voluntary.

These documents (and associated links) are publicly available via the DCCCE website (and associated links): www.climatechange.gov.au

The details of the NGERS are presented in Appendix 5.

3.2 FRANCE

In France there are no specific regulations concerning GHG emissions for the water sector. Nevertheless, all industries/companies with more than 50 employees must perform a carbon footprint assessment (new law July 2010). In the case of wastewater treatment plants, the French EPA (ADEME) recommends taking direct emissions of CH₄ and N₂O into account.

3.3 UNITED STATES OF AMERICA

In the United States of America (USA), environmental regulations may be enacted at the federal, state or local level. States or local authorities may issue regulations that are more restrictive than the federal government; however, those regulations apply only to those respective regions. The following summary of greenhouse gas legislation and regulations includes enacted regulations that affect, or may affect, publicly-owned treatment works (POTW). The summary includes current federal regulation and regulations by one state, California. No other states or coalitions of states have regulations on greenhouse gas (GHG) emissions that directly affect publicly owned wastewater treatment facilities. Proposed federal and state regulations are not discussed here.

3.3.1 REGULATIONS THAT (MAY) AFFECT PUBLICLY-OWNED TREATMENT WORKS (POTW)

On April 2, 2007, the Supreme Court of the United States, ruling in *Massachusetts vs. EPA*, 2007, found that GHGs, including carbon dioxide, are air pollutants covered by the federal Clean Air Act (CAA). On December 7, 2009, the USEPA Administrator signed two distinct findings regarding GHGs under the CAA a) that six, key, well-mixed GHGs threaten the public health and welfare of current and future generations and b) that the combined emissions of these well-mixed GHGs from new motor vehicles contribute to greenhouse gas pollution that threatens public health and welfare.

A series of clarifying actions then followed that culminated in the clarification of terms such as “regulated New Source Review pollutant” and “subject to regulation” and when a regulatory requirement to control emissions of a pollutant actually “takes effect.” The action also confirmed that the next regulation dealing with GHGs was the light duty vehicle rule (LDVR) whose requirements would not “take effect” any earlier than January 2, 2011.

The USEPA determined that one effect of the LDVR would be significant impacts on state permitting authorities for both Prevention of Significant Deterioration (PSD) and Title V permitting programs if these programs were left at current criteria pollutant trigger thresholds. PSD is that portion of the federal New Source Review program that regulates pollutants in regions of the country that have already “attained” the national ambient air quality standard (NAAQS). PSD permitting was impacted since no national ambient air quality standard (NAAQS) for GHGs existed or will exist.

Title V was impacted because PSD is implemented through the Title V operating permit program, and GHGs can, by themselves, cause a facility to be a major source subject to Title V.

If the traditional 100/250 tons (USA) per year criteria pollutant trigger levels remained in place on January 2, 2011, they would lead to dramatic increases in the number of required permits—tens of thousands of PSD permits and millions of Title V permits—overwhelming permitting authorities unless the USEPA acted to “tailor” the GHG thresholds upwards.

On May 13, 2010, the USEPA issued a final rule that “tailors” GHG emissions from stationary sources under the CAA permitting programs and defines when permits under the PSD and Title V Operating Permit programs are required to address GHGs for new and existing industrial facilities. Facilities responsible for nearly 70 percent of the national GHG emissions from stationary sources will be subject to permitting requirements under this rule. The rule establishes a schedule that will initially focus CAA permitting programs on the largest sources that would have had to get PSD and Title V permits in any event, for their criteria pollutant emissions (“anyway sources”). The rule then expands to cover the largest sources of GHG emissions, at specific trigger levels, not previously covered by the CAA. Finally, it describes USEPA plans for additional, later steps in this process.

Compliance with PSD and Title V would be required by facilities that directly emit $\geq 75,000$ or $\geq 100,000$ tons of carbon dioxide equivalent (CO₂e) GHG emissions, depending on whether the emissions are from a new facility or from a change within an existing facility. In 2011 USEPA published a final rule, that for three years exempts biogenic CO₂ emissions from the above thresholds for PSD and Title V permitting purposes, while USEPA studies the issue further. Only biogenic CO₂ and not N₂O or CH₄ are covered by the temporary exemption.

3.3.2 FEDERAL MANDATORY REPORTING OF GREENHOUSE GASES RULE

In 2009, USEPA issued a rule that requires mandatory reporting of GHG emissions from large sources in the United States. The rule requires the collection and submission of accurate and comprehensive emissions data to inform future policy decisions. Suppliers of fossil fuels or industrial greenhouse gases, manufacturers of vehicles and engines, and facilities that emit 25,000 metric tons or more per year of GHG emissions are required to submit annual reports to USEPA.

The key reporting requirement applies to stationary combustion sources above 25,000 Million Tons (USA) CO₂e/yr. Stationary fuel combustion sources include, but are not limited to, boilers, simple and combined-cycle combustion turbines, engines, incinerators, and process heaters. The only wastewater treatment process emissions to be reported in this rule are those from onsite wastewater treatment located at industrial facilities. POTW process emissions have not been included in the rule because USEPA does not expect emissions from POTWs to exceed the applicable thresholds. The calculation of total emissions for the purposes of determining whether a facility exceeds the threshold does not include biogenic CO₂ emissions (e.g., those resulting from combustion of biofuels, including the elimination of biosolids from the calculation of combustion emissions).

However, to the extent POTWs utilize fossil fuel in their combustion facilities; these would be included in the threshold applicability determination. If the 25,000 MT (USA) CO₂e/yr threshold is exceeded based upon fossil emissions, then biogenic emissions would also need to be reported. There is no federal reporting requirement for combustion emissions below 25,000 MT CO₂e/yr for a facility.

Emissions from stationary combustion units are determined using equations in the final rule, with the quantity of fuel combusted and emissions factors as the equation parameters. USEPA has clarified that utilities do not need to calculate or report emissions from combustion of

biosolids. One key question for wastewater utilities is whether or not they have a facility whose collective combustion unit heat input rated capacity exceeds 30 mmBtu/hr. If this is so, they are required to calculate emissions for that facility to determine if they meet the threshold for reporting. For biogas combustion, only the CH₄ and N₂O portion of the emissions need be included, and for fossil fuels, all of the six well-mixed GHG constituents for which there are emission factors, need be included.

3.3.3 CALIFORNIA'S AB 32 THE GLOBAL WARMING SOLUTIONS ACT OF 2006

In 2006, the California Legislature enacted AB 32, the Global Warming Solutions Act of 2006. This Act establishes the first comprehensive GHG regulatory program in the USA, and commits California to achieving significant GHG emissions reductions by 2020. The Act will regulate all public and private entities that emit GHGs (defined as carbon dioxide, CH₄, N₂O, hydrofluorocarbons, perfluorocarbons, and sulfur hexafluoride) to achieve a state-wide emissions limit equivalent to the GHG emissions level back in 1990. This legislation directs the California Air Resources Board (CARB) to develop and implement a comprehensive program to achieve this goal by 2020. The reduction measures to meet the 2020 target are required to be largely adopted by the start of 2011. The CARB is required to coordinate with other state agencies to develop the AB 32 program. (The CARB is also charged with achieving a longer-range goal of complying with an earlier Executive Order of the Governor that sets state GHG emission target of 80% below 1990 levels by 2050.)

There are a number of elements required by AB 32, including reporting under a California Mandatory Reporting Program, and reductions prescribed in a comprehensive Scoping Plan. By January 1, 2008, CARB required annual reporting and verification of GHG emissions, beginning with the sources or categories of sources that contribute most to statewide emissions. The CARB's Mandatory Reporting Program uses reporting thresholds similar to the federal program, however, biogenic emissions do count in determining if the reporting threshold is exceeded, although they are reported separately from fossil emissions. Therefore, more POTWs will report in California compared to the federal program.

In the Scoping Plan, CARB has five regulatory measures directed at the water/ wastewater sector including increasing water use efficiency, increasing water recycling, reduction in the magnitude and intensity of California's water systems, increased usage of urban runoff, increased renewable energy production from water systems and a public goods charge on water meters to pay for most of the above activities.

3.4 THE NETHERLANDS

Treatment of wastewater in the Netherlands is delegated to the Waterboard Authorities. These Waterboard Authorities are obliged to report the emission of greenhouse gases from wastewater treatment plants with a capacity larger than 136,360 p.e.⁹, or from wastewater treatment plants that treat more than 50 tonnes wet sludge (primary and secondary) per day (IPCC) from other WWTPs. The emission of N₂O and CH₄ are in these cases based on different emission factors as used in national inventory reports (NIR¹⁰). For N₂O an emission factor of

⁹ Based on 136 g total oxygen demand

¹⁰ Required under the Kyoto protocol. Emission factors used in the Netherlands are for N₂O 1 % of NKj influent and 0.007 kg CH₄ / kg COD_{influent} for WWTP without sludge digestion and 0.0085 kg CH₄ / kg COD_{influent} for WWTP with sludge digestion.

0.07% of the total nitrogen load is used. For CH₄ it is assumed that the majority of the CH₄ is emitted from sites that store digested sludge. In this case an emission factor of 18 g CH₄·p.e.⁻¹·year⁻¹ is used.

Since April 2010 the Dutch Waterboards signed a “Dutch Climate Agreement” with the government. Part of this agreement is that the waterboards committed themselves to reduce the emission of non CO₂ GHG with 30% (equal to approximately 200 ktonnes CO₂e from 1990 to 2020). This number can be adjusted based on the outcomes of current research (performed from September 2010 – September 2011) on the emission of N₂O and CH₄ from WWTPs.

4

BACKGROUND AND OBJECTIVES RESEARCH

4.1 AUSTRALIA

4.1.1 EMISSION OF N₂O

In May 2008, WSAA commissioned The University of Queensland (UQ) to undertake field-based research to improve the level of certainty in the estimation methodologies for direct CH₄ and N₂O emissions from wastewater systems.

One of the aims of the study was to improve the level of certainty in the estimation methodologies for direct N₂O emissions from wastewater systems, as calculated in the NGERS Technical Guidelines. The default emission factor for wastewater treatment under NGERS (NGERS Technical Guidelines, 2009) is equivalent to 0.01 kg N₂O per kg nitrogen denitrified.

Specifically, the study addressed a knowledge gap identified in the earlier WSAA Literature Review (Foley and Lant, 2008): There is a high level of uncertainty in the magnitude and variability of N₂O emissions from biological nutrient removal (BNR) processes, under different physical configurations and process conditions.

The purpose of the research element was to determine the N₂O generation and emission rates in full-scale treatment plants, of varying physical configuration and under different process conditions. The study did not address the uncertainty in emissions from wastewater discharges to different receiving environments.

4.1.2 EMISSION OF CH₄

With a global warming potential 25 times that of carbon dioxide, CH₄ is a potent GHG. Significant CH₄ production and emission from wastewater collection systems was not revealed until very recently. Indeed, the lack of data has led the IPCC to conclude, in its 2006 Guidelines (IPCC, 2006a,b), that “wastewater in closed underground sewers is not believed to be a significant source of methane”. As such, CH₄ in sewers is currently not considered in the accounting of greenhouse gas emissions from wastewater systems. However, the plentiful supply of readily biodegradable carbon, the high (>20°C) temperatures in certain locations and the presence of anaerobic biofilms, particularly in fully surcharged rising mains, suggest that there is potential for the formation of CH₄ in the raw sewage, and this is presently unaccounted in national GHG inventories (Foley and Lant, 2008).

For this reason several investigations were performed in Australia on the emission of CH₄ from rising mains. This research included measurements in the gas and liquid phase in the field and at laboratory scale. Based on the measurements two types of models were developed, one of which will be presented in this report. Further research was performed to study the impact of trade waste on the emission of CH₄ from rising mains. Finally research was performed to limit the formation of CH₄ in rising mains.

The methodology of the measurements in the gas and liquid phase in the field and at laboratory scale is discussed in section 5.5. The methodology of the other research topics is presented together with the results in section 6.5.

4.2 UNITED STATES OF AMERICA

4.2.1 BACKGROUND

The push to achieve greater nutrient removal from wastewater treatment plant effluents has resulted in the development of a wide range of innovative biological nitrogen removal (BNR) processes. However, BNR strategies could be a potential contributor to atmospheric N₂O and NO depending upon the reactor configurations and operating conditions. In the future, as BNR is implemented at wastewater treatment plants, the flux of these gases to the atmosphere could increase.

Based on recent field-scale measurements, engineered BNR facilities, while effective to varying degrees in reducing *aqueous* nitrogen pollution, could emit up to 7% of the influent nitrogen load as *gaseous* N₂O and NO (Kampschreur et al., 2008b).

The WERF research project represents one of the first attempts at characterizing nitrogenous GHG emissions from wastewater treatment plants, and at developing a methodology for collection of full scale plant data from a range of nutrient removal facilities in the United States. Building on previous work by the project team, this information will be integrated into a mechanistic activated sludge process model, which will be refined through this project by the addition of autotrophic pathways for N₂O and NO emission. The refined mechanistic model will allow the industry to codify the results of this research, and develop a tool that will aid in the prediction and therefore mitigation of N₂O, NO and NO₂-N emissions from WWTPs utilizing a range of wastewater treatment processes. Ultimately, this would allow the wastewater sector to engineer strategies for wastewater treatment that minimize gaseous nitrogen oxide emissions.

Although, from a fundamental perspective, N₂O and NO are known intermediates in heterotrophic denitrification (Knowles, 1982, Zumft, 1997) and autotrophic nitrification and denitrification (Anderson and Levine, 1986, Anderson et al., 1993, Kester et al., 1997, Ritchie and Nicholas, 1972, Stuenkel et al., 1992), the net contribution of BNR processes to N₂O emissions from wastewater treatment has only recently been explicitly acknowledged (USEPA, 2009). Based on the latest USEPA report on sources and sinks of N₂O from wastewater treatment operations, denitrification in anoxic zones is implicated as the dominant source of N₂O from BNR activated sludge reactors (USEPA, 2009).

However, nitrification could also play a role in N₂O generation and emission from WWTPs, especially under cycling between anoxic and aerobic conditions (Kampschreur et al., 2008b), as is common in BNR reactors. Therefore, N₂O fluxes from aerobic zones of WWTPs also need to be included in N₂O emissions inventories.

At this point, while there is considerable debate and interest on the 'potential' of constituent BNR processes for N₂O generation and emission, there are few reports that systematically quantify such emissions from full-scale BNR operations (Czepiel et al., 1995, Kampschreur

et al., 2008b, Kimochi et al., 1998, Sommer et al., 1998, Sümer et al., 1995, Wicht and Beier, 1995). Of these, only one has been conducted in the USA and focuses on a single non-BNR WWTP in New Hampshire (Czepiel et al., 1995).

Other full-scale studies have been conducted in Europe and have employed different methods for measuring N₂O emissions including the use of grab samples (Czepiel et al., 1995, Kampschreur et al., 2008b, Sommer et al., 1998, Sümer et al., 1995, Wicht and Beier, 1995) or online measurements (Kampschreur et al., 2008b, Kimochi et al., 1998). Given the broad diversity of BNR and non-BNR configurations that exist in the USA, a more detailed N₂O emissions database of WWTPs was needed, specifically obtained using a consistent protocol. Despite recognition of the possible role of BNR processes in N₂O emission, a measured database of N₂O emissions from these processes at the national scale does not currently exist. The WERF project focused on the quantification of N₂O emissions at twelve wastewater treatment plants (WWTPs) across the United States using a newly developed, USEPA-reviewed protocol. A subsequent focus was to determine the mechanisms and triggers of these emissions.

The principal motivation of conducting such a detailed monitoring campaign is the limited data currently used to “estimate” the N₂O emissions of WWTPs. The current method is based on emission factor values of 3.2 g N₂O/population equivalents/year from non-BNR operations and 7.0 g N₂O/population equivalent/year for BNR operations (Czepiel et al., 1995, USEPA, 2009). Both emission factors are based on a limited data set and may not be broadly representative.

4.2.2 EMISSION FACTORS UNITED STATES OF AMERICA

From a regulatory and policy perspective, organizations such as the USEPA are now beginning to explicitly include the contribution of BNR processes such as denitrification on the overall N₂O emission inventory from WWTPs (USEPA, 2009). A common approach to estimating the N₂O inventory of wastewater treatment processes is by using a ‘single’ emission factor with a value of 3.2 g N₂O/population equivalent/year and 7.0 g N₂O/population equivalent/year for non-BNR and BNR processes (USEPA, 2009). In these calculations, one population equivalent is defined as 100 gallons of wastewater discharged per capita per day. Conceptually, given that the inputs to a wastewater treatment plant and correspondingly the activity of the activated sludge bacteria are highly variable (Grady et al., 1999), it can be expected that there would be some degree of diurnal variability in N₂O emissions. Furthermore, owing to the fact that activated sludge bacteria have varying activities in different zones of the bioreactors (Grady et al., 1999), a certain degree of spatial variability in N₂O emissions is also expected from anoxic, anaerobic and even aerobic zones.

However, such spatial and diurnal variability in N₂O emissions is not considered in the simple emission factor approach adopted by the USEPA and IPCC for estimating the N₂O inventory of BNR and non BNR processes. Furthermore, by approximating the N₂O emissions from wastewater treatment processes using single emission factors across the board, the ability of certain operating conditions to selectively promote or minimize N₂O emissions is not understood.

Furthermore, not all emission factors are equivalent, owing to different conventions for normalizing, including wastewater flow rate, wastewater influent total nitrogen load or nitrogen load removed.

4.2.3 OBJECTIVES

The overall goal of this study was to quantify the emission of N₂O from WWTPs across the nation operated under different process conditions and configurations. The specific objectives were to:

- Develop a database of N₂O emissions fluxes from different activated sludge process configurations using a standard protocol.
- Identify key factors that are correlated with N₂O emission from activated sludge
- Determine the spatial and temporal variability in N₂O emissions from WWTPs (both BNR and non BNR) across the nation and examine the validity of the single emission factor approach in estimating the inventory of N₂O from WWTPs.

4.3 THE NETHERLANDS

BACKGROUND

In the Netherlands the Dutch Waterboards aim to operate their wastewater treatment plants in a sustainable way. This implies that they try to minimize their energy consumption and (non CO₂) greenhouse gas emission. However, until 2007 very limited data were available on the emission of N₂O and CH₄ from Dutch WWTPs. For this reason there was a need to investigate the emission of GHG from Dutch WWTPs.

OBJECTIVE

The objective of this research was to determine the level of N₂O and CH₄ emission from Dutch WWTPs and to understand the accuracy of the existing emission factors. In order to reach this objective the following research questions were raised:

- What is the emission of N₂O and CH₄ from a WWTP representative for the Dutch situation?
- Where do the emissions of N₂O and CH₄ originate from?
- Are the current emission factors for N₂O and CH₄ useful to estimate the total GHG emission from a Dutch WWTP?
- Is additional research required to identify reliable emission factors for N₂O and CH₄?

5

METHODOLOGY

5.1 NITROUS OXIDE MEASUREMENTS AUSTRALIA

5.1.1 FIELD SAMPLING SITES

The fieldwork program was conducted at seven full-scale BNR wastewater treatment plants in Australia. These plants were chosen to provide a range of plant sizes, process configurations, effluent qualities and climatic conditions. Their basic features are listed in Table 3. The widest possible range of WWTP types (in Australia) was selected with the expectation that differences between plant design and process conditions might help elucidate the factors influencing N₂O production.

TABLE 3 WASTEWATER TREATMENT PLANT SITES

Type of process	Sludge load ¹⁾ (kgTKN _{in} · kg d.w. ⁻¹ · d ⁻¹)	Temperature (°C)	SRT (appr.) (d)	Effluent TN ¹ (mg/l)	Process description
1. Ox.Ditch (4 rounds)	0.043 0.042 0.03 0.042	23	13	3	Inlet works, anaerobic contact tank, 2 x extended aeration oxidation ditches (in parallel) with diffused aeration, secondary sedimentation; mechanical sludge thickening and dewatering.
2. Johannesburg (3 rounds)	0.012 0.01 0.013	21	20	5	Inlet works, 2 x extended aeration Johannesburg bioreactors (in parallel) with submerged aspirating OKI™ aerators, secondary sedimentation, sludge thickening, aerobic digestion, mechanical and solar dewatering.
3. SBR (3 rounds)	0.021 0.017 0.021	20	16	15	Inlet works, primary sedimentation, sequencing batch reactor (4 compartments) with diffused aeration and bio-selector zone, sludge thickening, anaerobic digestion and mechanical dewatering.
4. MLE-1 (1 round)	0.031	21	13	12	Inlet works, primary sedimentation, 11 x covered Modified Ludzack Ettinger (MLE) bioreactors (in parallel) with diffused aeration, secondary sedimentation, sludge thickening, mechanical dewatering and lime stabilisation.
5. MLE-2 (4 rounds)	0.06 0.065 0.072 0.07	20	8	11	Inlet works, primary sedimentation, 2 x Integrated Fixed Film Activated Sludge bioreactors + 1 x MLE bioreactor (in parallel) with diffused aeration, secondary sedimentation, sludge thickening and anaerobic digestion.
6. MLE-3 (3 rounds)	0.01 0.01 0.012	18	15	13	Anaerobic lagoon, 1 x MLE bioreactor with diffused aeration, secondary sedimentation, maturation lagoons, and sludge wasting to an aerated facultative lagoon.
7. A ² /O (2 rounds)	0.009 0.01	21	14	3	Inlet works, 2 x parallel trains: 1) Primary sedimentation, 4-stage Bardenpho bioreactor with diffused aeration, secondary sedimentation; 2) Pre-fermenter, four-stage bioreactor (similar to A ² /O configuration) with diffused aeration and supplemental COD dosing by primary sludge from Train 1, secondary sedimentation, tertiary filtration, sludge thickening, aerobic digestion, sludge lagoon and mechanical dewatering.

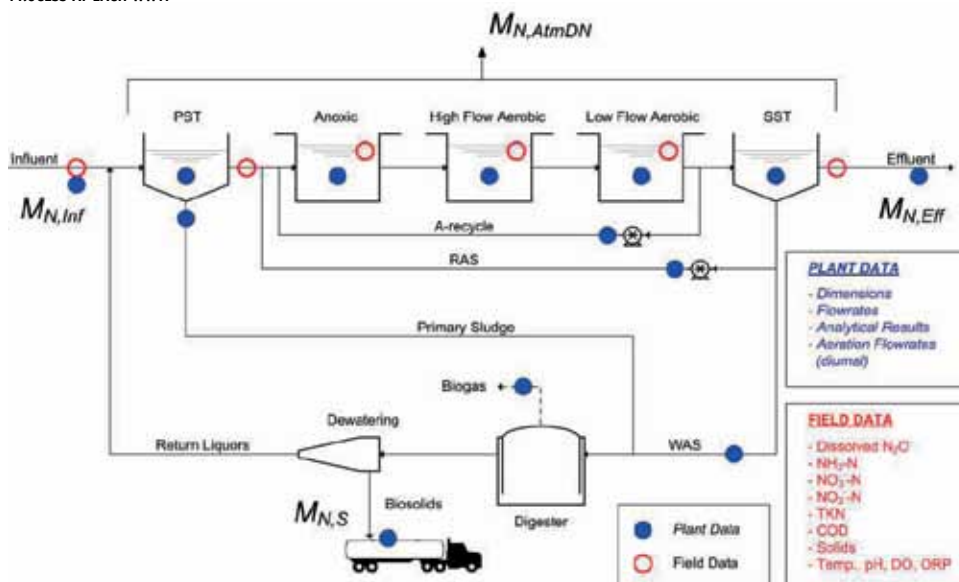
5.1.2 SAMPLE COLLECTION AND ANALYSIS

For each WWTP, it was intended to conduct four sampling rounds (2 – 4 h duration each, morning and afternoon on two consecutive days). Due to a combination of circumstances, this was not possible at all plants. However, 20 of the intended 28 sampling rounds were completed over a five month timeframe in the Australian winter/spring of 2008.

For each sample round, data was collected to enable the construction of chemical oxygen demand (COD), total nitrogen (TN) and N₂O-N mass balances over the entire WWTP. The sampling locations and types of data collected are illustrated in Figure 5. Field data collection consisted of a combination of:

- Wastewater grab samples;
- Measurement of process conditions, namely temperature, pH, dissolved oxygen and oxidation-reduction potential (ORP), using a portable water quality meter (TPS 90FLMV); and
- Dissolved N₂O concentration measurements using a Clark-type microsensor (N₂O 25 with 70 µm outside tip diameter, Unisense A/S, Aarhus, Denmark), logged via a Pico ammeter to a laptop.

FIGURE 5 TYPICAL DATA (TYPES AND LOCATIONS) COLLECTED FOR THE CONSTRUCTION OF THE COD AND TOTAL NITROGEN MASS BALANCES OVER THE ENTIRE PROCESS AT EACH WWTP



5.1.3 DETERMINATION OF N₂O EMISSIONS

Chemical oxygen demand and Total Nitrogen Mass Balances over Entire WWTP Processes

At each of the seven WWTP sites, total COD and TN mass balances were constructed across the entire process. These mass balances drew upon the analytical data collected in the field (i.e. COD, TKN, -N, NO₂-N, MLSS, MLVSS concentrations), as well as the plant data (i.e. flowrates, reactor volumes, solids capture efficiencies, biosolids tonnages and composition, biogas production and composition) supplied by the WWTP operators (refer to Figure 5). The purpose of this initial mass balance analysis was to:

- Ensure an accurate characterisation of the WWTP operation, such that both COD and TN balances over the WWTP generally achieved greater than 90% closure; and

- To determine the mass of nitrogen denitrified and emitted to the atmosphere, according to Equation 3:

$$M_{N,AtmDN} = M_{N,Inf} - M_{N,Eff} - M_{N,S} \quad (3)$$

Where:

$M_{N,AtmDN}$	= Mass of N denitrified to atmosphere, either as N ₂ or N ₂ O gas (kg·d ⁻¹)
$M_{N,Inf}$	= Mass of nitrogen in influent (kg/d)
$M_{N,Eff}$	= Mass of nitrogen in effluent (kg/d)
$M_{N,S}$	= Mass of nitrogen in wasted solids (kg/d)

Equation 3 assumes that the WWTP is operating at steady-state, with no net accumulation of nitrogen within the biomass inventory. All of the plants investigated had medium to long solids retention times (SRT) (i.e. 8 – 20 d). Hence, the change in biomass inventory is relatively slow, and the assumption of near steady-state conditions should hold for the two day sampling period at each WWTP.

Given the large variation in physical size and treated load of the seven WWTPs surveyed, Equation 3 provided a means of normalising the generation and emissions of N₂O (i.e. as a percentage of the total nitrogen denitrified to atmosphere).

Liquid Phase N₂O Mass Balances over Individual WWTP Zones

The second phase of mass balance analysis examined bulk liquid phase N₂O across the individual zones of each WWTP. For five of the seven sites, the WWTP was divided into five reactor zones for this mass balance analysis (i.e. primary sedimentation tank or anaerobic zone; anoxic zone; highly aerated aerobic zone; less aerated aerobic zone; and secondary sedimentation tank). At WWTP No.6, the plant was divided into seven reactor zones (three anoxic zones, three tapered flow aerobic zones, and secondary sedimentation tank). At WWTP No.3 (SBR), the mass balance was divided across each operational phase (i.e. fill/aerate, settle, decant). The general formulation of the mass balance construction is given in Equation 4:

$$\frac{dM_{N_2O-N,R}}{dt} = \sum M_{N_2O-N,In} - \sum M_{N_2O-N,Out} - Tr_{N_2O-N,R} + G_{N_2O-N,R} \quad (4)$$

Where:

$\frac{dM_{N_2O-N,R}}{dt}$	= change in mass of N ₂ O-N in the reactor zone, over time (kg·d ⁻¹)
$\sum M_{N_2O-N,In}$	= sum of i mass flows of N ₂ O-N into the reactor zone (kg·d ⁻¹)
$\sum M_{N_2O-N,Out}$	= sum of j mass flows of N ₂ O-N out of the reactor zone (kg·d ⁻¹)
$Tr_{N_2O-N,R}$	= mass transfer of N ₂ O-N from the reactor liquid to gas phase (kg·d ⁻¹)
$G_{N_2O-N,R}$	= net generation of N ₂ O-N in the reactor zone (kg·d ⁻¹) (i.e. net result of N ₂ O-N production and consumption due to biological reactions in the reactor)

Similar to Equation 3, it was assumed that the reactor zones operate at near steady-state conditions, and are well-mixed. Equation 4 can then be expanded and re-formulated to solve for:

$$G_{N_2O-N,R} = \sum_j Q_{Out,j} \times [N_2O-N]_R - \sum_i Q_{In,i} \times [N_2O-N]_{In,i} + V_R \times k_L a \times ([N_2O-N]_R - [N_2O-N]_S^*) \quad (5)$$

Where

- $Q_{In,i}, Q_{Out,j}$ = individual flows in and out of the reactor zone (Ml/d)
- $[N_2O-N]_{In,i}$ = concentration of N₂O-N in the incoming streams (mg/l or kg/Ml),
which is generally equal to the N₂O-N concentration in the originating reactor
- $[N_2O-N]_R$ = concentration of N₂O-N in the reactor zone (mg/l or kg/Ml)
- V_R = volume of the reactor zone (Ml)
- $k_L a$ = volumetric mass transfer coefficient (d⁻¹)
- $[N_2O-N]_S^*$ = saturation concentration of N₂O-N in water at atmospheric conditions
= $2.57 \cdot 10^{-4}$ kg/Ml at 20°C (Weiss and Price, 1980)

This calculation was completed for each reactor zone at each WWTP. It was then repeated at each reactor zone for the “best-case” combination of the lower limit values of the 95% confidence intervals of the measured N₂O-N concentrations and estimated mass transfer coefficients (refer to Figure 52). The calculation was then repeated again for the “worst-case” combination of the upper limit values of the 95% confidence intervals. These calculations determined the uncertainty range of net N₂O-N generation (i.e. production minus consumption) in each reactor zone. Negative values of $G_{N_2O-N,R}$ indicate N₂O-N consumption is greater than N₂O-N production in that particular zone.

The net generation and emissions of N₂O-N in each reactor zone were then summed to give the net generation, $G_{N_2O-N,WWTP}$, and emissions, $Tr_{N_2O-N,WWTP}$, of N₂O-N for the whole WWTP:

$$G_{N_2O-N,WWTP} = \sum G_{N_2O-N,R} \quad (6A)$$

$$Tr_{N_2O-N,WWTP} = \sum Tr_{N_2O-N,R} = \sum \{V_R \times k_L a \times ([N_2O-N]_R - [N_2O-N]_S^*)\} \quad (6B)$$

The ratio of N₂O-N mass transfer emissions to net generation was then calculated for each WWTP:

$$\%Tr_{WWTP} = \frac{Tr_{N_2O-N,WWTP}}{G_{N_2O-N,WWTP}} \times 100\% \quad (7)$$

NORMALISATION OF NITROUS OXIDE MASS BALANCE RESULTS

To compare results between different reactor zones, $G_{N_2O-N,R}$, was normalised by dividing by the corresponding total mass of nitrogen denitrified to the atmosphere from the entire WWTP:

$$GF_R = \frac{G_{N_2O-N,R}}{M_{N,AtmDN}} \quad (8A)$$

Where:

GF_R = N₂O-N generation factor for each reactor zone
(kgN₂O-N/kgN_{denitrified})

$M_{N,AtmDN}$ is calculated according to Equation 1.

Similarly, the mass transfer emissions to atmosphere from each reactor zone, $Tr_{N_2O-N,R}$, were normalised by dividing by $M_{N,AtmDN}$:

$$EF_R = \frac{Tr_{N_2O-N,R}}{M_{N,AtmDN}} \quad (8B)$$

Where:

EF_R = N₂O-N emissions factor for each reactor zone
(kgN₂O-N/kgN_{denitrified})

To compare results across sites, $G_{N_2O-N,WWTP}$ for each WWTP was normalised by dividing by its corresponding total mass of nitrogen denitrified to the atmosphere:

$$GF_{WWTP} = \frac{G_{N_2O-N,WWTP}}{M_{N,AtmDN}} \quad (9)$$

Where:

GF_{WWTP} = N₂O-N generation factor for entire WWTP (kgN₂O-N/kgN_{denitrified}).

The determination of the Nitrous Oxide Volumetric Mass Transfer Coefficients is presented in Appendix 6.

5.1.4 QUALITY CONTROL

The electrochemical microsensor was two-point calibrated before and after each field sampling round at ambient temperature, using distilled water (zero point) and a freshly prepared 0.15 mM N₂O solution. Laboratory trials with the N₂O25 microsensor in 0.27 mM N₂O solution showed its maximum measurement error to be $\pm 0.3\%$. The response of the electrochemical microsensor is known to be linear in the range of 0 - 1.2 mM (Andersen et al., 2001). At each sampling location, the microsensor was fully immersed in the reactor and allowed to stabilise and log data at 3 s intervals over a 5 - 10 minute period. The signal data from this sampling period at each location was then statistically analysed to calculate its 95% confidence interval (t-dist, $\alpha = 0.05$). The microsensor was also zero point calibrated with distilled water in the field after every measurement to provide a baseline signal and correct for any drift.

The TPS 90FLMV water quality meter was calibrated before and after the two-day sampling exercise at each WWTP. Grab samples for soluble species were immediately filtered using 0.22 μ m syringe filters, acid-preserved and kept on ice before analysis.

COD was measured by the colorimetric method described in APHA (1995) using commercial

Lovibond tubes in a range of 0 to 150 mgCOD.L⁻¹. The ammonium (NH₄⁺-N), nitrate (-N) and nitrite (NO₂⁻-N) concentrations were analysed using a Lachat QuikChem8000 Flow Injection Analyser (Lachat Instrument, Milwaukee, USA). Total Kjeldahl Nitrogen (TKN), mixed liquor suspended solids (MLSS) and volatile solids (MLVSS) were analysed according to Standard Methods (APHA, 1995).

Physical plant data (e.g. reactor dimensions, plant flowrates, diurnal aeration flowrates) were provided by the WWTP operators for the specific days and times of field sampling. Where possible, the WWTP operators also supplied their own routine process and analytical data that provided a useful cross-check against results from the grab samples collected during the field study.

5.2 NITROUS OXIDE MEASUREMENTS FRANCE

5.2.1 FIELD SAMPLING SITES

Four WWTPs have been monitored, all of them with N removal: two 'classical' BNR activated sludge facilities, one 'plug-flow' process and one Membrane Bioreactor (MBR). The characteristics of these WWTPs are summarized in Table 4.

TABLE 4 CHARACTERISTICS FOUR WWTPS STUDIED IN THE N₂O RESEARCH

WWTP	Water line	Biological treatment
WWTP 1	Pre treatment- Activated Sludge – Secondary Clarifier	Three-zones completely Stirred Tank Reactor. (anaerobic- anoxic- aerobic with sequenced aeration)
WWTP 2	Pre treatment - Activated Sludge - Secondary Clarifier	Plug flow process (anoxic –aerobic)
WWTP 3	Pre treatment - Activated Sludge - Secondary Clarifier	Three-zone completely Stirred Tank Reactor. (anaerobic- anoxic- aerobic with sequenced aeration)
WWTP 4	Pre treatment – Activated Sludge – Membrane Bio Reactor	Activated sludge with sequenced aeration followed by MBR

The configuration of WWTP 1 - 4 are presented in Annex 7.

5.2.2 SAMPLE COLLECTION AND ANALYSIS

The way the samples were taken for N₂O analysis depended on if the surface was aerated or non-aerated. Both protocols are described below.

Sample collection from aerated surfaces

The square wooden sampling box (1mx1m) is used in aerated conditions (see Figure 6). A pipe connected to a gas mass counter allows the flow of air supplied by the aeration system - "real" Q - to be collected and quantified (in m³/h). A spur from this pipe allows the sample of air to be directed to the gas analyzer.

FIGURE 6

SAMPLING BOX FOR AERATED SURFACES



SAMPLE COLLECTION FROM NON-AERATED SURFACES

In case of non-aerated surfaces the sampling system used was the Odoflux dynamic flow chamber (see Figure 7).

FIGURE 7

SAMPLING BOX FOR NON-AERATED SURFACES



The flow chambers covered an area to be studied as hermetically as possible in order to isolate the surface from external conditions. The gases emitted by the isolated surface are collected by a vector gas which is injected into the chamber (Q1). The mixture of vector gas and gaseous effluent is then collected for analysis. The flow chamber used consists of an acrylic resin cylinder capped by a hemisphere, also of acrylic resin. The air-tightness of these two parts is ensured by polypropylene screws. The sweep air supply is fed by a pipe coiled against the wall and pierced with holes (flow Q1). The air sample is taken at the top of the hemisphere by means of a probe which has several holes along its length in order to ensure homogeneous sampling. The air is sucked in at flow Q2, of which 1l/min is directed to the analyser; the other part (Q2 - 1l/min) is released into the atmosphere.

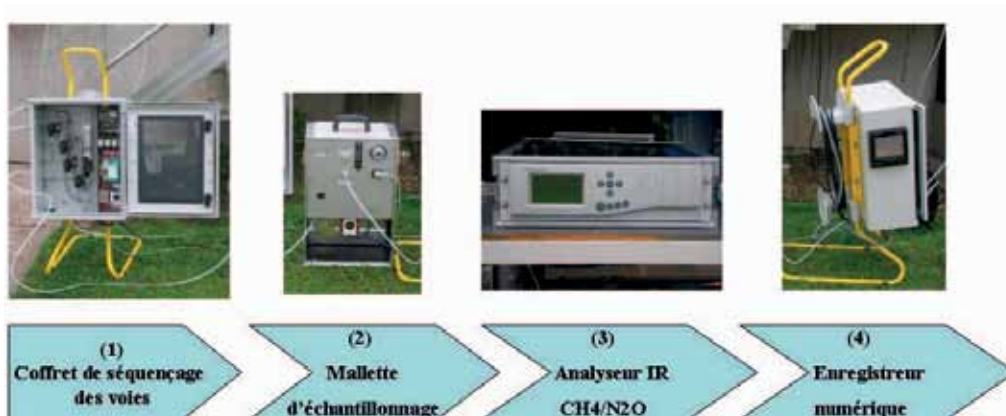
To respect “pseudo-isokinetic” conditions, the applied flow Q1 must be equal to the flow sucked in Q2. The protocol developed in the framework of this project for measuring GHG recommends a relatively weak flow ($Q1 = Q2$) of the order of 3 to 10l/min; this flow thus ensures “static chamber” conditions in which the surface of the liquid is not disturbed and the surface emissions are very slightly diluted.

ANALYSIS

The measured gas is sampled at the level of each spur using semi-supple teflon pipe, 4/6 mm in diameter. The measurement chain detailed below is illustrated in Figure 8.

FIGURE 8

MEASUREMENT CHAIN TO DETERMINE N₂O CONCENTRATION IN GAS SAMPLES; (ENGLISH VERSION OF THE PICTURE WAS NOT AVAILABLE, MEASUREMENT CHAIN IS EXPLAINED IN THE TEXT)



The collected gas is carried to the first module of the mobile analysis system, i.e. the measurement channel scrutiny system (1). On leaving this channel sequencing box, a single measurement channel is provided in the form of an exit via a semi-supple teflon pipe, 4/6 mm in diameter; this pipe is then connected to a sampling unit (2) which ensures the pre-conditioning of the measurement gas (Cooling of the gas using the Peltier effect, to reduce the dew point of the measurement gas and its exit regulation at a value of +5°C, Fine filter, with a 2µm filter cartridge, with humidity detection, flow-meter to ensure exact regulation of the flow of collected gas). When the collected gas leaves the portable sampling unit, it is then connected to the Servomex model 4210 gas analyser (3). The Servomex model 4210 analyser uses infra-red technology with gas filter correlation (GFC) for the compounds N₂O and CH₄. The data are taken continuously by the numerical recorder model QX/ HONEYWELL (5). The data thus recorded may be exported using Excel. The concentrations measured are expressed in ppm volume/volume.

GAS FLOW AND N₂O EMISSION

The resulting area flow of gas measured is calculated from the concentration measured at exit from the sampling system (N₂O or CH₄ in mg/m³ air) and from the air flow applied to the sampling system (m³/h).

The flow is related to the sampling area (area of the flow chamber in the case of non-aerated surfaces or the area of the sampling box in case of aerated surfaces), and is expressed in mg·h⁻¹·m⁻².

ADDITIONAL DATA

In order to understand the formation of N₂O during the treatment of waste water, the appearance of the different nitrous forms in biological treatment tanks, i.e. ammoniac (NH₄⁺-N), nitrates (NO₃⁻-N) and dissolved N₂O were monitored. These measurements would appear to be very necessary for monitoring the dynamics of the formation of N₂O emitted during waste water treatment processes. It likewise contains the protocol for measuring dissolved N₂O which was developed by the Rennes CEMAGREF and transferred to CIRSEE.

Ammonium and nitrate analysis

A VARIONPlus NH₄⁺-N / NO₃⁻-N (WTW) probe allows continuous measurement of these different nitrous forms in water. It uses specific ion measurements, which is a potentiometric method.

DISSOLVED N₂O

The protocol for the analysis of dissolved N₂O presented here was drawn up by the Rennes CEMAGREF. The water samples are taken and conditioned in 330 ml flasks, sealed hermetically with a septum. The sample conditioning stages are detailed below:

- Weigh 100 g of water sample in a 330 ml glass flask.
- Add 1 drop of sulphuric acid and close the flasks hermetically (stopper + septum).
- Place the samples in a container with warm water at 80 °C for 1 hr, the dissolved N₂O is then released into the head space.
- The gaseous fraction in the head space is then sampled using a gas syringe and injected into three 3 ml pill-boxes which have been previously conditioned in a vacuum;
- The analysis of the dissolved N₂O is done by gas chromatography - electron capture detector (GC/ECD) by the Rennes CEMAGREF, in France.

5.3 NITROUS OXIDE MEASUREMENTS USA**5.3.1 FIELD SAMPLING SITES**

Nitrous oxide emissions were monitored during this study from a wide range of activated sludge processes (both non-BNR and BNR). The modes of operation are summarized in this section according to the process configuration. For the sake of confidentiality, the specific locations of these processes are not be described. However, for geographic representativeness, it was ensured that these processes were broadly distributed around the North-East (4), Mid-Atlantic (2), Mid-West (2) and South-West (4) regions of the United States. The dimensions of the process bioreactors sampled and the sampling locations are further provided in Appendix 2 of this report. A description of the type of sampled processes is presented in Table 5.

TABLE 5 OVERVIEW OF TYPE OF PROCESSES SAMPLED IN N₂O RESEARCH

Plant Configuration	Description
Separate-stage BNR	The low-rate separate-stage nitrification denitrification process at this WWTP was sampled. The process was configured as a sequence of five reactors in series, as shown in Appendix 2. The influent to this process consisted of the clarified effluent from an upstream high-rate process, mainly engaged organic carbon removal. The influent was fed in a step-feed fashion to the first two aerobic zones. The last three zones of this process were non-aerated and the second non-aerated zone received methanol to promote denitrification. The effluent channel of this process was aerated prior to secondary clarification.
Four-stage Bardenpho	The four-stage Bardenpho process consisted of pre-denitrification (without external carbon addition) followed by a primary aerated zone, as shown in Appendix 2. The effluent of the primary aerated zone was internally recycled to the anoxic zone. Following the primary aerated zone was a de-oxygenation zone to scavenge dissolved oxygen, prior to methanol addition for enhanced denitrification. The final zone in this process was aerated primarily for stripping off the dinitrogen gas produced during denitrification, prior to secondary clarification.
Step-feed BNR 1	The four-pass step-feed BNR process sampled consisted of pre-anoxic zones comprising about 1/3 of the pass volume followed by aerated zones, as shown in Appendix 2. The transition zone between each pass was non-aerated to facilitate deoxygenation. The approximate influent flow split was 10%-40%-30%-20% to passes A, B, C and D, respectively. Pass A also received pre-settled anaerobic digestion centrate, which constituted approximately 30% of the influent TKN load to the process. Return activated sludge was also fed to Pass A.
Step-feed non-BNR	The step-feed non-BNR process sampled was configured and operated in four-pass step-aeration mode. The process was completely covered primarily for odor control. The headspace off-gases were consolidated and fed to a biofilter. The approximate influent flow split was 10%-40%-30%-20% to passes A, B, C and D, respectively. Return activated sludge was fed to Pass A.
Separate centrate	The separate centrate treatment process was operated to process pre-settled anaerobic digestion centrate and partially convert the influent NH ₄ ⁺ -N to NO ₂ ⁻ -N. The separate centrate treatment process was operated in plug flow mode, as shown in Appendix 2. Effluent from the separate centrate tank was fed to the overall plant return activated sludge line for possible bioaugmentation with primarily ammonia oxidizing bacteria (AOB) and for nitrogen removal via the short-cut nitrite pathway.
Plug-flow 1	The first plug-flow process sampled was designed and operated primarily for organic carbon removal and nitrification and did not have dedicated anoxic zones or external organic carbon addition. The process was configured in four-pass mode, as shown in Appendix 2.
Plug-flow 2	The second plug-flow process sampled was also designed and operated for organic carbon removal and nitrification and did not have dedicated anoxic zones or external organic carbon addition. The process was configured in two-pass mode, as shown in Appendix 2.
MLE 1	The first modified Lutzack Ettinger (MLE) process sampled was originally designed for operation in enhanced biological phosphorous removal mode, but subsequently operated in MLE mode. As shown in Appendix 2, the process consisted of pre-denitrification without external organic carbon addition. Biomass from zone 6, which was aerated, was internally recycled to anoxic zone 2. Return activated sludge was fed to anoxic zone 1.
MLE 2	The second modified Lutzack Ettinger (MLE) process sampled was also originally designed for operation in enhanced biological phosphorous removal mode, but subsequently operated in MLE mode. As shown in Appendix 2, the process consisted of pre-denitrification without external organic carbon addition. Biomass from zone 6, which was aerated, was internally recycled to anoxic zone 2. Return activated sludge was fed to anoxic zone 1.
Step-feed BNR 2	The second step-feed process sampled was configured in four-pass mode as shown in Appendix 2. Each pass consisted of pre-anoxic zones comprising 1/3 of the pass volume followed by aerobic zones. The approximate influent flow split was 50%-30%-20%-0% to passes A, B, C and D, respectively. The anoxic zones were mixed via low intensity pulse aeration. The return activated sludge was fed to Pass A.
Oxidation ditch	The oxidation ditch process shown in Appendix 2 was operated to achieve simultaneous nitrification and denitrification by operation are uniformly low aeration intensities and dissolved oxygen concentrations. The influent flow to the process was fed to the inner loop and was mixed and circulated using surface mixers. No external organic carbon was added to enhance denitrification. Return activated sludge was fed to the inner loop of the process.
Step-feed BNR 3	The third four-pass step-feed BNR process sampled consisted of pre-anoxic zones comprising about 1/3 of the pass volume followed by aerated zones, as shown in Appendix 2. The approximate influent flow split was 33.3%-33.3%-33.3%-0% to passes A, B, C and D, respectively. Pass A also received pre-settled anaerobic digestion centrate, which constituted approximately 40% of the influent TKN load to the process. Return activated sludge was also fed to Pass A. The reactors of this process were also covered and thus only composite measurements of the overall headspace could be performed.

5.3.2 SAMPLES COLLECTION AND ANALYSIS

One of the most significant outputs from the WERF project has been the development of a protocol that enables collection of N₂O emission fluxes from open-surface activated sludge bioreactors using consistent methodology. The protocol was submitted to the USEPA for review during late 2008 and the comments provided have been duly incorporated herein. The protocol is intended to provide utilities and field sampling teams with a detailed description of the data collection methodology and analysis requirements to enable calculation of gaseous nitrogen fluxes from different zones of activated sludge trains in a wastewater treatment facility.

The protocol was officially released during the 2nd Water Environment Federation Nutrient Removal Conference in Washington DC during June 2009 and is also available online at www.werf.org.

SAMPLING DESIGN FOR FULL-SCALE MONITORING

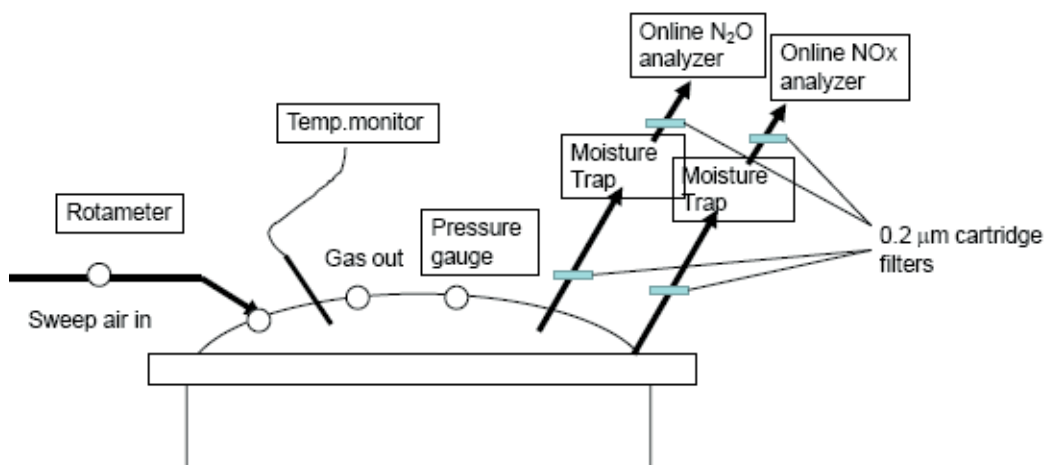
The N₂O emission fluxes of several wastewater treatment plants that are accomplishing nitrification and denitrification were determined. Testing was conducted at each plant during which gas phase monitoring was performed in real-time continuous mode and liquid phase sampling was performed via discrete grab sampling. Plants were subjected to two campaigns conducted once in warm temperature conditions (i.e. summer, early fall), and cold temperature conditions (winter/early spring) in the Northeast and Midwest and twice in plants along the West Coast (Fall and Spring), not subject to significant temperature changes.

SAMPLING PROCEDURES-HEADSPACE GAS MEASUREMENT

The overall procedure for measuring N₂O, NO and NO₂-N fluxes from the head-space of activated sludge tanks involves a variant of the EPA/600/8-86/008 and the South Coast Air Quality Management District (SCAQMD) tracer methods. This variant was developed to measure those sources that have a relatively high surface flux rate when compared to diffusion (for instance, spilled oil containment).

Commercially available replicas of the US EPA surface emission isolation flux chamber (SEIFC, Figure 9) were used to measure gaseous N fluxes from activated sludge reactors.

FIGURE 9 SCHEMATIC OF FLUX-CHAMBER EMPLOYED FOR N₂O MEASUREMENT.



The SEIFC consists of a floating enclosed space from which exhaust gas is collected in a real-time or discrete fashion. Since the surface area under the SEIFC can be measured, the specific flux of the gaseous compound of interest can be indirectly determined. The SEIFC ‘floats’ on the activated sludge tank surface and several replicate measurements can be taken at different locations in a single tank as well as from different tanks (nitrification, denitrification) along a treatment train.

The SEIFC is also equipped with mixing (physical mixer or via sweep gas circulation) to ensure adequate gas mixing and in some cases, an online temperature probe. The SEIFC is currently one of the few devices accepted by the USEPA for measuring gaseous fluxes (Tata et al., 2003) and as such will be employed for this study. Gas-phase analyses was conducted via infra-red (N₂O) and chemiluminescence (NO_x).

In general, sampling was conducted at multiple locations of the activated sludge train in each wastewater treatment facility. These locations included aerobic, anoxic and anaerobic zones, depending upon the configuration of the given facility. During the course of the gas phase sampling, liquid phase samples were collected adjacent to the hood location. The samples were filtered immediately upon collection in the field and analyzed by host plant personnel for ammonia, nitrite and nitrate concentration, utilizing readily available field methods (i.e. Hach kits) and standard laboratory analytical methods.

The specific locations selected were the geometric center of each demarcated anoxic or aerobic zone in the WWTP, or alternately locations where nitrification could be inferred based on initial screening of NH₄⁺-N and DO concentrations (as in the plug-flow processes). For discrete measurement at each of these locations, 30 replicate measurements of gaseous N₂O and 1 measurement of aqueous N₂O were obtained over a period of 30 min. During continuous measurement at each of these specific locations over a 24 hour period, gaseous N₂O concentrations were still measured at 1/min, while aqueous N₂O concentrations were measured four-five times per day. Independent replication at each location (on different days) was not conducted owing to practical limitations associated with such an extensive campaign.

SAMPLING PROCEDURES-AQUEOUS N₂O CONCENTRATION

Aqueous phase N₂O concentrations were measured using a miniaturized Clark-type sensor with an internal reference and a guard cathode (Unisense, Aarhus, Denmark). The sensor is equipped with an oxygen front guard, which prevents oxygen from interfering with the N₂O measurements. The sensor is connected to a high-sensitivity picoammeter and the cathode is polarized against the internal reference. Driven by the external partial pressure, N₂O from the environment will penetrate through the sensor tip membranes and be reduced at the metal cathode surface. The picoammeter converts the resulting reduction current to a signal. The internal guard cathode is also polarized and scavenges oxygen in the electrolyte, thus minimizing zero-current and pre-polarization time.

SAMPLING PROCEDURES-MEASUREMENT OF ADVECTIVE GAS FLOW RATE FROM AERATED ZONES

Advective flow of gas through the flux-chamber (Q_{emission}) in aerated zones was measured using a modification of ASTM method D1946. Briefly, a tracer gas consisting of 10,000 ppmv ($C_{\text{helium-tracer}}$) He was introduced into the flux-chamber at a known flow rate, Q_{tracer} (equation 1). He concentrations in the off-gas from the flux-chamber ($C_{\text{helium-Fc}}$) were measured using a field gas-chromatograph equipped with a thermal conductivity detector (GC-TCD). Q_{emission} was computed using equation 10.

$$Q_{\text{tracer}} * C_{\text{helium-tracer}} = (Q_{\text{tracer}} + Q_{\text{emission}}) * C_{\text{helium-FC}}$$

$$Q_{\text{emission}} = \frac{Q_{\text{tracer}} * (C_{\text{helium-tracer}} - C_{\text{helium-FC}})}{C_{\text{helium-FC}}} \quad (10)$$

SAMPLING PROCEDURES-MEASUREMENT OF ADVECTIVE GAS FLOW RATE FROM NON AERATED ZONES

The only modification to the protocol to measure the emission flow rate from *non-aerated zones* was the introduction of sweep gas (air) or carrier gas through the flux-chamber at a known flow rate (Q_{sweep}), in addition to the He tracer gas. The corresponding Q_{emission} was computed using equation 11. Addition of sweep gas is needed to promote mixing of the SEIFC contents, owing to the low advective gas flow from the anoxic-zone headspace. Sweep-air N₂O concentrations were always measured and typically below the detection limits of the N₂O analyzer.

$$Q_{\text{tracer}} * C_{\text{helium-tracer}} = (Q_{\text{tracer}} + Q_{\text{sweep}} + Q_{\text{emission}}) * C_{\text{helium-FC}}$$

$$Q_{\text{emission}} = \frac{Q_{\text{tracer}} * (C_{\text{helium-tracer}} - C_{\text{helium-FC}})}{C_{\text{helium-FC}}} - Q_{\text{sweep}} \quad (11)$$

During continuous N₂O measurements, Q_{emission} was determined several times a day.

Sampling Procedures-Wastewater and Process Characterization

PRELIMINARY DATA GATHERING AND STEADY STATE PROCESS ANALYSIS.

The integral dependence of N₂O and NO emissions on the process operating conditions made the development of a preliminary reconnaissance analysis crucial. The following background information was collected from candidate evaluation sites:

- **Overall Plant Description.** First, general information related to treatment plant configuration, liquid and solids process flow diagrams, design criteria, major mechanical process equipment, etc from the plant's design reports and/or O&M manuals were obtained. In addition, the following secondary process operating data was gathered via meetings with plant operations teams and process engineers:
- **Secondary Process Configuration.** including zone configuration, zone volumes, operating set points, basins in service, aeration flow and distribution, recycle streams and flow rates (if applicable)
- **Plant Operating Data.** Summary of a minimum of three months plant data applicable to the treatment process to allow for characterization of the process influent, target and actual operating set points for key operational parameters (DO, SRT), effluent concentrations.

INTENSIVE ON-SITE SAMPLING AND ANALYSIS

Intensive diurnal sampling was conducted during each N₂O sampling campaign at the different plants to develop correlations between process state variables and gas-phase N₂O and NO_x concentrations.

5.3.3 CALCULATION N₂O EMISSION

Determination of fluxes

The net flux of gaseous N species (mg/min-m²) was calculated based on the gas flow rate out of the flux chamber (Q_{emission}, L/min), gas concentration (parts per million) and the cross-sectional area of the SEIFC (m²) (Equation 12).

$$\text{Flux} = \frac{Q_{\text{emission}} * C}{A} \quad (12)$$

DETERMINATION OF LUMPED EMISSION FACTORS

The surface flux calculated from equation 12 was translated into the flux of a given zone by multiplying with the specific zone area. The N₂O emission fractions (mass/mass) for each WWTP at any given time point were computed by normalizing the measured flux from each zone in the facility to the daily influent total Kjeldahl nitrogen (TKN) loading according to equation 13. Emission fractions were averaged over the course of the diurnal sampling period and reported as the average (avg.) ± standard deviation (sd). for each individual process sampled.

During each campaign, wastewater nitrogen species concentrations including influent, bioreactor and effluent total Kjeldahl nitrogen (TKN), ammonium (NH₄⁺-N), nitrite (NO₂⁻-N) and nitrate (-N) were measured simultaneously about six-times per day according to Standard Methods (Eaton et al., 2005) to supplement the gas-phase measurements. The discrete measurements were averaged to generate the emission fractions described in equation 13. Additionally, seven out of the twelve processes were sampled at minimum and maximum annual wastewater temperatures to examine seasonal temperature impacts on N₂O generation and emission.

$$\text{Emission fraction} = \frac{\sum_{i=1}^n \text{Flux}_i * \text{Area}_i \text{ (kg N}_2\text{O-N)}}{\text{Daily influent TKN load (kg-N)}} \quad (13)$$

Where:

Flux_i = N₂O emission flux calculated from the ith zone (kgN₂O-N·m⁻²·d⁻¹)

Area_i = Surface area of the ith zone (m²)

n = number of zones in a given facility from which N₂O fluxes are captured

Daily influent TKN load: Average influent load (influent flow rate * influent TKN concentrations) over the course of 24 hours.

On average, wastewater characterization was performed at each gas sampling location as well as in the tank influent and effluent about six times per day. At facilities where analysis was not as frequent, daily composite measurements were employed, for instance in the influent and effluent samples. Alternately, in some facilities, online measurement devices (for measuring pH, dissolved oxygen, redox potential and select N-species, including NH₄⁺-N and -N) were employed at different locations of the activated sludge tank, which also facilitated the wastewater characterization efforts.

For aerobic zones, the helium-based advective gas-flow data were correlated to plant-recorded airflow rates for any given zone via linear regression and used to calculate diurnal N₂O emissions. For anoxic (non-aerated) zones lacking associated plant airflow data, the average of the experimentally obtained helium-based gas flow rates was used to calculate diurnal N₂O emissions.

CORRELATION BETWEEN WWTP OPERATING CONDITIONS AND N₂O EMISSIONS

Possible links between WWTP operating conditions and N₂O emission fluxes were examined via multivariate linear regression modeling of emissions and several wastewater state variables using SAS (Cary, NC). For aerobic zones, N₂O fluxes were correlated with liquid temperature and sampling location-specific pH, temperature, dissolved oxygen (DO), NH₄⁺-N and NO₂⁻-N concentrations and multiplicative combinations thereof. For anoxic zones, N₂O fluxes were correlated with sampling location-specific soluble chemical oxygen demand (sCOD), pH, temperature, DO, NO₂⁻-N and -N concentrations and interactive combinations thereof. Assumptions of state variable normality and equal variance were evaluated using error residual and covariance plots (not shown). The variables not normally distributed were log-transformed, verified for normality and equal-variance and subsequently used for regression modeling. Time points where all state variables had not been measured simultaneously were not included in regression analysis.

5.3.4 QUALITY CONTROL

A detailed quality assurance project plan (QAPP) was prepared and submitted along with the protocol to the USEPA for review. The specifics of the QAPP and related procedures used were verified independently by a Project Quality Officer from Rutgers-The State University of New Jersey.

The validity of the measurements using the protocol developed for this study was determined via a parallel sampling effort among three teams on September 9 and 10, 2008 at one of the test wastewater treatment facilities. The plant staff used an USEPA flux chamber technology and sampled nitrogen oxide compounds using a field photo-acoustic analyzer. The Columbia University-WERF team used a flux chamber manufactured by St. Croix Sensory and measured N₂O off-gas concentrations via gas-filter correlation, described above. A third team led by Dr. Charles Schmidt used an USEPA flux chamber and sampled the off-gas into opaque Tedlar® bags for FTIR analysis (NIOSH 6660) by a commercial laboratory (Peak Analytical, Boulder, CO).

Based on these parallel measurements, the three methods resulted in similar results (Figure 10), with good correspondence in both the N₂O concentrations and off-gas flow-rate (only done by the Columbia-WERF team and Dr. Schmidt's team, Figure 11). Briefly, the following observations were made based on the results obtained and incorporated into subsequent full-scale measurement campaigns.

The use of an inert gas tracer was demonstrated to be an appropriate method to determine the advective off-gas flow rate. This was an improvement over the initial method developed by the Columbia-WERF team based on successive dilution of the N₂O concentrations. The initial method was discontinued following the validation study and replaced with He-tracer based method to determine advective flow rate.

Significant spatial and temporal variability in the measured concentrations of headspace N₂O was observed by the Columbia-WERF and Schmidt teams (Figure 12). Therefore, for subsequent full-scale measurements, discrete measurements (once a day) of N₂O at different locations in

any given WWTP was discontinued. Rather, a substantially more involved sampling strategy that entailed 24 hour “real-time online monitoring” of emissions at each location was initiated.

FIGURE 10 COMPARISON BETWEEN N₂O FLUXES OBTAINED VIA THREE INDEPENDENT METHODS. NOTE, ONLY THE CES AND WERF FLUXES CAN BE DIRECTLY COMPARED SINCE THEY WERE CONDUCTED CONCURRENTLY; (1 LB IS EQUAL TO 0.45 KG)

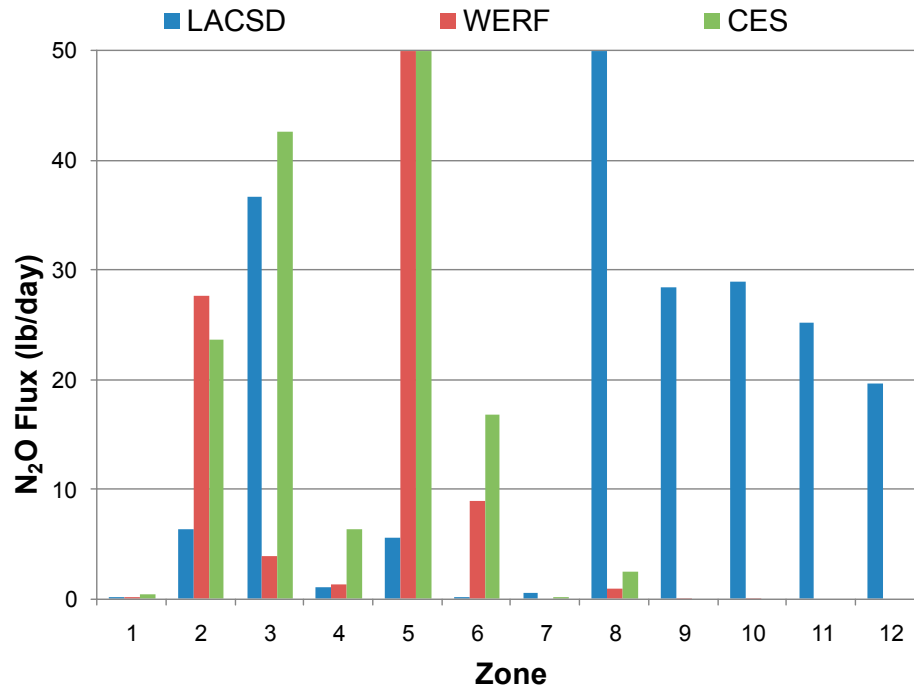


FIGURE 11 COMPARISON BETWEEN GAS FLOW RATES OBTAINED VIA THE TRACER GAS (CES) AND SUCCESSIVE DILUTION (WERF-COLUMBIA) METHODS

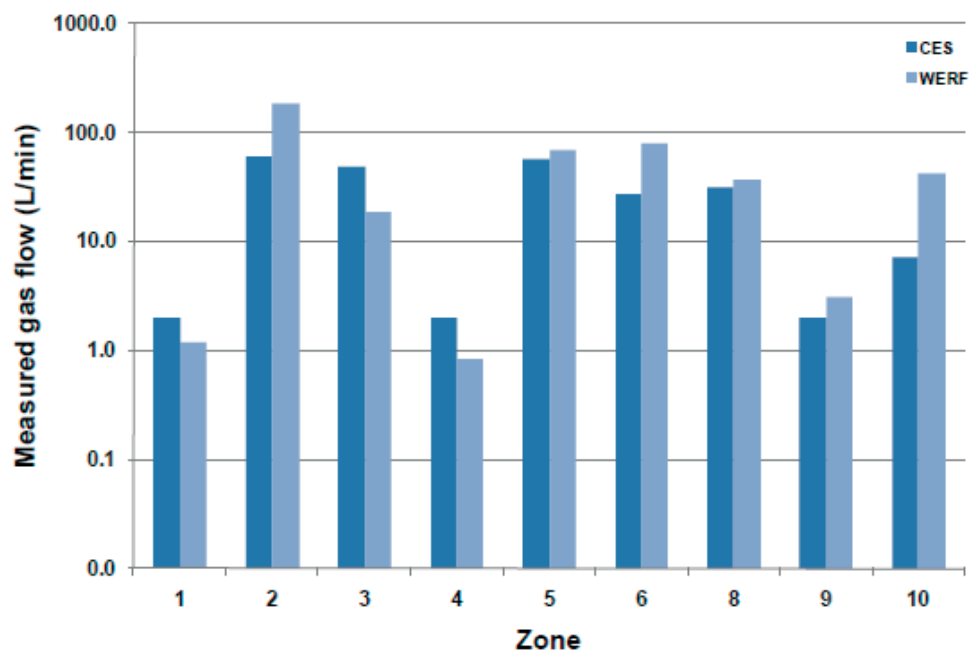
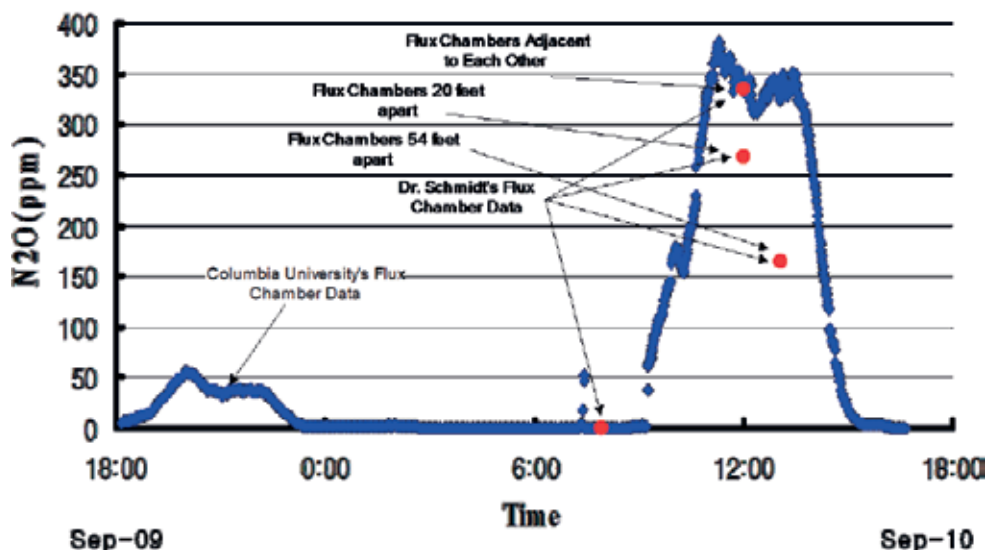


FIGURE 12 ILLUSTRATION OF SPATIAL AND TEMPORAL VARIABILITY IN N₂O CONCENTRATIONS IN THE HEADSPACE OF AN AEROBIC ZONE THAT NECESSITATES REAL-TIME ONLINE MONITORING. COLUMBIA UNIVERSITY'S FLUX CHAMBER DATA GIVEN BY NEAR CONTINUOUS BLUE DIAMONDS; DR. SCHMIDT'S FLUX CHAMBER DATA GIVEN BY 4 RED DOTS



5.4 NITROUS OXIDE MEASUREMENTS THE NETHERLANDS

5.4.1 FIELD SAMPLING SITES

Four measurement campaigns were performed in total. The WWTPs were selected to have covered activated sludge compartments. This allows for comparatively accurate knowledge on the gas flow. The measurement campaigns were done at two BNR carousel activated sludge plants, Carrousel 1 (25-29th of September 2008), Carrousel 2 (29th May - 4th of June 2009), and twice at a BNR (biological nitrogen removal) plant (14-21st of October 2008 and 9-16th of February 2009). The reason that at the BNR plant the emission of N₂O was measured twice was to study the effect of the temperature on the N₂O emission. In Table 6 the design characteristics of the WWTPs are shown.

TABLE 6 DESIGN CHARACTERISTICS OF THE INVESTIGATED WWTPS

	Unit	Carrousel 1	Carrousel 2	BNR
Location	-	Papendrecht	Kortenoord	Kralingseveer
Hydraulic load	m ³ /day	12,000	25,000	112,000
SRT (approximately)	days	23	unknown	20
Population equivalents	p.e. (54 gBZV)	40,000	100,000	360,000
Effluent requirements				
Nitrogen	mg/l	10	10	12
Phosphorus	mg/l	2	1	1.3
Organic load	kgCOD/day	3,540	13,880	25,000
Nitrogen load	kgN/day	410	1,200	3,700 ¹
Organic sludge load	kgCOD · kg dw ⁻¹ · d ⁻¹	0.031	0.060	0.048
Nitrogen sludge load	kgN · kg dw ⁻¹ · d ⁻¹	0.009	0.012	0.024
Pre-sedimentation		No	No	Yes
P-removal		Biological	70% biological, 30% chemical	Biological
Type		carrousel	Carrousel	Pre-denitrification and -nitrification followed by two parallel carrousels
Sludge digestion		No	No	Yes

1) Nitrogen load of the activated sludge units (after pre-sedimentation); the load includes nitrogen-rich internal flows from the sludge line

5.4.2 SAMPLES COLLECTION AND ANALYSIS

Nitrous oxide concentration

The emissions of N₂O, NO, O₂ and CO₂ were measured on-line using a custom-made analyzer (Rosemount, designed by the TU Delft). A switching-box equipped with four channels, controlled and measured four separate gas flows successively. The analyzer was set to take measurements every 60 seconds for a 180-second period, after which it would switch to the next channel, purge for 300 seconds and repeat. In each gas flow gaseous N₂O, NO, and CO₂ was measured by infra red, O₂ analysis was performed by para-magnetism (Emerson Process management Rosemount Analytical). All air flows through the analyzer were maintained at a flow of 1 L/min. Prior to passing through the analyzer all flows were led through condensers to prevent moisture from entering the analyzer.

At each WWTP the four channels were used to measure the N₂O concentration at different locations. An overview of the exact locations per WWTP is presented in Appendix 3. At all WWTPs one channel was used to measure the ambient air as a reference.

Additional grab-samples of off-gas from various (other) onsite processes were collected in gas sample bags. These were later analyzed for N₂O at the laboratory using gas chromatography (Varian CP-3800 Gas Chromatograph).

GAS FLOW

The flow of off gas through the pipes was measured using a pitot tube. The pitot tube was connected to a pressure probe (Testo 445, max. 10 hPa, accuracy +/- 0.03 hPa), which converted the pressure difference to a gas velocity. In this way the gas velocity was measured at least three times during the measuring campaign. During each measurement the gas velocity was measured at 20 – 50 individual points throughout the whole pipe. Based on the internal diameter of the pipe the gas flow was calculated. The pitot tube was used during the measurement campaigns in Papendrecht and Kralingseveer. During the measuring campaign at Kortenoord the gas velocity was measured with a hot wire anemometer (Testo 435-1, max. 20 m/s, accuracy +/- 0.03 m/s and +4% of measuring value). After the measurements the error of the measurement was determined. If this error was larger than the variation in the gasflow, the emission was calculated with an average gasflow, if the error was smaller than the emission was calculated with the daily gasflow.

ADDITIONAL DATA

In addition to the gas measurements, concentrations of ammonium, nitrate, COD, BOD, Nkj and phosphate were obtained via daily analysis (24-hour flow-dependent sample collection) by the Waterboard. During the measurement campaign, field test kits were used as an indication of fluctuations of key parameters over the day (especially nitrite). Furthermore, on-line data of the available sensors at the WWTPs are logged as part of the general plant operation and were obtained from the Waterboards operating the plants. One parameter that was continuously logged was the influent flow. With this flow and the available analysis of the influent the nitrogen load could be calculated.

5.4.3 CALCULATION N₂O EMISSION

The emission of N₂O is calculated based on the following measurements:

- N₂O concentration;
- gas flow;
- influent flow;
- NKj concentration (after primary settling including internal flow).

The total N₂O load was calculated, by averaging the N₂O concentration during the day. This average concentration was multiplied by an average gas flow. This could be done because it was found that the variation in the gas flow was smaller than the calculated error in the measurement.

The nitrogen load that entered the WWTP was calculated based on the measured NKj concentration and the average flow over the day.

5.4.4 QUALITY CONTROL

The N₂O analyser was two-point calibrated before every measuring campaign, using a standard gas for each individual gas (N₂O, NO, CO₂ and O₂). The accuracy of the analyser was 1%. The analyser was equipped with four measuring channels, one of them was always used to measure the ambient air. The concentrations in the ambient air were used to calculate the gas production (N₂O, NO, CO₂) or consumption (O₂). In this way it was also possible to neutralize a possible shift in the calibration line by per example a pressure difference.

For the quality control of the gas flow is referred to section 5.4.2.

5.5 METHANE MEASUREMENTS AUSTRALIA

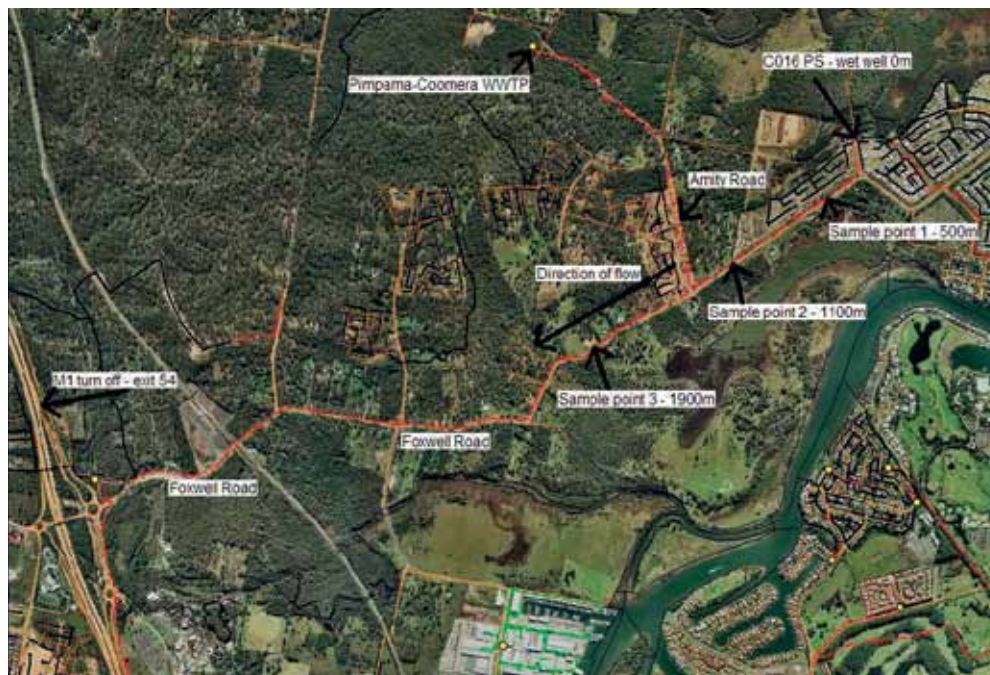
5.5.1 LIQUID PHASE MEASUREMENT

While on-line sensors for measuring dissolved CH₄ are available, the measurement campaigns conducted in Australia so far measuring dissolved CH₄ in rising main sewers have solely relied on the use of manual sampling followed by laboratory analysis of the CH₄ content in wastewater samples using GC.

FIELD SAMPLING SITES

The design of the measurement campaign is important. As will be discussed later, the hydraulic retention time (HRT) of wastewater in a sewer line is a critical parameter influencing the production of CH₄. To obtain a representative picture of CH₄ formation in a given sewer line, it is important to design the sampling campaign so that a wide range of HRT is covered. This would involve taking wastewater samples at multiple locations along the sewer line at multiple times. Figure 13 shows the site map of the C016 rising main sewer at the Gold Coast, Queensland, Australia, with the four sampling locations marked (Pumping Station, Sample Points 1, 2 and 3). In the sampling campaign reported in Foley et al. (2009), each of the sites was measured 3 – 4 times with an interval of 1 hour, which allowed to capture wastewater samples with HRT in the sewer line for 0 – 8.7 hours.

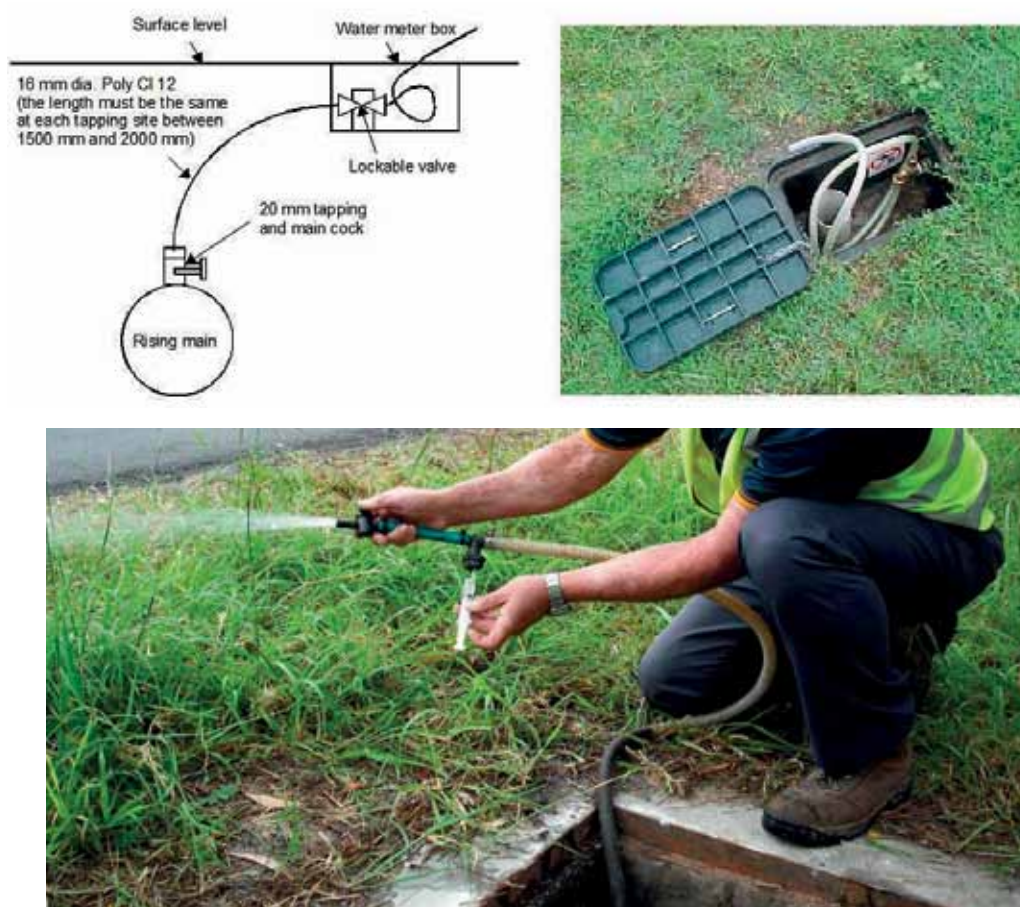
FIGURE 13 AERIAL PHOTOGRAPH OF THE C016 RISING MAIN. SAMPLES WERE COLLECTED FROM THE C016 PUMP STATION WET WELL (0M), SAMPLE POINT 1 (AT 500 M), SAMPLE POINT 2 (AT 1100 M) AND SAMPLE POINT 3 (AT 1900 M)



SAMPLE COLLECTION AND ANALYSIS

Wastewater sampling from **pressurized lines** was done through a special sampling arrangement (Figure 14). It consists of a 16 mm diameter pipe connecting a sampling tap at the ground level to the tapping arrangement attached to the underground pipe. Samples were collected using a hypodermic needle and 5 ml plastic syringe, attached directly to the pressurized rising main via a flexible hose. This procedure avoided any contact of the wastewater with atmosphere and possible oxygen interference.

FIGURE 14

UPPER LEFT AND UPPER RIGHT: THE SAMPLING SYSTEM. LOWER: COLLECTION OF WASTEWATER SAMPLES CONTAINING DISSOLVED CH₄.

Wastewater samples from **wet wells** were collected with a sampling device consisting of an open-head cylindrical container 40 cm in height and 1.5 l in volume. The sampler was submerged in the wet well for 5 – 10 seconds before its recovery. Samples were taken with 5ml plastic syringe from a location 10 cm below the water surface in the sampler container to avoid contact with air.

The above method was also used for wastewater sampling from a **tapping point** (as an alternative method to what described in Figure 14). Prior to fill up the sampling container, sewage was thoroughly flushed for a period of one minute. Then, the hose-tube connected to the sampling tap was placed in the sampling container, which was filled from the bottom. Container was flushed for 30 seconds prior to a sample collection. Samples were taken 10 cm below the water surface.

A sample collected was subsequently injected into freshly vacuumed BD Vacutainer[®] tubes through a 0.22 μ m pore diameter unit (Millipore, Millex GP). The Vacutainer tube was mixed overnight in a shaker to allow equilibration of gas and liquid phases. Most of the CH₄ (~97% at 25°C) would be transferred to the gas phase in this process (Alberto et al., 2000). The CH₄ concentration in the gas phase of the tube was measured using a Shimadzu GC-9A Gas Chromatograph equipped with a flame ionization detector (FID). The concentration of CH₄ in the initial liquid phase was then calculated using mass balance and Henry's law (Guisasola et al., 2008). The Vacutainer tube was weighed before and after sampling to determine the sample volume collected. This volume, along with the known volume of the Vacutainer tube, enables to calculate dissolved CH₄ contained in the original wastewater sample.

5.5.2 GAS PHASE MEASUREMENT

Due to safety considerations when operating in confined spaces, sewer workers in Australia commonly use portable gas detectors to detect toxic and flammable gasses. These sensors typically measure a number of gases including O₂, CO, H₂S, CO₂ and combustible gases such as CH₄. For CH₄ measurement, these sensors give direct readings of volumetric concentrations of CH₄ (%v/v) and/or percentages (%) of the lower explosive limit (LEL, which is approximately 5.1% v/v at 20°C). The detecting systems used include infrared sensors (IR), flame ionization detectors (FID) or catalytic bead sensors. These detectors are not specific for CH₄, and as such the measurement could be interfered by other hydrocarbons (e.g. ethane, propane, butane, etc). However, unless directly discharged, hydrocarbons other than CH₄ are not expected to be present in sewers in significant quantities. Therefore, the sensors should give correct readings of CH₄ concentrations in most cases despite of the possible cross interference. Automatic versions of the above gas detectors with data logging capabilities are also available for on-line CH₄ measurement. However, most of these sensors were developed for applications in other areas such as mining and landfills. Their application in the corrosive sewer environment is yet to be fully tested.

Methane concentration in sewer gas has also been measured through gas sampling (e.g. using Tedlar bags) followed by composition analysis using gas chromatography (GC). Compared to on-line sensors, this grab-sample based method could only give snapshots of CH₄ concentrations, which is a serious limitation. As will be discussed later, CH₄ concentration in both the liquid and gas phases is highly dynamic. It is difficult to capture the dynamics through offline sampling.

5.6 METHANE MEASUREMENTS FRANCE

The protocol for the CH₄ measurements in France is equal to the protocol used for the N₂O measurements as presented in section 5.2.

5.7 METHANE MEASUREMENTS USA

Work to date on the WERF (U2R08) project focuses on evolution of CH₄ from the collection system and from digester gas using equipment.

The research on the collection system has been performed in two phases, of which the first phase is completed and the second is underway. In the first phase the presence of CH₄ in the wet well was monitored instantaneously, in the second phase the presence of CH₄ was measured continuously (diurnal). In this report only the results of the first phase will be presented.

The work on the digester gas using equipment evaluated and estimated the fugitive CH₄ emissions from digester gas using equipment including Combined Heat and Power (CHP) technologies.

The work on the digester gas using equipment evaluated and estimated the fugitive CH₄ emissions from digester gas using equipment including Combined Heat and Power (CHP) technologies.

5.7.1 COLLECTION SYSTEM PHASE 1: FIELD SAMPLING SITES

The investigation on the collection system was performed in DeKalb County, Georgia. DeKalb County covers approximately 280 square miles (725 square km) and serves a population of over 700,000. In Annex 8 a map of the County is provided and delineates the major drainage basins.

The map also identifies major wastewater collection system infrastructure. DeKalb's wastewater collection system is extensive, comprising of approximately 2,900 miles (1,931 km) of sanitary sewer, 80,000 manholes and 64 sanitary sewage lift stations¹¹. Wastewater generated within the Snapfinger and Pole Bridge basins flows into one of DeKalb's two advance wastewater treatment facilities (AWTFs):

- Snapfinger Creek AWTF (36-mgd treatment capacity).
- Pole Bridge Creek AWTF (20-mgd treatment capacity)

A significant fraction of DeKalb's population lives within the Nancy Creek basin. Wastewater generated in the Nancy Creek basin is conveyed to City of Atlanta's R.M Clayton's WWTP via an 8 mile (12.87 km) long, 16 foot (4.88 m) diameter Nancy Creek Tunnel.

Of the County's 64 lift stations, 60 are small ranging in capacity from 80 to 700 gallons per minute (gpm) (0.3 – 2.6 m³/minute) and 4 are large with firm capacities of over 2,000 gpm (~7.6 m³/min) The small lift stations are not mechanically ventilated and have a gravity/passive ventilation.

The smaller lift stations comprise two general types of prefabricated units:

- Above-grade or Recessed. Includes a prefabricated concrete wetwell, four-level float controls, and two or four vacuum-primed pumps located directly above the wetwell (above grade) or in a recessed adjacent pit (recessed). Figure 16 shows a typical above-grade lift station.
- Below-grade. Has a separate prefabricated concrete or onsite constructed brick wetwell and prefabricated steel dry wells. Flooded-suction centrifugal pumps, motors, panel and switches are located below grade with ladder access.

A typical lift station in DeKalb County's selecting system is presented in Figure 15.

FIGURE 15

TYPICAL LIFT STATION IN DEKALB COUNTY'S SELECTING SYSTEM



¹¹ The lift stations were predominantly fed by gravity sewers and in more limited cases by rising mains or a combination of the two.

Three of the large lift stations include Lower Crooked Creek Pumping Stations (LCC) 1, 2 and 3. These three lift stations operate in series serving to convey the bulk of the flow coming from Gwinnett County to the Pole Bridge Creek AWTF. They are very similar in design including their pumping capacities and wetwell dimensions. Each lift station has a wetwell and dry well with constant-speed pumps. The wetwells of LCC-1 and -2 are ventilated, whereas LCC-3 has an open, unventilated wetwell. Figure 16 shows a photo of LCC-2. Honey Creek Pumping Station is the fourth large pumping station and also has a dry well/wetwell configuration. Similar to LCC-3, the wetwell of this lift station is open to the atmosphere.

FIGURE 16

LCC-PS-2



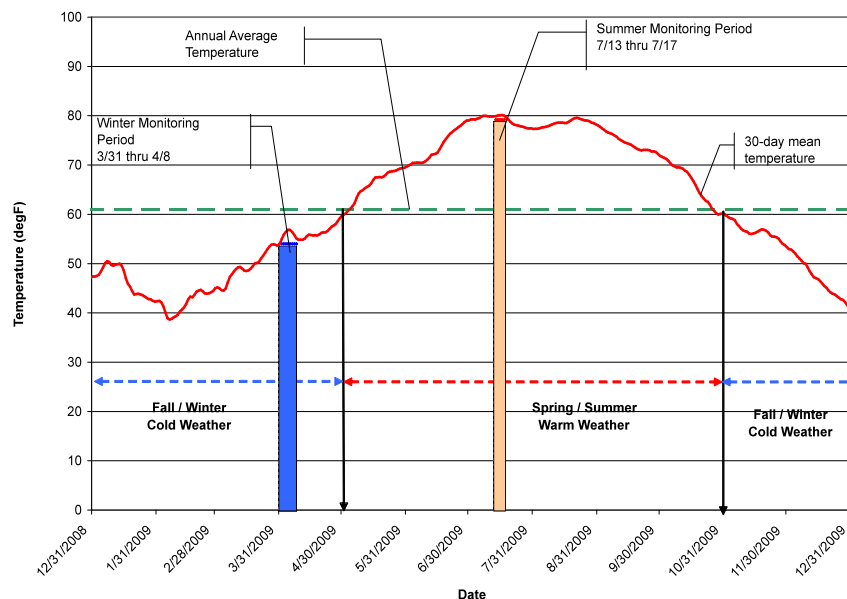
5.7.1 COLLECTION SYSTEM PHASE 1: TIME PERIODS

In order to study the effects of seasonal variations and temperature changes on the CH₄ emissions CH₄ monitoring was conducted during two discrete events: once during cold weather (winter monitoring) and once during warmer weather (summer monitoring).

The year was divided into 'cold' and 'warm' weather months. Cold weather months were selected as those with monthly average temperatures lower than the annual average temperature. Similarly, warm weather months were selected as those with monthly average temperatures higher than the annual average temperature. The selected cold and warm weather periods are depicted in Figure 17 together with the temperature profile of DeKalb County for the year 2009.

FIGURE 17

TEMPERATURE PROFILE OF DEKALB COUNTY FOR THE YEAR 2009 AND THE SAMPLING PERIODS



5.7.2 COLLECTION SYSTEM PHASE 1: SAMPLE COLLECTION AND ANALYSIS

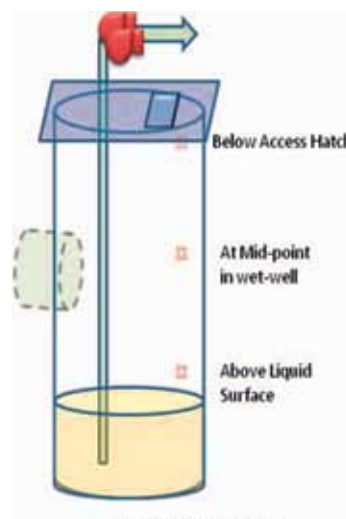
Instantaneous CH₄ readings from each of the 64 lift stations were conducted as part of this phase. In order to better understand the chemistry and biological pathways, other parameters such as temperature, H₂S, CO₂ and CO content were also collected.

SAMPLE COLLECTION IN UNVENTILATED LIFT STATIONS

Figure 18 shows the inside of a typical unventilated lift station in the DeKalb collection system. Methane concentrations were measured at three locations in each wetwell during each sampling event: immediately above the liquid surface, mid-way up the wetwell headspace; and immediately below the access hatch. While all three concentrations were collected, the highest of the three was used in order to somewhat mitigate the lack of accounting for dispersion of lighter-than-air CH₄ in between pumping cycles or the air dilution that results when air is pulled into the well during a pumping cycle. A similar procedure for data collection was used at large unventilated lift stations with open forebays including LCC PS-3 and Honey Creek.

FIGURE 18

UNVENTILATED WETWELL



SAMPLE COLLECTION IN VENTILATED LIFT STATIONS

There is only one ventilated lift station within DeKalb's collection system. At both LCC-1 the wetwell is covered and foul air exhaust fans continuously withdraw air from and send it to an odor control systems. Openings for the influent screens and at the wetwell covers allow air to enter the wetwell as foul air is exhausted. Instantaneous measurements for CH₄ and other parameters were sampled directly from the discharge header of the exhaust fan upstream of the biofilter.

ANALYSIS

Portable instruments were used for taking instantaneous readings in the field. A hand-held flame ionization detector (FID, MicroFID manufactured by PhotoVac) with hydrogen as a fuel source was used to measure CH₄. A portable four-gas analyzer with photoionization detector (PID, RKI Eagle Gas Portable Monitor) was used to measure H₂S, CO₂, %lower explosive limit (LEL) and CO. Ambient air and raw sewage temperatures were also monitored to check for a correlation with the measured CH₄ concentration. A dissolved oxygen (DO) probe (YSI 550A DO) was used to measure ambient air temperature, raw sewage temperature, and dissolved oxygen concentration in the raw sewage.

GAS FLOW IN UNVENTILATED LIFT STATIONS

The operation of constant-speed unventilated wetwells creates synchronized “breathing cycles” in the wetwell headspace. As the water level in a wetwell rises, the air inside the wetwell volume is expelled into the atmosphere through the vent or the wetwell cover. Conversely, when the wetwell water level is drawn down by pumping, outside air is drawn into the wetwell headspace.

Since all of the wetwells are at or near the atmospheric pressure, the amount of air breathed in and exhausted out during each pumping cycle is approximately equal to the active volume inside the wetwell or the volume of raw sewage pumped during each cycle. Thus, the total volume of air exhausted out of a given lift station is assumed equal to the gallons of raw sewage pumped over the same period.

During the separate DeKalb collection systems surveying effort, the active volume of each lift station was determined along with the pumping rate for each of the pumps in all 61 lift stations. In order to keep track of the monthly flow rate of the lift stations, DeKalb has installed pump-hour meters at each lift station and maintains monthly logs of hour-meter readings. These hour-meter logs along with the pumping capacities established during the collection systems study were used to determine the monthly flow rates for each of the lift stations. Using this method, monthly flows for the one-year period from September 2008 to August 2009 (the “model period”) were established for each of the smaller constant-speed lift stations.

For each of the unventilated lift stations, CH₄ readings from cold weather monitoring (March/April) were applied to monthly flows from November through April and CH₄ readings from warm weather monitoring were applied to monthly flows from May through October to estimate the annual CH₄ emissions.

While this approach allows an annual mass emission to be calculated for the DeKalb pumping stations, it is likely subject to be a number of limitations, namely:

- Dispersion of CH₄, which is lighter than air, is not accounted for. Many stations cycle only a few times per day and those stations likely lose a considerable portion of the evolved CH₄ to the atmosphere in between cycles. This phenomenon would result in under-reporting of actual emissions.
- Dispersion effects can be very significant at some of the larger pumping stations, like LCC-3. LCC-3 has a large, uncovered forebay with significant interchanges of outside air. On calm days the forebay air changes could be 1 to 5 times per hour and could increase to as much as 10 or 30 times per hour on windy days. This limitation, combined with the fact that these stations have the highest volume and mass throughput which should produce more CH₄, could represent a very significant source for under-reporting.
- During a pumping cycle, a significant volume of outside air is drawn into the wet well. If a sampling event occurred soon after such a dilution, the applied concentrations could under-report the actual CH₄ emissions.
- All of the sampling occurred during daylight hours and as such, correlated with higher load periods during the day that would be thought to coincide with periods of higher CH₄ production. This limitation could result in over-reporting of actual emissions.

GAS FLOW IN VENTILATED LIFT STATIONS

In order to compute the mass of CH₄ emissions, exhaust fan air flow rate and CH₄ concentration measured in the discharge header were used. In the absence of measured air flow data, the foul air fan flow rate was determined to be 1,950 cfm (cubic foot per minute) using

the manufacturer's fan curve and field-observed pressure conditions. The instantaneously-measured winter and summer CH₄ concentrations were applied to the 1,950 cfm in order to calculate the mass emissions from this station.

It is possible that this approach over reports the CH₄ emissions from LCC-1 in that the sampling occurred during higher flow periods when more flow and mass would be thought to produce higher-than-average CH₄ emissions.

5.8 METHANE MEASUREMENTS THE NETHERLANDS

5.8.1 FIELD SAMPLING SITES

The emission of CH₄ was determined at the same WWTPs as for the N₂O emission. The WWTPs at Papendrecht and Kortenoord do not have a sludge digester; one is present at WWTP Kralingseveer. Compared to the N₂O emission the emission of CH₄ was not measured continuously, but was measured via grab samples.

5.8.2 SAMPLES COLLECTION AND ANALYSIS

SAMPLE COLLECTION

Grab samples for the analysis of the CH₄ concentration were taken at the locations where emission of CH₄ could be expected. The amount of grab samples taken at each WWTP is summarized in Table 7.

TABLE 7 SUMMARY LOCATIONS CH₄ MEASUREMENTS AND NUMBER OF SAMPLES TAKEN

WWTP	Sample location	Number of samples	
Papendrecht	After inlet work and coarse screen	3	
	Selector	2	
	Anaerobic tank	2	
	Sludge loading	1	
	Carousels	1	
Kortenoord	After inlet work and coarse screen and grid removal	5	
	Selector	5	
	Aeration tank (3 channels N ₂ O measurements)	3	
	Sludge thickeners	3	
	Sludge storage	3	
	Sludge dewatering site	4	
Kralingseveer		October	February
	After inlet work and coarse screen	2	3
	Primary settling	2	3
	Selector	2	3
	Sludge thickeners	2	3
	Sludge storage	2	3
	Aeration tank	2	2
	Carousel 1 and 2	2	3

ANALYSIS

The grab samples were analyzed for CH₄ at the laboratory using gas chromatography (Varian CP-3800 Gas Chromatograph).

GAS FLOW

The gas flow was determined in the same way as for the emission of N₂O (see section 5.4.2).

CALCULATIONS

The emission of CH₄ is based on the following measurements:

- CH₄ concentration in grab samples;
- Gas flow;
- Influent flow;
- COD concentration influent (after primary settling, including internal flow).

The CH₄ emission was calculated based on the average concentration in the grab samples. In almost all cases¹² the average concentration was multiplied by the average gas flow as the error in the measurements was larger than the variations in the gas flow.

5.9 TOTAL CARBON FOOTPRINT WWTP

In the research performed in the Netherlands the emission of N₂O and CH₄ was measured at the same three WWTPs. Based on these measurements it was possible to determine the contribution of N₂O and CH₄ to the total carbon footprint of a WWTP including the use of electricity and natural gas. To determine the total carbon footprint of a WWTP all sources were converted to CO₂ equivalents. The conversions factors that have been used in this case are summarised in Table 8.

TABLE 8

CONVERSION FACTORS FOR CLIMATE FOOTPRINT CALCULATIONS USED IN THE NETHERLANDS

	Conversion factor	Unit
Electricity	0.67	kg CO ₂ /kWh
Natural gas	1.8	kg CO ₂ /Nm ³
N ₂ O	298 ¹⁾	kg CO ₂ /kg N ₂ O
CH ₄	25 ¹⁾	kg CO ₂ / kg CH ₄

1) GWP AR4: Adjusted GWP in IPCC Fourth assessment report, 2007 (IPCC,2007)

The conversion factors mentioned for electricity and natural gas in Table 8 are specific for the situation in the Netherlands, for any other country other factors do apply.

¹² Exceptions were selector Papendrecht, sludge storage in Kralingseveer (February), here the daily gas flow was used.

6

RESULTS

6.1 NITROUS OXIDE EMISSION AUSTRALIA

6.1.1 EMISSION AND ORIGIN

Figure 19 shows the dissolved N₂O concentrations (and associated 95% confidence intervals) measured by the microsensor in each reactor zone, at each WWTP. A wide range of values is apparent across the reactor zones, sampling rounds and WWTPs, from near zero to greater than 1 mg/l.

FIGURE 19 DISSOLVED N₂O CONCENTRATIONS MEASURED AT EACH WWTP. ERROR BARS INDICATE THE 95% CONFIDENCE INTERVAL, BASED ON THE MICROSENSOR SAMPLING DATA

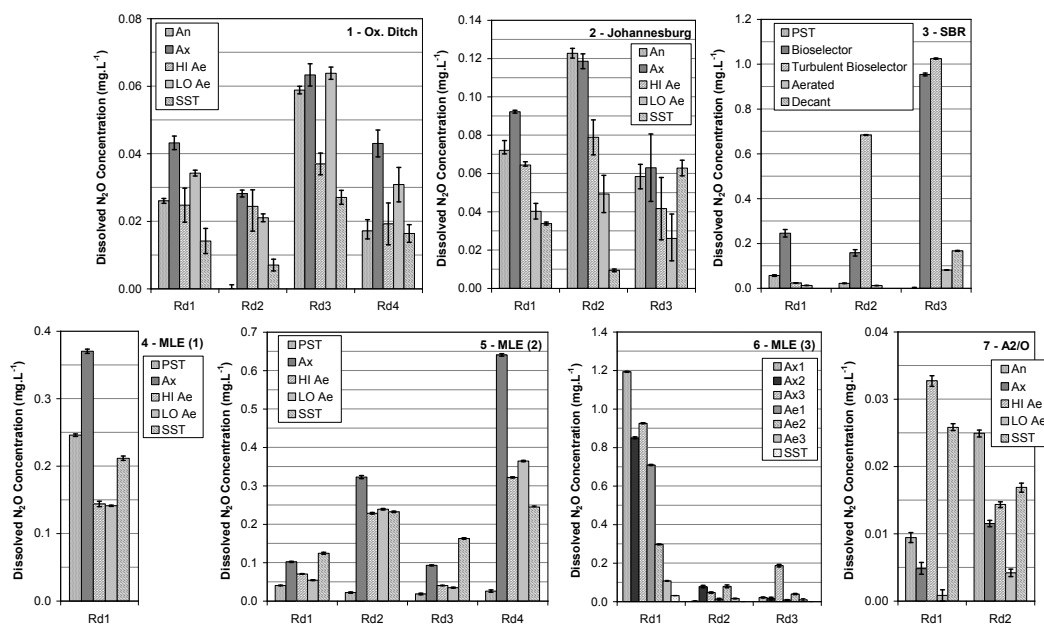
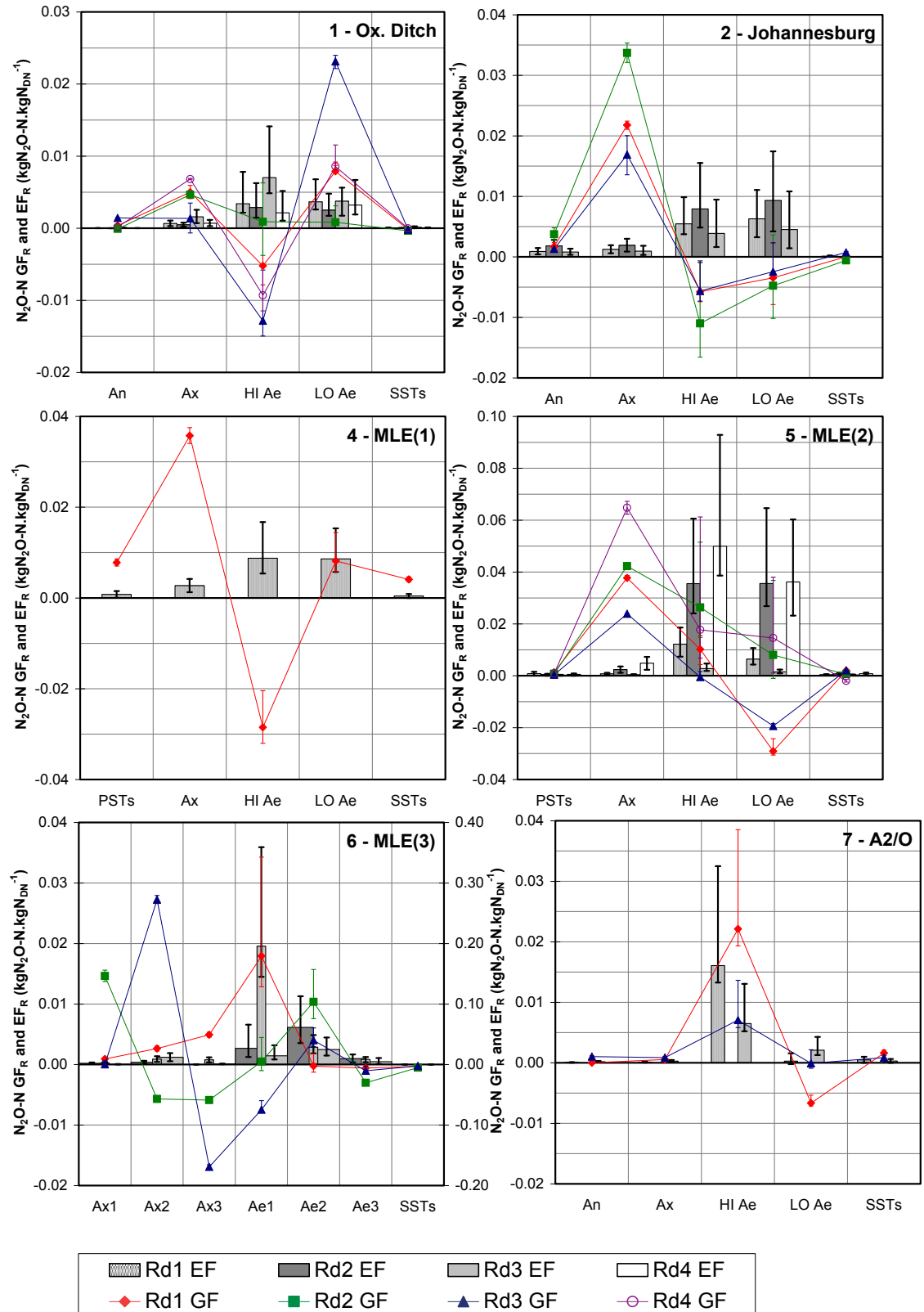


Figure 20 shows the normalised net N₂O-N generation (GF_R) and mass transfer emission profiles (EF_R), per reactor zone and sampling round, at each of the continuous flow WWTPs

FIGURE 20

NET N₂O-N GENERATION (GF_R) AND MASS TRANSFER EMISSIONS (EF_R) PROFILES, PER REACTOR ZONE, IN EACH SAMPLING ROUND AT THE SIX CONTINUOUS FLOW WWTPS. "AN" – ANAEROBIC ZONE, "AX" – ANOXIC ZONE, "HI AE" – HIGHLY AERATED AEROBIC ZONE, "LO AE" – LESS AERATED AEROBIC ZONE, "PSTs" – PRIMARY SEDIMENTATION TANKS, "SSTs" – SECONDARY SEDIMENTATION TANKS. RESULTS FROM ROUND 1 AT WWTP NO. 6 ARE PLOTTED ON THE SECONDARY AXIS – NOTE THE CHANGE IN SCALE. ERROR BARS INDICATE THE COMBINED UNCERTAINTY FROM THE 95% CONFIDENCE INTERVALS OF BOTH KLA AND MEASURED DISSOLVED N₂O CONCENTRATIONS



The calculated total WWTP generation factors (GF_{WWTP}) for the 20 sampling rounds are reported in Table 9 and show a wide range of results across two orders of magnitude. The minimum generation factor was 0.002, and the maximum 0.112 kgN₂O-N/kgTKN_{influent}. The average of the 20 samples was 0.016 ± 0.012 (t-dist, $\alpha = 0.05$). However, close inspection of these results shows the average to be skewed upwards by four results greater than 0.02 kgN₂O-N per kgTKN_{influent}. Excluding these results from the statistical sample, the average of the remaining 16 results is 0.006 ± 0.002 kgN₂O-N/kgTKN_{influent}.

Table 9 also reports the percentage of generated N₂O-N that is lost via mass transfer to the atmosphere. Generated N₂O-N can also be lost as dissolved N₂O-N in the WWTP effluent and waste solids. However, it is clear from the results in Table 9, that these losses are generally minor (i.e. < 5%). Whilst N₂O is a highly soluble gas in water, it also has a high mass transfer coefficient (similar to oxygen), so it is not unexpected that the majority of N₂O generated in the bioreactors is quickly stripped to the atmosphere.

TABLE 9 NITROUS OXIDE GENERATION FACTOR (GF_{WWTP}) AND % MASS TRANSFER EMISSIONS ($\%TR_{WWTP}$) FOR EACH WWTP AND SAMPLE ROUND

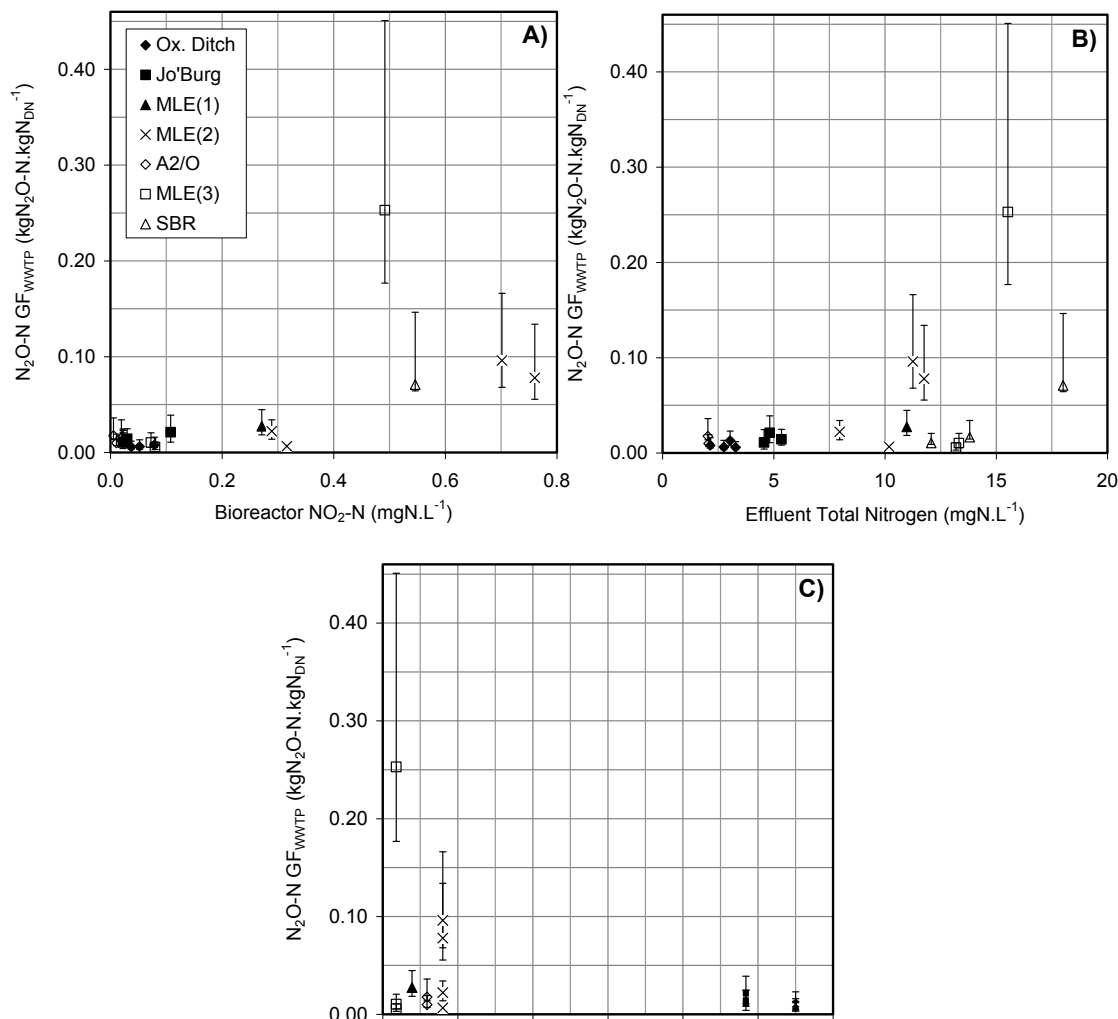
WWTP No.	N ₂ O-N GF _{WWTP} (kg N ₂ O-N/kgTKN _{influent})		N ₂ O-N Mass Transfer Emissions (%T _{WWTP})					
	Round 1	Round 2	Round 3	Round 4	Round 1	Round 2	Round 3	Round 4
1 – Ox. Ditch	0.003 (0.002 – 0.006)	0.002 (0.001 – 0.004)	0.004 (0.002 – 0.007)	0.002 (0.001 – 0.005)	98.2 – 99.0%	98.5 – 99.3%	95.3 – 98.2%	96.3 – 98.7%
2 – Johannesburg	0.010 (0.006 – 0.017)	0.017 (0.009 – 0.031)	0.007 (0.003 – 0.017)		96.8 – 98.8%	99.0 – 99.6%	88.4 – 97.7%	
3 – SBR	0.006 (0.005 – 0.011)	0.007 (0.006 – 0.014)	0.032 ¹ (0.029 – 0.065)		94.6 – 97.4%	94.6 – 97.5%	93.7 – 97.2%	
4 – MLE (1)	0.012 (0.008 – 0.020)				67.7 – 86.3%			
5 – MLE (2)	0.009 (0.006 – 0.015)	0.030 ¹ (0.021 – 0.051)	0.004 (0.002 – 0.005)	0.035 ¹ (0.025 – 0.061)	88.3 – 95.3%	94.1 – 97.6%	71.4 – 87.4%	94.5 – 97.8%
6 – MLE (3)	0.112 ¹ (0.078 – 0.200)	0.005 (0.003 – 0.010)	0.003 (0.002 – 0.006)		99.6 – 99.8%	99.7 – 99.9%	99.9%	
7 – A ² O	0.007 (0.006 – 0.015)		0.004 (0.003 – 0.008)		95.1 – 98.0%		94.0 – 97.6%	

Notes:

1. Bold results are greater than 0.02 kgN2O-N/kgTKN-1influent.
2. For the purposes of this report, the units of GFWWTP have been converted from kg N2O-N/kgN-1denitrified to kg N2O-N/kgTKN-1influent.
3. Figures in brackets represent the uncertainty range for GFWWTP results, based on the combination of 95% confidence intervals for both kLa values and measured dissolved N2O concentrations.

Shown in Figure 21 are the N₂O-N generation factor results plotted against bulk bioreactor NO₂-N concentration, and two process design parameters, namely effluent total nitrogen and a-recycle rate (i.e. recycle rate from aerobic zone to anoxic zone). Figure 21A suggests that the high N₂O-N generation factors generally correspond with high bulk NO₂-N concentrations in the bioreactor. Figure 21B and Figure 21C together show particular design parameters that potentially influence the stability of WWTPs in relation to N₂O generation.

FIGURE 21 NET N₂O-N GENERATION FACTOR, GF_{WWTP} , IN EACH SAMPLING ROUND, PLOTTED AGAINST A) BULK BIOREACTOR NITRITE-N CONCENTRATION, B) EFFLUENT TOTAL NITROGEN, AND C) A-RECYCLE RATE (AEROBIC \otimes ANOXIC) AS A MULTIPLE OF THE AVERAGE INFLUENT FLOWRATE, Q



The predominance of mass transfer emissions from the aerated zones is in agreement with other full-scale studies of aerated and quiescent bioreactors (Czepiel et al., 1995).

The profiles in Figure 20 show that whilst little atmospheric emission occurs from the quiescent anoxic zones, they actually are responsible for a substantial portion of the N₂O generated in the bioreactor. Examination of the net generation profile, rather than the emission profile, is therefore more useful in the context of understanding (and possibly controlling) the underlying mechanisms of N₂O formation.

The measurement of positive dissolved N₂O concentrations in all reactor zones across all seven WWTPs (refer to Figure 19) suggests that N₂O is pervasive in most BNR plants. Furthermore, the calculation of positive N₂O-N generation factors for all WWTPs in Table 9 confirms that these facilities are sources of anthropogenic greenhouse gas emissions for UNFCCC accounting purposes. It should also be noted that the generation factors for all sample rounds were positive, even when accounting for the uncertainty of a best-case combination of a low mass transfer coefficient and low dissolved N₂O concentration.

The variation in generation factor results in Table 9 highlights two key points. Firstly, WWTP Nos. 1 and 2 had low generation factors, with low variability. This was true also for the limited sampling rounds at WWTP No.7. Secondly, WWTP Nos. 3, 5 and 6 generally had higher generation factors, but also exhibited significantly higher variability over a short time frame. For example at WWTP No.5, the generation factor varied from 0.009 kgN₂O-N/kgTKN_{influent} in the morning sampling round, to 0.030 in the afternoon sampling, decreasing to 0.004 the following morning, and then increased again to 0.035 that afternoon. The results also highlight that some WWTPs might operate steadily with relatively low N₂O generation, and then suffer some process perturbation that leads to a temporary spike in N₂O formation¹³. This was witnessed most noticeably at WWTP No.6, which had a peak N₂O GF of 0.112 kgN₂O-N per kgTKN_{influent} in the first sampling round, but relatively low GFs in the remaining sampling rounds. Together, these two points indicate that both process design and variability in operating process conditions are likely to influence the magnitude and variability of N₂O-N generation factors.

6.1.2 PROCESS PARAMETERS OF INFLUENCE

Figure 21A suggests that high N₂O-N generation factors generally correspond with higher bulk NO₂⁻-N concentrations in the bioreactor. Although the number of data points is limited, there appears to be a threshold value at approximately 0.3 - 0.5 mg.l⁻¹ NO₂⁻-N at which the generation factor jumps to be >> 0.03 kgN₂O-N/kgN_{denitrified}. All four results > 0.03 kgN₂O-N/kgN_{denitrified} occurred under conditions where bulk NO₂⁻-N concentrations exceeded 0.4 mg/l.

Figure 21B and Figure 21C together show particular design parameters that potentially influence the stability of WWTPs in relation to N₂O generation. Both WWTP Nos. 1 and 2 have very high a-recycle rates and correspondingly low effluent TN concentrations. The WWTPs that had more highly variable N₂O-N generation factors tended to have lower a-recycle rates and correspondingly higher effluent TN concentrations. From Figure 21B, there appears to be a threshold at approximately TN 10 mg/l, below which the N₂O-N generation factor is relatively low and more stable.

¹³ That a temporary peak in N₂O emission is possible as a result of a process perturbation was also found by Burgess (2002). In this case a peak of N₂O was found after a peak in NH₄ or a disturbance in the aeration.

6.2 NITROUS OXIDE EMISSION FRANCE

6.2.1 EMISSION OF N₂O

The emission of N₂O that was measured at the four WWTPs is presented in Table 10.

TABLE 10 EMISSION OF N₂O AT FOUR WWTPS IN FRANCE

WWTP	Emission (kg N ₂ O/kg TKN _{influent})	Emission (kg N ₂ O-N/kg TKN _{influent})
WWTP 1	<0.0002	<0.0001
WWTP 2	<0.0002	<0.0001
WWTP 3	0.003	0.0017
WWTP 4	0.002	0.0011

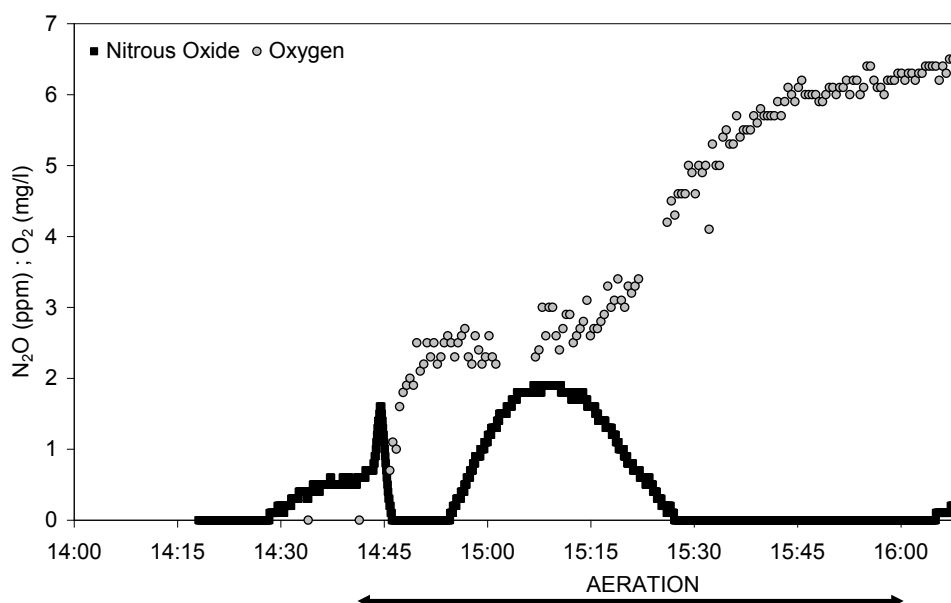
6.2.2 ORIGIN N₂O EMISSION

In all cases N₂O emissions were emitted in aerated zones. Emissions profiles show that some N₂O can be produced during non aerated (anoxic) steps but it is always emitted during aerated steps (stripping). This is illustrated with the results from WWTP 1, 3 and 4.

WWTP 1, CSTR

Emissions of N₂O during nitrogen removal were investigated in a completely stirred tank reactor (activated sludge) at full scale WWTP (WWTP1). The emission of N₂O around the start of the aeration is presented in Figure 22.

FIGURE 22 N₂O EMISSION DURING NITRIFICATION – 06/2007



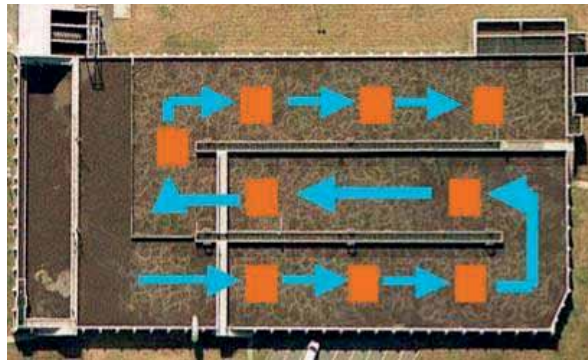
From Figure 22 the following can be observed:

- A first peak of N₂O at the start of the aeration. This is probably due to stripping of N₂O that was previously generated.
- A second peak of emission is observed during the nitrification phase. As can be observed N₂O emissions are linked to nitrification. Dissolved O₂ was around 2 mg/l during the nitrification and increased to 6 mg/l when all NH₄⁺-N has been oxidized (no data on the NH₄ concentration in the basin).

WWTP 3, PLUG FLOW REACTOR

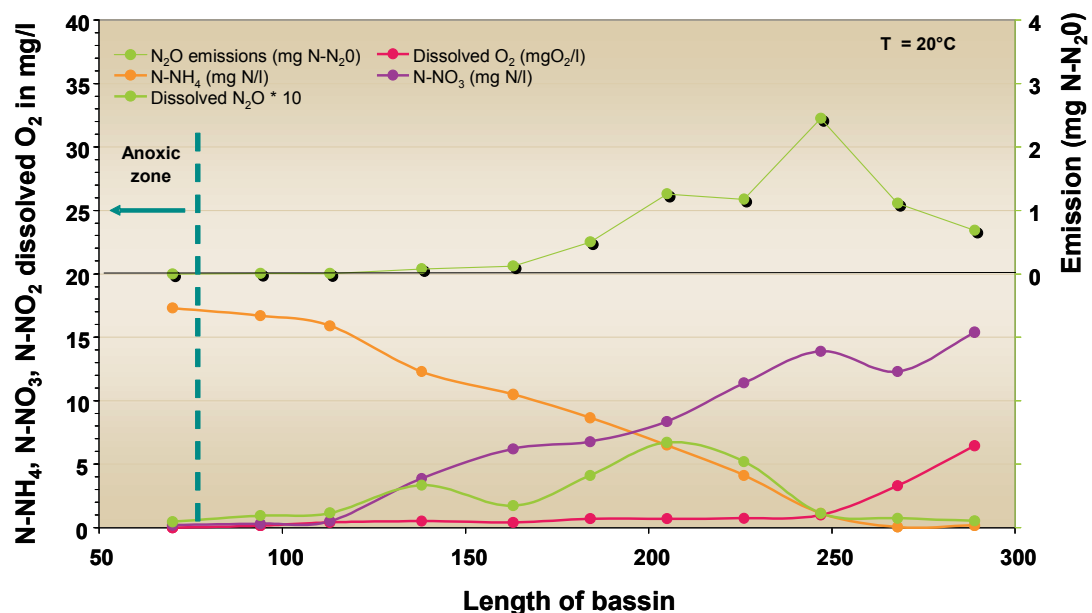
Emissions of N₂O during nitrogen removal were investigated in a plug flow activated sludge tank at a full scale WWTP (WWTP 3). Nitrous oxide was sampled at nine points (see Figure 23) with the square wooden sampling box as described in 5.2.2, following a 'kinetic' approach. All N forms were simultaneously monitored in water (NH₄⁺-N, NO₂⁻-N, ⁻-N, and dissolved N₂O) and in air (N₂O).

FIGURE 23

SAMPLING POINTS FOR N₂O MEASUREMENTS AT WWTP 3 IN FRANCE

The N₂O generation and emission during the aerated step in a plug flow system is presented in Figure 24. The first 80 metres of the basin is an anoxic zone where denitrification occurs. No N₂O emissions have been observed during denitrification. In the aerated length (80-290m) NH₄⁺-N and ⁻-N concentrations account for nitrification (NH₄⁺-N oxidation). The influent concentration was 17 mg NH₄⁺-N/l without nitrate neither dissolved N₂O. During nitrification NH₄⁺-N has been removed while the nitrate concentration has increased to 15 mg ⁻-N/l. Dissolved N₂O was measured and the curve indicates that N₂O is generated during the nitrification step. As shown on the upper part of the graph N₂O emissions were observed and have been quantified. Dissolved oxygen was 0.5-1mg/l and increased to 5 mg/l when nitrification has been completed –i.e. when overall ammonium has been oxidized.

FIGURE 24

EMISSION OF N₂O DURING NITRIFICATION IN A FULL SCALE PLUG FLOW ACTIVATED SLUDGE TANK AT WWTP 3

WWTP 4, ACTIVATED SLUDGE WITH SEQUENCED AERATION FOLLOWED BY A MBR

In this WWTP it was observed that:

- At the start of the aeration a first peak of N₂O appeared.
- A second peak of N₂O appeared during the course of the aeration period.

6.2.3 PROCESS PARAMETERS OF INFLUENCE

Based on the work in France the following process parameters of influence have been identified: NH₄⁺-N and DO. Furthermore air-flow must also be considered regarding stripping of N₂O.

6.3 NITROUS OXIDE EMISSION UNITED STATES OF AMERICA**6.3.1 EMISSION OF N₂O**

A wide range of N₂O emissions was measured across the twelve WWTPs operated at different temperatures, configurations and influent characteristics (Table 11).

TABLE 11 SUMMARY OF N₂O FLUXES AND EMISSION FACTORS MEASURED AT FULL-SCALE WWTPS.¹⁴

Plant Configuration	Temp (°C)	Reactor influent TKN load (g-N/day)	Reactor effluent TN load (g-N/day)	Q (MGD)	kg N ₂ O-N / kg N _{influent} (%)	kg N ₂ O-N / kg N _{denitrified} (%)
Separate-stage BNR	15 ± 0.48	1.8 · 10 ⁶	3.6 · 10 ⁵	23	0.03 ± 0.00	0.03 ± 0.01
	23 ± 0.28	2.3 · 10 ⁶	4.3 · 10 ⁵	27	0.01 ± 0.00	0.01 ± 0.00
Four-stage Bardenpho	14 ± 0.26	8.6 · 10 ⁵	1.7 · 10 ⁵	7.8	0.16 ± 0.10	0.19 ± 0.12
	23 ± 0.20	7.4 · 10 ⁵	7.6 · 10 ⁴	8.1	0.60 ± 0.29	0.66 ± 0.32
Step-feed BNR 1	19 ± 0.22	3.1 · 10 ⁶	1.4 · 10 ⁶	29	1.6 ± 0.83	2.9 ± 1.5
	25 ± 0.28	2.9 · 10 ⁶	9.4 · 10 ⁵	30	0.62 ± 0.27	0.90 ± 0.39
Step-feed non-BNR	17 ± 0.12	8.6 · 10 ⁶	4.4 · 10 ⁶	71	0.18 ± 0.18	0.37 ± 0.36
	26 ± 0.81	8.9 · 10 ⁶	4.2 · 10 ⁶	93	1.8 ± 0.79	3.3 ± 1.5
Separate centrate*	30 ± 2.3	8.8 · 10 ⁶	5.5 · 10 ⁶	2.0	0.24 ± 0.02	0.63 ± 0.06
	34 ± 0.32	8.5 · 10 ⁶	4.2 · 10 ⁶	1.6	0.54 ± 0.16	0.96 ± 0.32
Plug-flow 1	11 ± 0.20	1.8 · 10 ⁶	1.0 · 10 ⁶	18	0.40 ± 0.14	0.92 ± 0.32
	23 ± 0.46	1.8 · 10 ⁶	7.3 · 10 ⁵	15	0.41 ± 0.14	0.70 ± 0.24
Plug-flow 2	11 ± 0.41	6.3 · 10 ⁵	4.0 · 10 ⁵	8.7	0.62 ± 0.15	1.7 ± 0.41
	22 ± 0.58	6.6 · 10 ⁵	4.0 · 10 ⁵	6.6	0.09 ± 0.03	0.22 ± 0.06
MLE 1	26 ± 1.8	6.8 · 10 ⁵	1.9 · 10 ⁵	4.0	0.07 ± 0.04	0.09 ± 0.05
MLE 2	26 ± 0.17	6.9 · 10 ⁵	1.5 · 10 ⁵	4.1	0.06 ± 0.02	0.07 ± 0.03
Step-feed BNR 2	29 ± 0.18	2.2 · 10 ⁶	2.9 · 10 ⁵	14	1.5 ± 0.02	1.7 ± 0.02
Oxidation ditch	19 ± 0.58	3.9 · 10 ⁵	4.3 · 10 ⁴	3.4	0.03 ± 0.01	0.03 ± 0.01
Step-feed BNR 3	24 ± 0.78	7.8 · 10 ⁶	8.6 · 10 ⁵	57	0.05 ± 0.03	0.06 ± 0.03

* Flow normalized emission factors for centrate are inappropriate since centrate constitutes a miniscule flow rate, while containing up to 30% of the influent TKN load

14 At the first two facilities sampled, both discrete short-term (lasting about 30 min) and continuous (lasting 24 hours) N₂O measurements were conducted in multiple locations or zones. However, subsequently, the 30 min discrete measurements are not reported, since they did not capture the considerable temporal variability observed in each zone. For following campaigns, each location or zone was subjected only to 24 diurnal continuous monitoring

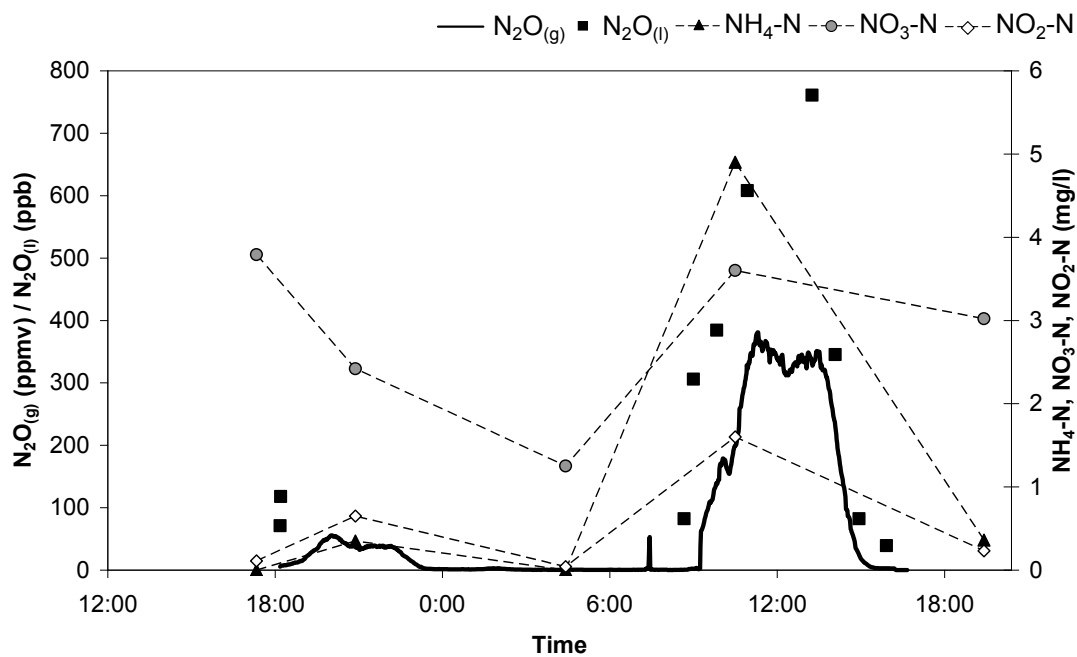
From the emission numbers presented in Table 11 the following observations can be made:

- On average, N₂O emission fractions varied from 0.01 - 1.8% or 0.01 - 3.3%, when normalized to influent TKN load or influent TKN load processed, respectively.
- These emission fractions were on the lower end of the range reported by previous studies, which varied between 0-15% of influent TKN load (Czepiel et al., 1995, Kampschreur et al., 2008b, Kimochi et al., 1998, Sommer et al., 1998, Sümer et al., 1995, Wicht and Beier, 1995)
- The emission fractions are mostly statistically higher (at the $\alpha=0.05$ confidence level) than currently used values of 0.00035 kg N₂O-N/kg N(non-BNR processes (Czepiel et al., 1995)).
- Emission values from the separate-stage BNR and oxidation ditch process were statistically lower, (at the $\alpha=0.05$ confidence level) and the those from the two MLE processes were statistically not dissimilar, (at the $\alpha=0.05$ confidence level) relative to the current estimates (Czepiel et al., 1995, USEPA, 2009).

The high emissions for the separate centrate treatment process are primarily because centrate streams have disproportionately low flow rates compared to their TKN concentrations.

A high degree of diurnal variability in N₂O emissions was also observed during a day at the full-scale step-feed BNR process (Figure 25) This variability could be linked to diurnal variations in influent N-loading as reported by (Ahn et al., 2009).

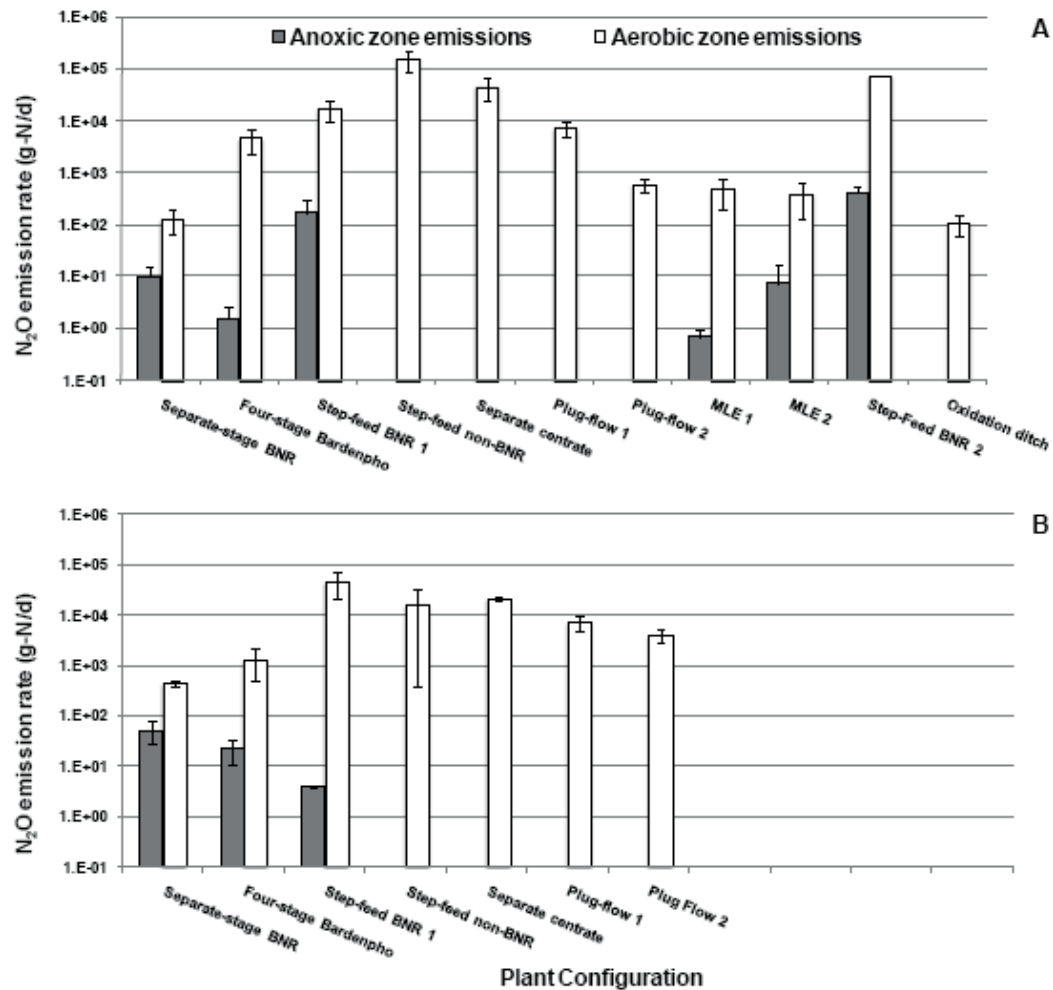
FIGURE 25 DIURNAL VARIABILITY IN GASEOUS N₂O CONCENTRATIONS MEASURED FROM AN AEROBIC ZONE OF THE FULL-SCALE STEP-FEED BNR PROCESS



6.3.2 ORIGIN OF N₂O EMISSIONS

Based on the measurements at the 12 WWTP the emission of N₂O was in general higher from aerated zones than from non-aerated zones as can be seen from Figure 26. Therefore the currently held premise that N₂O emissions from WWTPs mostly occur in the anoxic zones (USEPA, 2009) is not accurate.

FIGURE 26 N₂O EMISSIONS FROM AEROBIC AND ANOXIC ZONES IN DIFFERENT WWTPS MEASURED AT HIGH (A) AND LOW (B) TEMPERATURES. SPECIFIC TEMPERATURES DESCRIBED IN TABLE 11. STEP-FEED BNR 3 IS NOT INCLUDED SINCE THE EMISSIONS FROM THE COVERED AEROBIC AND ANOXIC ZONES COULD NOT BE DISTINCTLY MEASURED



6.3.3 PROCESS PARAMETERS OF INFLUENCE

AEROBIC ZONES

Based on multivariate regression modeling, the factors positively correlated with N₂O emissions from aerobic zones, were NH₄⁺-N, NO₂⁻-N and DO concentrations (isolated effect), and NH₄⁺-N and NO₂⁻-N concentrations (interactive effect) as can be observed from Table 12.

TABLE 12 FACTORS CORRELATING WITH N₂O EMISSION FLUXES (GN₂O-N/DAY) FROM AEROBIC ZONES

Variable	Parameter estimate	Standard error	t value	Pr > t
Intercept	6.1	0.48	13	0.00
Log(ammonia)	1.0	0.25	4.1	0.00
Log(nitrite)	0.60	0.16	3.7	0.00
Log(DO)	0.59	0.18	3.2	0.00
Log(ammonia)-log(nitrite)	0.18	0.08	2.3	0.03

The outcome of the multivariate regression modeling is also supported by the results from the Bardenpho plant as can be observed from Figure 27.

ANOXIC ZONES

Based on multivariate regression modeling, the factors positively correlated with N₂O emissions from anoxic zones, was the DO and NO₂⁻-N concentration (interactive effect) as can be observed from Table 13.

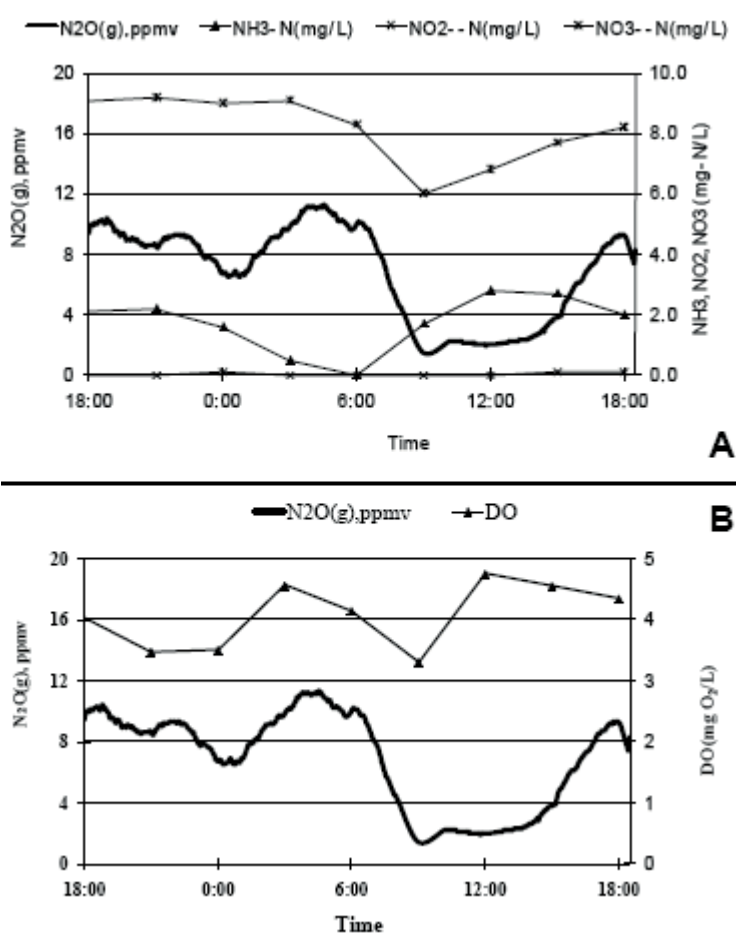
TABLE 13

FACTORS CORRELATING WITH N₂O EMISSION FLUXES (GN₂O-N/DAY) FROM ANOXIC ZONES

Variable	Parameter estimate	Standard error	t value	Pr > t
Intercept	-1.2	0.89	-1.3	0.21
Log(DO)-log(nitrite)	0.67	0.25	2.7	0.01

The regression model did not correlate temperature and N₂O fluxes in either aerobic or anoxic zones.

FIGURE 27

DIURNAL VARIABILITY IN GASEOUS N₂O CONCENTRATIONS MEASURED FROM THE FIRST AEROBI ZONE (A-B) OF THE FULL-SCALE FOUR-STAGE BARDENPHO PROCESS

TRANSITION FROM ANOXIC TO AEROBIC ZONES

Besides the parameters correlating to the emission of N₂O another factor of influence could be the transition from anoxic zones to aerobic zones. This was illustrated at different type of plants (processes) as can be observed from Figure 28 and Figure 29. From Figure 28 it can be observed that from the anoxic zone to the aerobic zone the concentration of N₂O in the gas phase increases as well as in the aqueous phase. The same increase in the gas phase was seen in the plug flow reactor (Figure 29), but here the liquid-phase N₂O concentrations at both the influent end and middle regions of the passes were statistically similar ($p=0.26$ and 1.0 , respectively for two-tailed t-test comparisons at $\alpha = 0.05$ for the two processes).

FIGURE 28

SPATIAL PROFILE OF GASEOUS N₂O CONCENTRATIONS AND TYPICAL ACTIVATED SLUDGE VARIABLES IN A STEP-FEED BNR 1 PROCESS SHOWING INDIVIDUAL SAMPLING LOCATIONS. RESULTS ARE FROM DISCRETE SAMPLING OVER A PERIOD OF 30 MINUTES AT EACH SAMPLING POINT. ARROWS INDICATE WASTEWATER FLOW. SHADED AND NON-SHADED BOXES REPRESENT NON-AERATED AND AERATED ZONES, RESPECTIVELY. GASEOUS N₂O CONCENTRATIONS ARE EXPRESSED AS AVG. \pm SD. OF 30 MEASUREMENTS






Species	Anoxic 	Aerobic 1 	Aerobic 2 
NH ₄ ⁺ (mg-N/L)	14	12 \pm 5	1.5 \pm 0.71
NO ₂ ⁻ (mg-N/L)	0.00 \pm 0.00	0.00 \pm 0.00	0.00 \pm 0.00
NO ₃ ⁻ (mg-N/L)	0.85 \pm 0.10	2.7 \pm 0.35	10. \pm 0.21
DO (mg O ₂ /L)	0.10	2.3	4.2
Aqueous N ₂ O (μ g/L)	55	190	570
Gaseous N ₂ O (ppmv)	1.5 \pm 0.14	16 \pm 0.27	23 \pm 0.67

FIGURE 29

SPATIAL PROFILE OF GASEOUS N₂O CONCENTRATIONS AND TYPICAL ACTIVATED SLUDGE VARIABLES IN A FULL-SCALE PLUG-FLOW PROCESS. RESULTS ARE FROM DIURNAL SAMPLING OVER A PERIOD OF 24 HOURS AT EACH SAMPLING POINT. ARROWS INDICATE WASTEWATER FLOW. ALL ZONES WERE AERATED

Species	Aerobic 1 	Aerobic 2 
NH ₄ ⁺ (mg-N/L)	10 \pm 0.99	6.2 \pm 2.2
NO ₂ ⁻ (mg-N/L)	0.27 \pm 0.10	0.22 \pm 0.00
NO ₃ ⁻ (mg-N/L)	1.2 \pm 0.42	2.7 \pm 1.1
DO (mg O ₂ /L)	0.80 \pm 0.47	1.8 \pm 1.0
Aqueous N ₂ O (μ g/L)	56 \pm 9.8	62 \pm 6.0
Gaseous N ₂ O (ppmv)	2.9 \pm 0.73	51 \pm 21

6.4 NITROUS OXIDE EMISSION THE NETHERLANDS

6.4.1 EMISSION OF N₂O

The N₂O emission that was measured during the four measuring campaigns at the three WWTPs is summarized in Table 14.

TABLE 14 OVERVIEW N₂O EMISSION AT THE SURVEYED WWTPS

Location	Date	N ₂ O-N emission (g N ₂ O-N/h)	NKj influent (kg/d)	Emission ¹⁾ (%)
Papendrecht	25-09-08	7.07	n.a.	-
Carrousel	26-09-08	7.69	484	0.038
	27-09-08	7.78	n.a.	-
	28-09-08	6.62	n.a.	-
	29-09-08	6.73	382	0.042
	Average	7.18	433	0.040
Kortenoord	29-05-09	7.4	839	0.021
Carrousel	30-05-09	28.9	965	0.072
	01-06-09	10.7	833	0.031
	02-06-09	24.4	927	0.063
	03-06-09	32.5	n.a.	-
	Average	20.8	891	0.048
Kralingseveer	14-10-08	346	2,348	0.35
BNR (October 2008)	15-10-08	504	2,141	0.57
	16-10-08	294	2,169	0.33
	17-10-08	256	2,642	0.23
	18-10-08	446	2,239	0.48
	19-10-08	436	2,339	0.45
	20-10-08	517	2,295	0.54
	Average	400	2,310	0.42
Kralingseveer	11-02-09	7,900	5,251	3.6
BNR (February 2009)	12-02-09	13,094	5,968	5.3
	13-02-09	11,464	2,998	9.2
	14-02-09	9,733	2,524	9.3
	15-02-09	4,528	2,359	4.6
	16-02-09	8,172	2,331	8.4
	Average	9,148	3,572	6.1
Emission factor NIR ²⁾				1

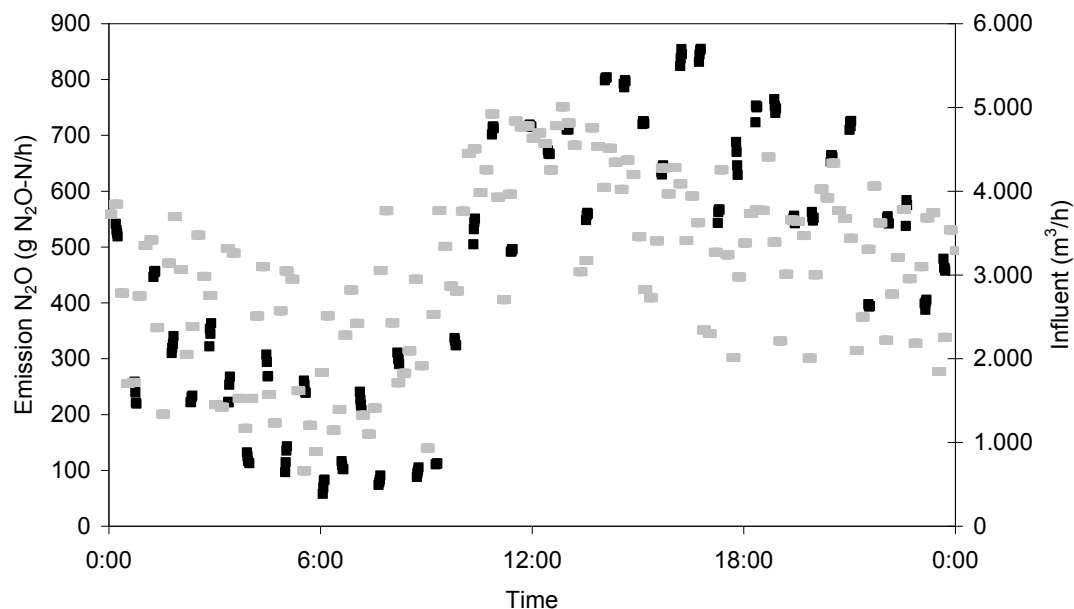
1) The uncertainty in the emission is approximately +/- 5% for Kortenoord and Papendrecht and +/-6% for Kralingseveer.

2) National Inventory Reports.

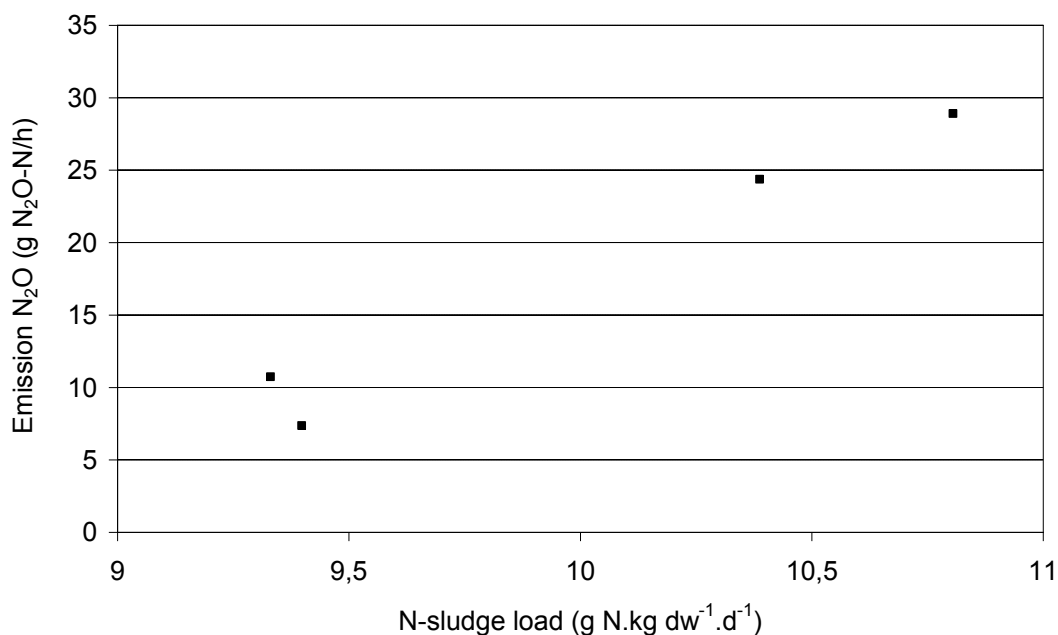
From the emission numbers presented in Table 14 the following observations can be made:

- The emission found at Papendrecht and Kortenoord were around 20 times lower than the emission factor currently used to estimate the emission of N₂O from WWTPs.
- The emission factor found at Kralingseveer in October was around two times lower than the current used emission factor.
- In February at Kralingseveer the emission was more than six times higher than the current used emission factor.
- The emission of N₂O differ at two WWTPs during one week (Kortenoord and Kralingseveer)
- The emission of N₂O differ between seasons at one WWTP (Kralingseveer)
- The emission of N₂O differ between WWTPs (Papendrecht and Kortenoord versus Kralingseveer)

Besides the observed variations in N₂O emission as mentioned above, also a strong variation was observed during a day at one WWTP. An example of this is presented in Figure 30. In Figure 30 the variation in N₂O emission is presented as observed during the measuring campaign in October at Kralingseveer.

FIGURE 30 N₂O EMISSION AND INFLUENT FLOW AT 18-10-2008 AT WWTP KRALINGSEVEER

From Figure 30 it can be observed that the emission of N₂O varies with the influent flow. This suggests that the emission of N₂O is related to the variation in sludge load. However, a linear relation between sludge load and N₂O emission could not be found at Kralingseveer, but this relation was found in Kortenoord as can be seen from Figure 31.

FIGURE 31 THE EMISSION OF N₂O AS FUNCTION OF THE N-SLUDGE LOAD AS OBSERVED DURING THE MEASURING CAMPAIGN IN KORTENOORD

6.4.2 ORIGINS OF N₂O EMISSION

One of the main objectives of the research performed in the Netherlands was to investigate to which extent WWTPs emitted N₂O and what the dynamics of this emission was. To study this, the emission of N₂O was measured in the total off gas of WWTPs that are completely covered. The total off gas originated from tanks in which both nitrification and denitrification processes took place. For this reason it was not possible to distinguish which process nitrification or denitrification was responsible for the emission of N₂O.

6.4.3 PROCESS PARAMETERS OF INFLUENCE

Based on the measuring campaigns differences in N₂O emissions were found between WWTPs and differences where found during day or in between seasons at one WWTP.

Possible process parameters that could explain these differences are summarized in Table 15.

TABLE 15

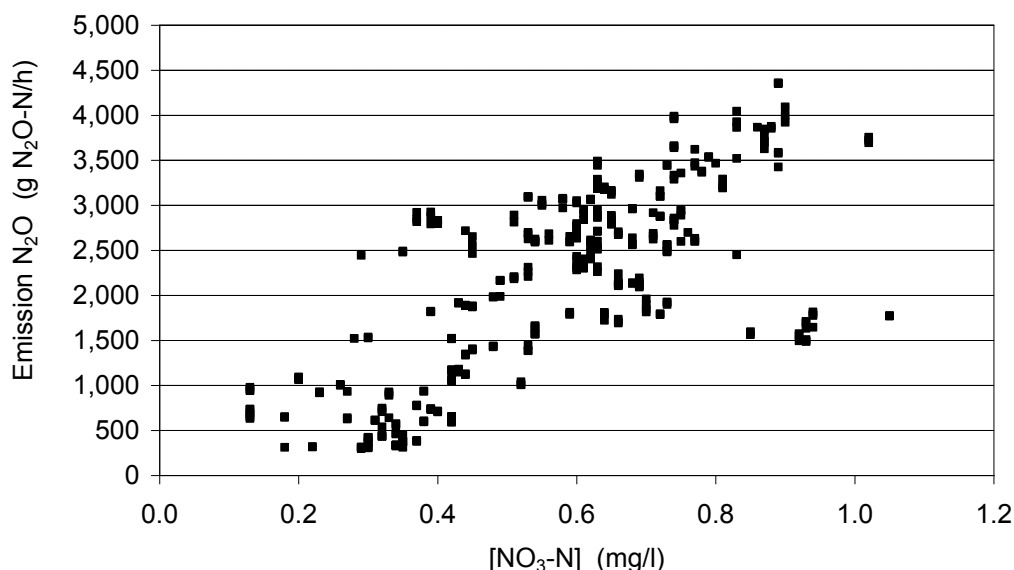
OVERVIEW PROCESS PARAMETERS THAT COULD EXPLAIN THE FOUND VARIATIONS IN N₂O EMISSION

Variation in N ₂ O emission	Possible process parameters of influence
During a week at one WWTP (p.e. Kortenoord)	Nitrogen sludge load
During seasons at one WWTP (Kralingseveer)	Temperature Rainfall Aeration intensity Nitrite Nitrogen sludge load
Between WWTPs Papendrecht/Kortenoord vs Kralingseveer	BOD / N ratio Nitrogen sludge load Type of process

The difference in N₂O emission at WWTP Kortenoord could possibly explained by the variation in nitrogen sludge loading (see Figure 31).

At the WWTP Kralingseveer the emission of N₂O was measured in October and February to study the effect of the temperature on the emission. In October the average temperature of the wastewater was 18.1 °C, in February the temperature was 9.6°C. However the effect of only the temperature could not studied, because at the start of the measuring campaign in February the WWTP suffered from heavy rainfall. At that time the WWTP had to treat wastewater flows of almost 13,000 m³/h, while the dry weather flow is normally ~5,000 m³/h. As a result of the heavy rainfall the nitrogen sludge load increased and the aeration intensity was increased to remove at least all the ammonium. Because different events (low temperature together with heavy rainfall) coincide at the same time during the measuring campaign in February it was not possible to point out one cause for the very high N₂O emission in February compared to the emission in October. What was found in February was a correlation between the N₂O emission and the nitrate concentration as can be observed from Figure 32. The found correlation between the nitrate concentration and the N₂O emission suggest that also nitrite was present. Nitrite is not a parameter that is measured at a WWTP, but based on a simple analysis during the measuring campaign in February the presence of nitrite was proven and concentrations up to ~10 mg/l were found.

FIGURE 32 THE EMISSION OF N₂O AS FUNCTION OF THE NITRATE CONCENTRATION AS OBSERVED IN THE FIRST AERATION TANK AT WWTP KRALINGSEVEER (FEBRUARY 2009)



The average N₂O emission found at WWTP Papendrecht and Kortenoord were comparable, but the emission at Kralingseveer (October) was almost ten times higher. The following differences between Papendrecht and Kortenoord on one side and Kralingseveer on the other side could possibly explain the difference in N₂O emission:

- Absence of primary clarifier at Papendrecht and Kortenoord and the presence of a primary clarifier at Kralingseveer. This resulted in a difference in the BOD/N ratio. During the measuring campaign in Papendrecht the BOD/N ratio was 2.9, in Kortenoord this ratio was 2.5 and in Kralingseveer this ratio was 1.7 in October, but was 2.7 in February.
- A lower nitrogen sludge load in Papendrecht and Kortenoord compared to Kralingseveer. During the measuring campaign in Papendrecht the nitrogen sludge load was 0.012 kgN·kg dw⁻¹·d⁻¹, in Kortenoord this was 0.010 kgN·kg dw⁻¹·d⁻¹, and in Kralingseveer this was 0.020 kgN·kg dw⁻¹·d⁻¹ in October, and in a February a nitrogen sludge load of 0.053 kgN·kg dw⁻¹·d⁻¹ was observed.
- Process configuration. In Papendrecht and Kortenoord the removal of nitrogen occurs in a carrousel, while in Kralingseveer nitrogen is removed in first a plug flow reactor followed by two carrouseles.

6.5 METHANE EMISSION AUSTRALIA

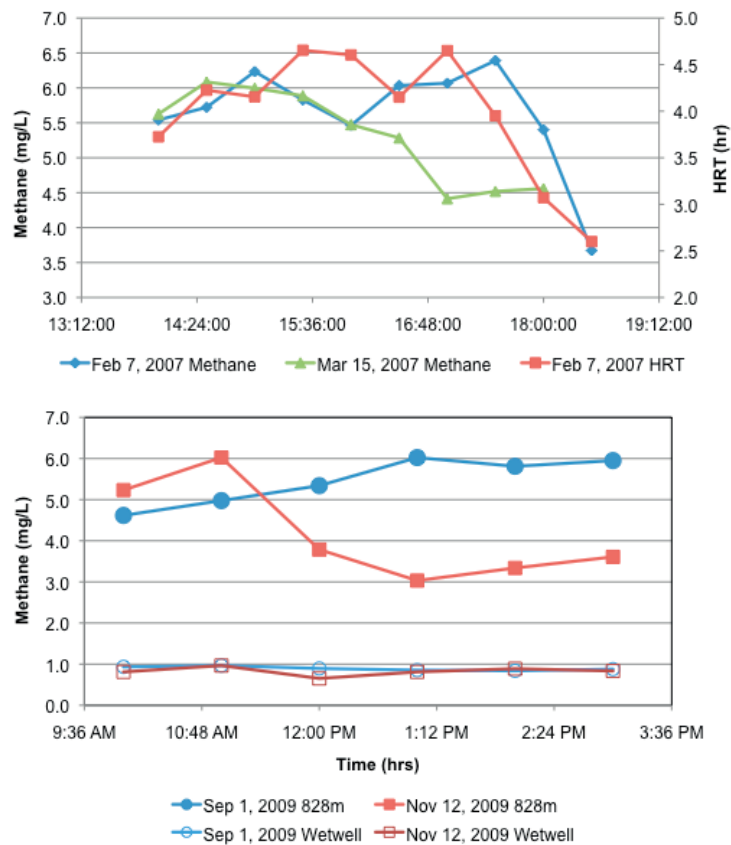
6.5.1 LIQUID PHASE DATA

RISING MAIN UC09

Rising main UC09 is a single line receiving domestic wastewater. It has a length of 1080 m and a pipe diameter of 150 mm (Area to Volume ratio: $A/V = 2/0.075 = 26.7 \text{ m}^{-1}$). The average daily flow was approximately 200 m³/d at the time of the study, giving rising to an average HRT of 2.5 hr. However, HRT varied between 1.5 to 6.0 hours caused by diurnal variations of the wastewater flow. The feed pump was operated intermittently with an interval ranging between below 30 minutes to above 2 hours. Each pumping event lasted for 1-3 minutes depending on the time of the day. The average flow velocity during a pumping event was estimated to be 1.55 m/s. The examination of a removable section of the pipe revealed that no sediments were present in this pipe.

Figure 33 shows the dissolved CH₄ concentrations measured at both the pumping station and 828m downstream from the pumping station during daytime (10:00 a.m. – 6:00 p.m.) over a time span of two years.

FIGURE 33 METHANE DATA MEASURED AT UC09. PART OF THE DATA WAS PREVIOUSLY PUBLISHED IN GUIASOLA ET AL. (2008). THE WASTEWATER TEMPERATURE WAS 27.7°C, 26.9°C AND 24.6°C ON 07/02/2007, 15/03/2007 AND 01/09/2009, RESPECTIVELY



In one case, the HRT of the wastewater samples taken at 828 m was calculated and also shown in the figure. The HRT of a wastewater sample, defined as the time of this sample spent in the rising main at the time of sampling, was calculated from the operational data of the pump feeding the given rising main (Sharma et al., 2008). The volume of wastewater pumped into the pipe during each pump run was calculated from the physical dimensions of the wet well and the measured stop/start water levels. Based on this estimated wastewater volume and the dimensions of the pipe, the number of pump runs required for a wastewater “slug” to travel from the pipe inlet to the sampling location was calculated. The HRT of the wastewater sample collected was then calculated from the on-line pump operational data recorded by the on-line supervisory control and data acquisition (SCADA) system.

In addition to CH₄ concentrations, several other physico-chemical parameters of the samples collected such as temperature, total and soluble COD, volatile fatty acids (VFAs), inorganic sulphur species (sulphate, sulphide, thiosulphate and sulphite) and pH were also measured.

Methane concentration at the pumping station was always approximately 1 mg/l. In comparison, CH₄ concentration at 828m was between 3 – 6.5 mg/l with an average of approximately 5.0 mg/l, indicating an average production of 4.0 mg/l. The average CH₄ production rate was 1.3 mg·l⁻¹·hr⁻¹, or 1.2 g·m⁻²·d⁻¹ by taking the A/V ratio of 26.7 m⁻¹ into consideration.

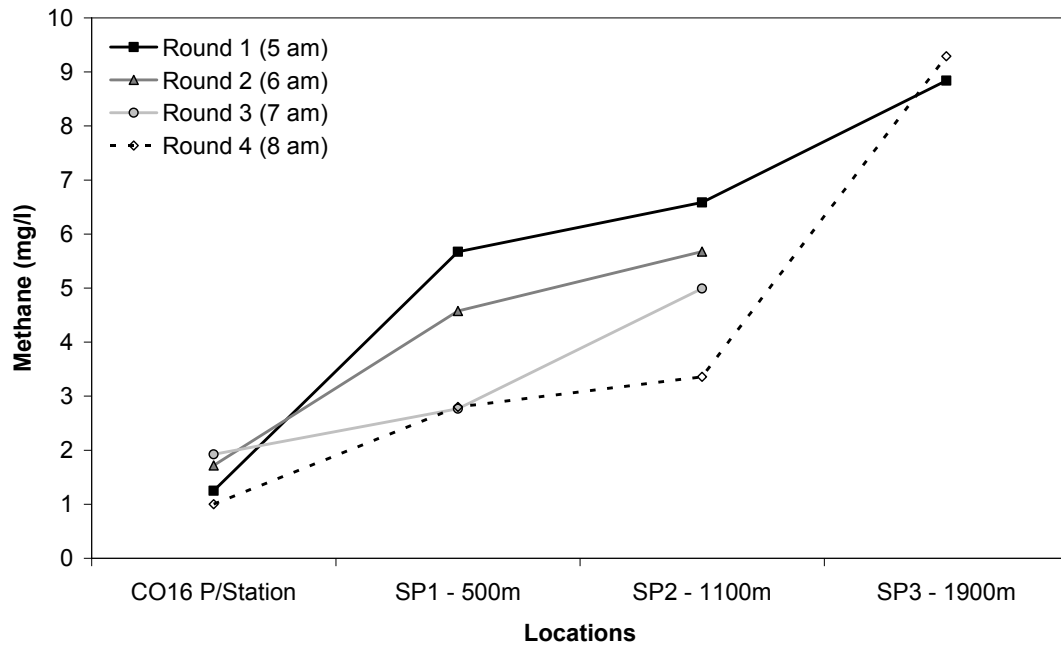
RISING MAIN C016

A second sampling site is C016, which were sampled at four locations, namely the pumping station and 500m, 1100m and 1900m downstream of the pumping station (Figure 13). Similar to UC09, the C016 rising main also receives domestic wastewater. It has an internal pipe diameter of 300 mm, giving an A/V ratio of 13.3 m⁻¹. The average daily wastewater flow was approximately 700 m³. Similar to UC09, the pumping station was operated intermittently with 30 - 40 pumping events per day, each lasting for 4 – 6 min in duration. During each pumping event, the calculated flow velocity was approximately 0.90 – 1.01 m/s.

In one sampling campaign, samples from the four locations were collected hourly between 5:00 a.m. and 8:00 a.m. to cover a wide range of HRT (0 to 8.7 hr), as the samples captured included wastewater discharged over night and hence with a relatively long retention time in the sewer line, and also relatively fresh sewage discharged in the morning. The CH₄ data are plotted against the sampling locations in Figure 34 (Foley et al., 2009). More detailed data can be found in Foley et al. (2009).

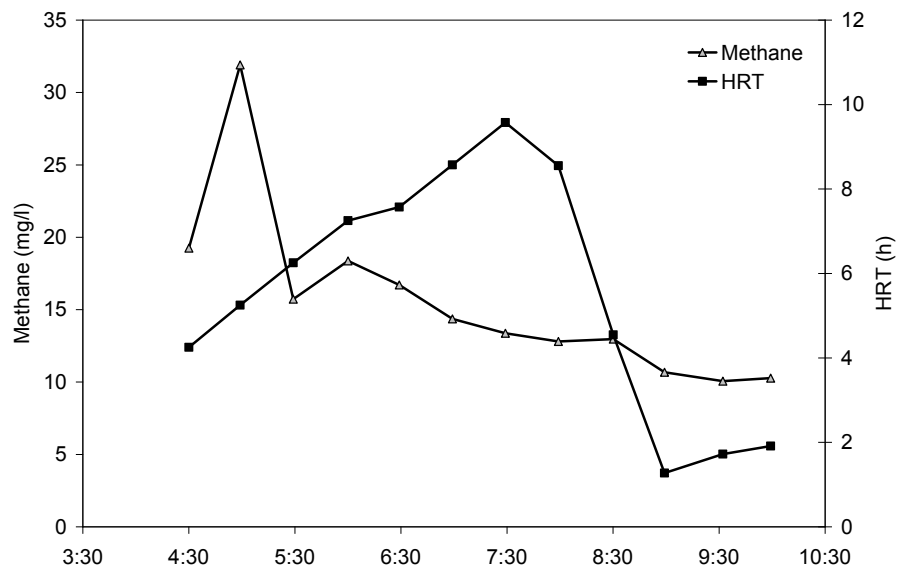
Similar to UC09, CH₄ concentration in the pumping station was between 1-2 mg/l. The concentration increased further downstream, reaching approximately 9.0 mg/l. Methane concentration varied considerably with time at the 500m and 1100m locations between 5am and 8am, likely due to the variation of HRT of the wastewater samples collected. The wastewater HRT at 500m reduced from 5.4 hr at 5:00 a.m. to just below 1 hr at 7 – 8:00 a.m. Similar, the wastewater HRT reduced from 7.3 hr at 4:00 – 5:00 a.m. to 1.5 hr at 7:00 – 8:00 a.m. By taking all results together, the average CH₄ production rate was calculated as 0.7 mg·l⁻¹·hr⁻¹ or 1.3 g·m⁻²·d⁻¹. The latter is very similar to the value observed at UC09.

FIGURE 34 METHANE DATA MEASURED AT FOUR LOCATIONS ALONG C016 (FOLEY ET AL., 2009). THE AVERAGE WASTEWATER TEMPERATURE WAS 23.5°C



In another sampling campaign, the 1100m location was sampled half-hourly during 4:15 a.m. and 10:00 a.m. The results are presented in Figure 35.

FIGURE 35 METHANE DATA MEASURED AT C016 AT 1100M IN ANOTHER CAMPAIGN. THE AVERAGE WASTEWATER TEMPERATURE WAS 22.5 °C



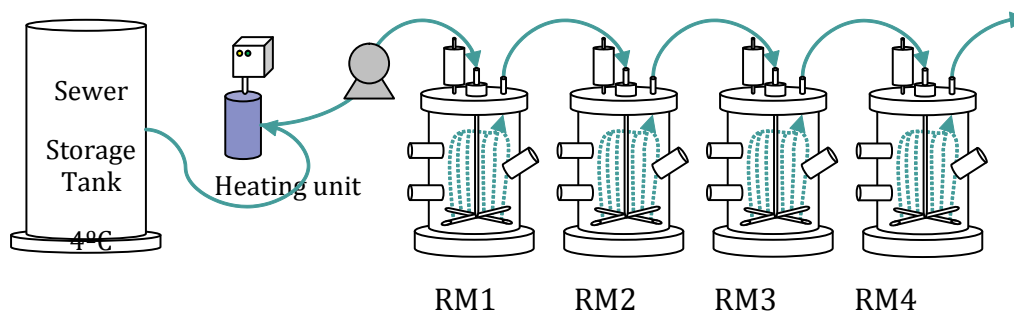
The CH₄ concentrations measured this round were approximately three times those presented in Figure 34. This difference cannot be fully explained by the differences in HRT (4 – 9.5 hr in this case in comparison with 1.5 – 7.3 hr in Foley et al., 2009). The average CH₄ production rate was calculated as 3.1 mg·l⁻¹·hr⁻¹ or 5.5 g·m⁻²·d⁻¹, three times higher than the values obtained in the previous round or for UC09. The average wastewater temperature was 22.5 °C, which was similar to that in the previous round. The reason for the much higher CH₄ production in this round is not known. This could be related to the COD (Chemical Oxygen Demand) concentration, which unfortunately was not measured during the measurement campaigns

LABORATORY-SCALE SEWER SYSTEMS

To investigate CH₄ production in sewers under controlled conditions, reactor-based laboratory scale systems were set up at The University of Queensland (Figure 36).

FIGURE 36

A LABORATORY-SCALE SEWER SYSTEM AT THE UNIVERSITY OF QUEENSLAND



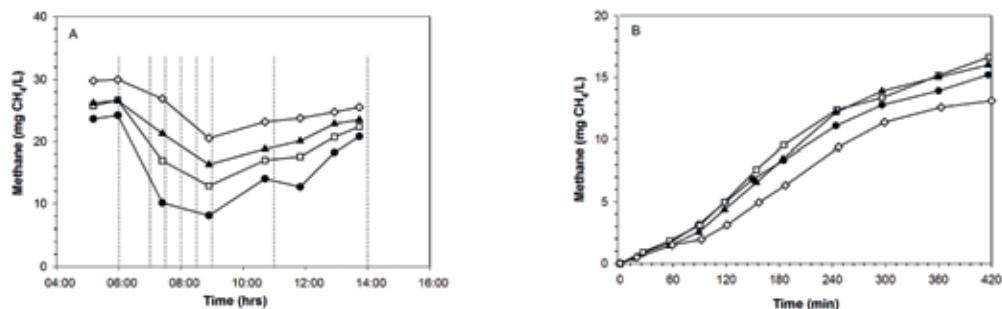
Each system consisted of four air-tight reactors in series (named RM1 to RM4), each with a volume of 0.75 L. The diameter of the reactors was 80 mm and the height was 150 mm. The A/V ratio was calculated to be 56.7 m⁻¹ by considering the biofilm areas both on the wall and on the top of the reactor (there was no visible biofilm growth on the bottom of the reactor due to the use of a magnetic stirrer, see below). Biofilm carriers were placed in each reactor to enable the collection of intact biofilms for micro-scale analysis. This increased the biofilm surface area by 25%, and the effective A/V ratio was thus increased to 70.8 m⁻¹. The system was intermittently fed with real sewage that had been screened and settled using a peristaltic pump following a typical operating pattern observed at UC09 in 2003. The feed pattern consisted of sixteen pumping events per day, each delivering 0.75 l (equivalent to the volume of one reactor) wastewater into the system. The average HRT in the system was 6 hrs with minimum and maximum HRT being 2 and 10 hrs, respectively. The system was operated under quiescent conditions except during pumping events, which resulted in turbulence in the reactors due to flow. The reactors were also stirred via magnetic stirrers during pumping events to minimize the accumulation of sediments.

Fresh sewage was collected weekly at the Robertson Park Pump Station, Indooroopilly (Brisbane), which primarily collects domestic wastewater from the local area, and thus contains a low amount of sulphide (≤ 1 mg S/l) and CH₄ (~ 1 mg/l). The fresh sewage contains on average 15 mg SO₄-S/l and 200 mg COD/l of soluble COD, of which the Volatile Fatty Acid (VFA) content was approximately 50 mg COD/l as acetate and 15 mg COD/l as propionate. Sewage was stored at 4°C to minimize biological transformations and warmed to 21°C through a heat exchanger and water bath prior to entering the system. The sewage temperature remained stable as the reactor system was located in a temperature controlled lab (21 ± 1°C).

After several months of operation, anaerobic biofilm was fully developed in the laboratory system, leading to pseudo-steady state conditions. The system was monitored with routine sampling of wastewater from all reactors. Figure 37A shows typical CH₄ profiles in all reactors during daytime.

FIGURE 37

A. SPATIAL AND TEMPORAL VARIATIONS OF CH₄ CONCENTRATIONS IN THE LABORATORY-SCALE SEWER SYSTEMS; B. METHANE PROFILES OBTAINED IN BATCH TESTS WITH THE FOUR REACTORS. RM1 (●), RM2 (□), RM3 (▲) AND RM4 (◇). TIME 0 IS OAM. THE VERTICAL DASHED LINES IN A CORRESPONDED TO PUMPING EVENTS. WASTEWATER TEMPERATURE DURING THE TEST WAS APPROXIMATELY 21 °C



The results confirmed that a substantial amount of CH₄ can be produced in anaerobic sewer systems. The measured CH₄ concentrations were between 8 - 30 mg/l, generally higher than the values measured in real sewers. This was likely due to the higher A/V ratio of the lab-system (70.8 m⁻¹ vs. 13.3 m⁻¹ of CO16 and 26.7 m⁻¹ of UC09). As will be further discussed later, however, the CH₄ production rate per area of sewer biofilm (calculated to be 1.0 – 1.2 g CH₄·m⁻²·d⁻¹) is very similar to the values determined for UC09 and CO16.

Batch tests were performed with all the four reactors to assess the methanogenic activities of the biofilms. The continuous operation of the system was stopped and the experimental reactors were temporarily isolated from the system. At the start of each batch test, the selected reactor was carefully emptied and drained to remove any sediment, and re-filled with fresh sewage, taking care to minimize oxygen entrainment during filling. The reactor was magnetically stirred during the batch experiments. As an average, two batch tests per month were conducted in each reactor.

Each batch test typically lasted around eight hours. Methane, VFAs, COD and soluble sulphur species (sulphate, sulphite, thiosulphate and sulphide) were measured regularly. A sample frequency of around 30 min was maintained for at least the first three hours of the test, while the last 5 hours were usually sampled on an hourly basis. Figure 37B shows the typical CH₄ profiles obtained during the batch tests. Methane production was approximately linear in all reactors in the first 5 hours. The slowing-down after 5 hours was likely due to the decrease of easily fermentable COD (Guisasola et al., 2008). The relatively slow rates in the first 1.5 hours were not expected, which could be due to the contact of anaerobic biofilm with oxygen during the draining process. Very similar CH₄ production rates were observed for the first three reactors. The fourth reactor had a slightly lower rate. The average CH₄ production rate was 2.9 – 3.7 mg·l⁻¹·hr⁻¹ (determined using the linear part of the curves presented in Figure 37B, i.e. between 90 – 240 minutes), or equivalently 1.0 – 1.2 g·m⁻²·d⁻¹. These values are very similar to those observed at UC09 and CO16.

6.5.2 GAS PHASE DATA

While it is a routine process for sewer workers to measure %LEL levels in confined spaces, CH₄ concentration in sewer air has not been widely reported in literature. In the last few years, South East Water Limited (SEWL) investigated CH₄ production in one of its sewer mains (Ibrahim, 2010). The main is 1.8 km in length. The pipe material is concrete and diameter starts at 150mm and goes to 525 mm. The main receives wastewater from a number of industrial dischargers and a small amount of domestic sewage towards the end of main. South East Water Ltd. detected high %LEL levels and low O₂ readings during scheduled cleaning of

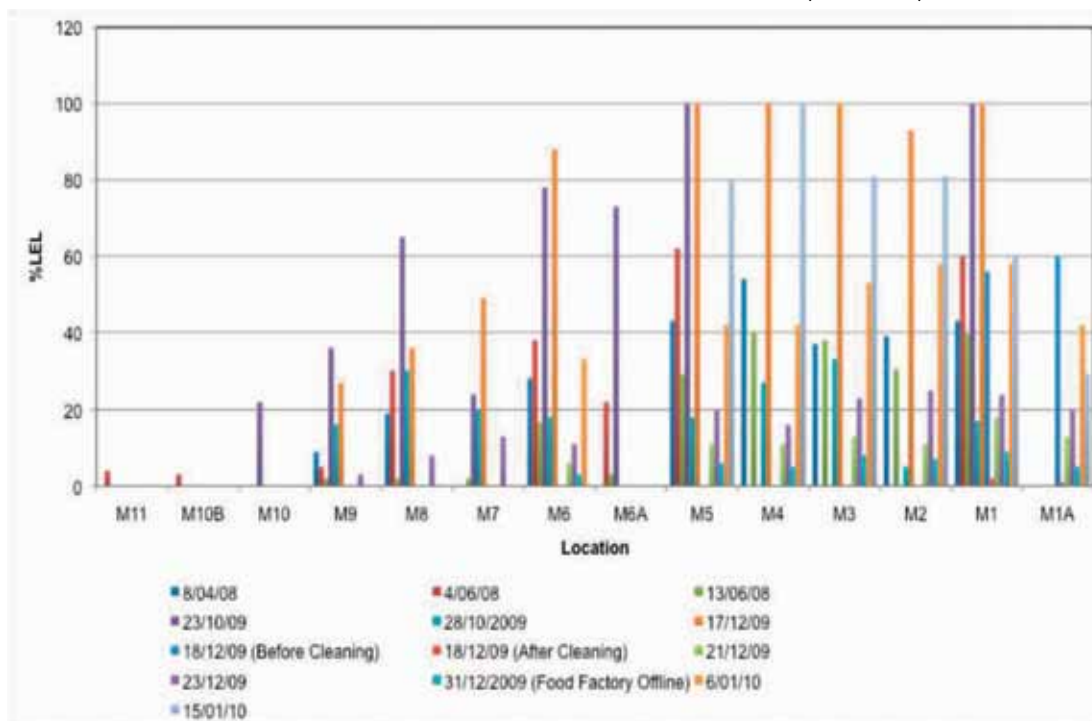
this main in January 2006, which triggered a closer monitoring of the %LEL levels and CH₄ concentrations in the manholes along the sewer line. Figure 38 shows the %LEL levels at various manholes over a period of 1.5 years.

Very high %LEL levels were repeatedly detected in this sewer, particularly at downstream locations. In several occasions, %LEL was above 100%, which implied a risk of explosion. Methane was confirmed to be responsible for the high %LEL readings through bag sampling and gas composition analysis with GC. In the two samples taken from Manhole M6 at 2am and 2pm, CH₄ concentrations were measured as 1.7% (34% LEL) and 0.6% (12% LEL), respectively.

The fact that %LEL was relatively low at upstream locations (M11 and M10B) suggests that CH₄ emitted at downstream locations was formed in this gravity section rather than as result of CH₄ formation in a pressurized section located further upstream, as otherwise high emissions would have been observed at M11 and M10B as well.

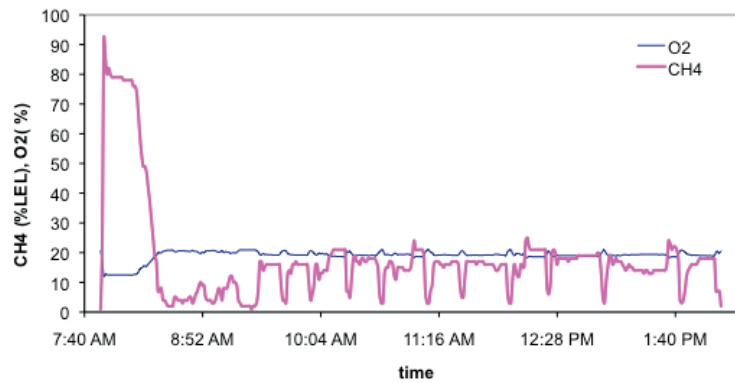
It should be noted that this system may not be representative to general sewer networks due to the industrial origin of the wastewater transported. Indeed, as will be further discussed, SEWL was able to identify the cause for the high-level CH₄ formation, which is linked to a particular food processing wastewater that contained a high-level of biodegradable COD. This will be further discussed later.

FIGURE 38 GAS PHASE %LEL IN MANHOLES ALONG A MAIN RECEIVING PREDOMINANTLY INDUSTRIAL WASTEWATERS (IBRAHIM, 2010)



However, high-levels of CH₄ can also be present in sewer networks that primarily transport domestic wastewater. Indeed, high concentrations of CH₄ would be expected at discharging points of rising mains. Figure 39 shows that the dynamic CH₄ and oxygen profiles measured at the discharging well of CO16, where high-levels of dissolved CH₄ were detected (referring to section 6.5.1).

FIGURE 39 GAS PHASE CH₄ %LEL AND OXYGEN CONCENTRATION AT A RISING MAIN DISCHARGE POINT



In the early morning, a %LEL level of 80 – 90% was measured, which was accompanied by a low oxygen level of 12 – 13%. The very high level of CH₄ can be explained by the discharge of septic wastewater that had stayed in the rising main for many hours over night. The %LEL levels varied between 2 – 22% during the remaining time of the monitoring period. The dynamics was likely caused by the intermittent discharge of CH₄-containing wastewater from the rising main. Unfortunately, the monitoring trial lasted for a few hours only. More on-line data of this nature should be collected in the future.

6.5.3 MODELLING

AN EMPIRICAL MODEL

Both field data and simulation studies revealed that, for a given wastewater composition, the key factors determining CH₄ formation in a rising main sewer are the HRT of wastewater, and the A/V ratio of the pipe. Based on this observation, Foley et al. (2009) proposed the following empirical equation for estimating CH₄ production in a rising main pipe:

$$C_{CH_4,end} = C_{CH_4,0} + r_{CH_4,A} * \frac{A}{V} * HRT \quad (1)$$

where $C_{CH_4,end}$ and $C_{CH_4,0}$ are CH₄ concentrations (mg/l or g/m³) at the end and at the beginning of the pipe, respectively; $A/V = 2/r$ is the A/V ratio (m⁻¹) of the pipe with r (m) being the pipe radius; HRT is the hydraulic retention time (hr) of wastewater in the pipe; $r_{CH_4,A}$ is the CH₄ production rate of sewer biofilms (g/m²).

Field measurements carried out in Australia so far have consistently shown that $C_{CH_4,0}$ has a value of 1-2 g/m³ in wet wells collecting domestic sewage.

With the field data measured, $r_{CH_4,A}$ was empirically determined to be 0.0524 g·m⁻²·hr⁻¹ in Foley et al. (2009), which was shown to agree well with the Guisasola et al. (2008) model (Foley et al., 2009). However, the value of this parameter is expected to be largely affected by, among other factors, wastewater composition and temperature and may need calibration in each case.

This simple predictive model provides a means for water authorities to estimate CH₄ formation from their pressurized sewerage systems.

LIMITATIONS OF CURRENT MODELS

The modelling work to date has focused on rising main sewers. In gravity sewers, both CH₄ formation and oxidation are expected to occur due to the presence of surface aeration of wastewater. However, to date there has been no study on CH₄ transformation in gravity sewers. In particular, the contribution of sewer sediments to CH₄ formation is currently unknown. This should be addressed in future research.

While there has been no study reported to date dedicated to the gas-liquid mass transfer of CH₄ in sewers, its modelling should be relatively straightforward given the large amount of work undertaken to model the transfer of oxygen and hydrogen sulphide in sewer systems (Hvitved-Jacobsen, 2002).

6.5.4 IMPACT OF TRADE WASTE

Many types of trade wastes contain a high-level of easily biodegradable COD, which may increase CH₄ production in sewers. Indeed a major contributor to the high-levels of CH₄ presented in Figure 38 was confirmed to be the wastewater discharged by a food factory. The trade wastewater contained total COD (TCOD) and soluble COD (SCOD) at 26,500 and 30,000 mg/l, respectively, elevating the TCOD and SCOD concentrations from 1,750 mg/l and 1,050 mg/l before the trade waste discharge to 14,500 mg/l and 12,000 mg/l, respectively, after the discharge. Methane generation testing was conducted to determine if this particular trade waste was responsible for the high-level of CH₄ measured. Testing was performed with mini-digesters to measure the overnight change in gas pressure in four samples. Samples were taken from the trade waste, upstream and downstream of the trade waste discharge point, as well as at the end of the sewer line. The samples were left overnight at 21°C and the gas pressure measured approximately after 18 hours. The upstream sample showed the lowest reading in pressure (see Table 16). A dramatic increase CH₄ production by the downstream samples, along with the high CH₄ production potential of the raw trade waste, suggests that the trade waste was likely a major source of CH₄. This was independently confirmed by the fact that CH₄ concentration in sewer air decreased dramatically when the factory was closed during a holiday period.

TABLE 16 METHANE PRODUCTION DURING 18 HR ANAEROBIC INCUBATION OF WASTEWATER SAMPLES COLLECTED AT VARIOUS LOCATIONS (IBRAHIM, 2010)

Wastewater sample collection location	Final pressure (mm Hg)	Soluble COD (mg/l)	Total COD (mg/l)
Upstream of the trade waste discharge point	22	1,050	1,750
Downstream of the trade waste discharge point	72	12,000	14,500
End of the sewer line	85	9,400	11,000
Trade waste	120	26,500	31,000

An in-depth study of the impact of brewery wastewater discharge on sulphide and CH₄ production in a sewer was conducted in Sudarjanto et al. (2010a) using laboratory sewer systems. Experiments were carried out on two laboratory scale sewer reactors, one used as an experimental and the other as a control. Each reactor had a volume of 1 l, and was fed with wastewater four times a day. During each pumping event, 1 l of wastewater was pumped into each reactor, resulting in a hydraulic retention time (HRT) of 6 hours.

As summarized in Table 17, both reactors were fed with domestic sewage for six months to enable the development of stable anaerobic sewer biofilms (baseline period). While the feed to the control reactor remained unchanged, the experimental reactor was fed with a mixture of brewery and domestic wastewater at two different proportions at later stages (10% v/v for

three months followed by 25% v/v for another 1.5 months). The composition of the sewage and brewery wastewater is shown in Table 18. Batch tests were conducted to measure the CH₄ production rates of the two reactors during the three phases.

TABLE 17 DETAILS OF THE OPERATIONAL PERIODS (SUDARJANTO ET AL., 2010A)

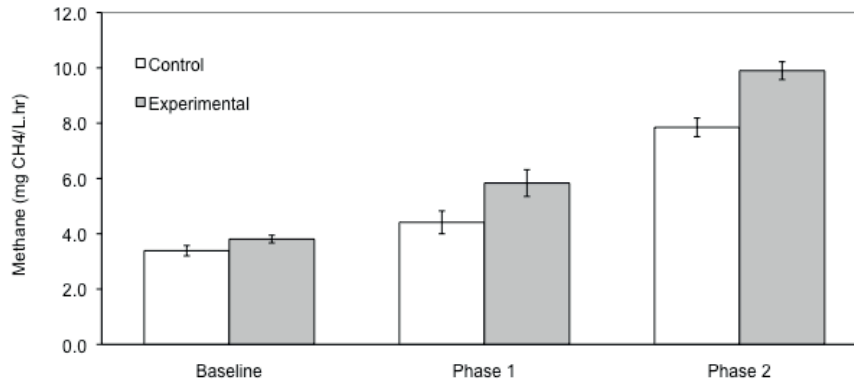
Phase	Duration	Composition of feed	
		Experimental reactor	Control reactor
1	6 months	100% domestic wastewater	100% domestic wastewater
2	3 months	90% domestic + 10% brewery wastewater	100% domestic wastewater
3	1.5 months	75% domestic + 25% brewery wastewater	100% domestic wastewater

TABLE 18 COMPOSITION OF SEWAGE AND BREWERY WASTEWATER USED IN THE STUDY (SUDARJANTO ET AL., 2010A)

Parameter	Unit	Composition of feed	
		Raw sewage	Brewery wastewater
TCOD	mg COD/L	270-485	5,000-7,000
SCOD	mg COD/L	170-310	3,000-6,000
VFA	mg COD/L	50-120	250-500
pH	-	7.2 ± 0.2	4.0 ± 0.6

As shown in Figure 40, 10% v/v discharge of brewery wastewater increased the CH₄ production rate of the sewer biofilm by 30%.

FIGURE 40 METHANE PRODUCTION RATES BY THE ANAEROBIC SEWER BIOFILMS IN THE CONTROL AND EXPERIMENTAL REACTORS DURING THE THREE PHASES OF THE STUDY (SUDARJANTO ET AL., 2010)



When the brewery wastewater fraction was increased to 25% v/v, the CH₄ production rate in the experimental reactor remained to be 30% higher than that in the control reactor. The results confirmed that the discharge of brewery wastewater into sewers significantly increase CH₄ production by sewer biofilms. It is also interesting to observe that the ratio of CH₄ production rates between the two reactors did not increase when the brewery wastewater fraction was increased from 10% to 25%, with reasons yet to be identified. It may be that CH₄ production does not increase further once a critical SCOD is reached in the wastewater. This is understandable considering the fact that biological reactions in sewers are primarily catalyzed by sewer biofilms (including sediments) (Gutierrez et al., 2009), and therefore the reaction rate may be limited by the biofilm surface area. A further contributor could be the low pH in the experimental reactor during brewery wastewater addition.

The average pH in the control reactor was 7.0, which was also the pH in the experimental reactor in the baseline period. With 10% and 25% addition of the brewery wastewater, pH in the experimental reactor decreased to 6.6 and 6.0, respectively. The low pH could have partially reduced the activities of methanogens.

It should be noted that different types of trade wastes may have different effects on CH₄ formation in sewers. In a recent study, Sudarjanto et al. (2010b) found that the discharge of a dairy wastewater, which also contains high-levels of SCOD and TCOD, at a ratio of 10%v/v did not cause any increase in CH₄ production in a laboratory sewer system. Indeed, both sulphide and CH₄ production was observed to decrease slightly. The results suggest that each trade waste should be assessed separately. Nevertheless, it is conceivable that trade wastes that contain high-levels of carbohydrates will likely enhance CH₄ formation in sewers.

6.5.5 MITIGATION

The mitigation of CH₄ emission from sewers is important for the water industry to minimize the carbon footprint of its operation. This would be best achieved through reducing CH₄ production by suppressing the growth of methanogens in sewer biofilms. The mineralization of CH₄ after its production is an alternative option, but may be difficult to achieve in the sewer environment. The biological oxidation of CH₄ is a slow process in general.

Chemical dosage to sewage is commonly used for the control of sulphide formation in sewers, or its transfer from wastewater to sewer air. The effect of several of these chemicals including nitrite, iron salts and magnesium hydroxide on CH₄ formation by sewer biofilms has recently been investigated (Mohanakrishnan et al., 2008; Gutierrez et al., 2009; Zhang et al., 2009; Jiang et al., 2010a). These chemicals substantially reduce or even completely suppress methanogenic activities. Once inhibited or suppressed, it takes weeks or even months for methanogens to fully recover (Jiang et al., 2010a). These results indicate that CH₄ emission can be mitigated in conjunction with sulphide emission. Both CH₄ and sulphide control should be considered when optimizing the use of these chemicals. Future research should also focus on the development of more cost-effective mitigation strategies, perhaps dedicated to CH₄ control.

ELEVATION OF PH

Elevation of pH through the addition of magnesium hydroxide (Mg(OH)₂) is commonly used as a means for reducing H₂S emission from wastewater to sewer air. When pH is lifted from a neutral level, which is typical in sewage, to 8.5 – 9.0, the molecular hydrogen sulphide (H₂S) fraction of total dissolved sulphide is reduced, and as a result its transfer from the liquid to the gas phase is reduced.

Gutierrez et al. (2009) investigated the impact of pH elevation on the microbial activities of anaerobic sewer biofilms, related to sulphide and CH₄ production as well as fermentation.

Experiments were carried out on laboratory scale sewer systems consisting of both an experimental and a control reactor. Each reactor, with a volume of 1 l, was fed with domestic wastewater every six hours.

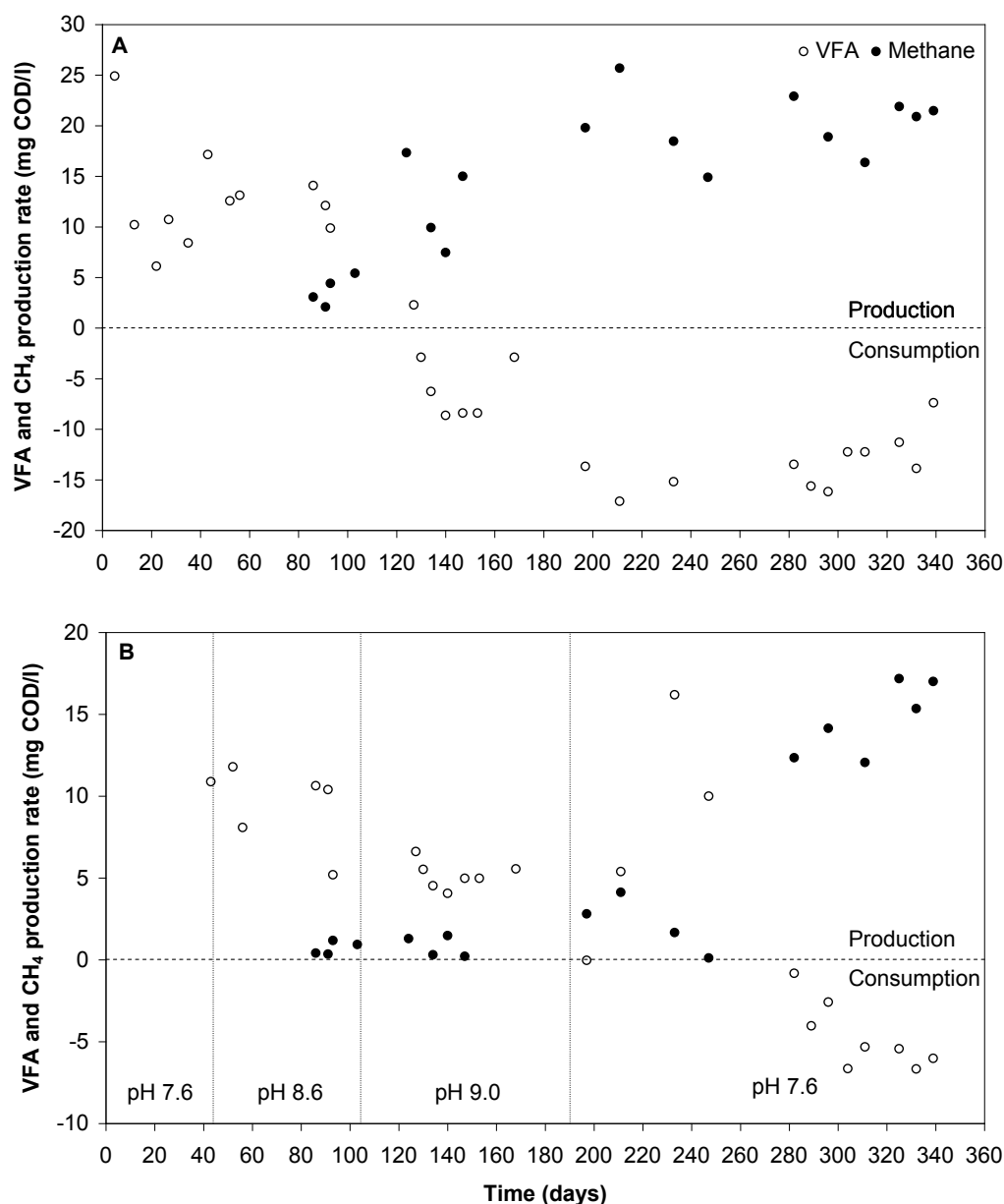
During each pumping event, 1 l of wastewater was pumped into each reactor, resulting in a hydraulic retention time (HRT) of 6 hours. The 12-month experimental study was divided into four phases with conditions summarized in Table 19.

TABLE 19 EXPERIMENTAL CONDITIONS APPLIED DURING THE ENTIRE COURSE OF THE PH STUDY (GUTIERREZ ET AL., 2009)

Operational phases	Length (days)	Control Reactor	Experimental Reactor
1	0 - 40	No pH control 7.6±0.1	No pH control 7.6±0.1
2	51 - 110	No pH control 7.6±0.1	pH adjusted to 8.6±0.1 with 88 ml 0.05M NaOH in each pumping event
3	111 - 170	No pH control 7.6±0.1	pH adjusted to 9.0±0.1 with 120 ml 0.05M NaOH in each pumping event
4	171 - 332	No pH control 7.6±0.1	No pH control 7.6±0.1

Compared to those in a control reactor without pH control (pH 7.6±0.1), the SRB activity was reduced by 30% and 50%, respectively, at pH 8.6 and pH 9.0. The results showed that pH elevation not only reduced the H₂S transfer but also its production by sewer biofilms. When normal pH was resumed, it took approximately two months for the SRB activity to fully recover. More details can be found in Gutierrez et al. (2009).

FIGURE 41 EVOLUTION OF THE VFA (○) AND CH₄ PRODUCTION RATES (●) IN THE CONTROL (A) AND EXPERIMENTAL (B) REACTORS. NOTE THAT CH₄ PRODUCTION RATES HAVE A UNIT OF MG COD · L⁻¹ · HR⁻¹ IN THIS FIGURE, RATHER THAN MG CH₄ · L⁻¹ · HR⁻¹, FOR DIRECT COMPARISON WITH THE VFA PRODUCTION/CONSUMPTION RATES. 1 MG CH₄ = 4 MG COD



Methanogenic activities developed in the control reactor in three months after the reactor start-up, while no significant methanogenic activities were detected in the experimental reactor until normal pH was resumed (Figure 40). The results suggest that elevated pH at 8.6 - 9.0 suppressed the growth of methanogens.

These experimental findings are significant for the optimal use of alkali addition to sewers for the control of H₂S and CH₄ emissions. A model-based study performed in Gutierrez et al. (2009) showed that, by adding the alkali at the beginning rather than towards the end of a rising main, substantial savings in chemicals can be achieved while achieving the same level of sulphide emission control, and complete CH₄ emission control.

Elevated pH conditions also reduced the activity of fermentative bacteria (FB) in the reactors. Prior to the development of the methanogenic activity, a stable VFA production rate of 11.5 ± 1.2 mg VFA-COD·l⁻¹·hr⁻¹ was observed in the control and experimental reactors. The FB activity decreased to 5.1 ± 0.4 mg VFA-COD·l⁻¹·hr⁻¹ in the experimental reactor under pH 9.0 (with negligible CH₄ formation). This indicates that the fermentation activity was reduced by 54% at pH 9.0 in comparison to pH 7.6.

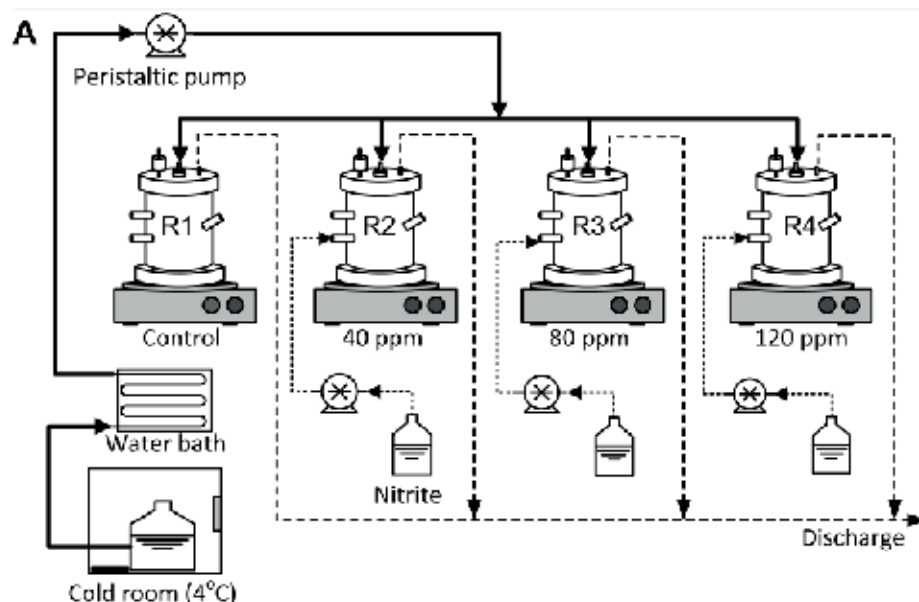
NITRITE ADDITION

Mohanakrishnan et al. (2008) and Jiang et al. (2010a) investigated the impact of nitrite addition on the sulphate-reducing and methanogenic activities of anaerobic sewer biofilms. It was found that intermittent addition of nitrite could suppress CH₄ production for weeks to months.

The laboratory systems used in Jiang et al. (2010a) are shown in Figure 42, which consisted of four air-tight reactors, namely R1 to R4, each with a volume of 0.75 l. The reactors were fed with sewage through a peristaltic pump every 6 hours, a typical sewage hydraulic retention time in sewers (Hvitved-Jacobsen, 2002).

Every feed pumping event lasted for 2 minutes, delivering one reactor volume (0.75 l) of sewage into each reactor.

FIGURE 42 SCHEMATIC OF THE LABORATORY-SCALE RISING MAIN SEWER REACTORS. R1 DID NOT RECEIVE NITRITE, AND SERVED AS A CONTROL. R2, R3 AND R4 RECEIVED NITRITE AT 40, 80 AND 120 MGN PER L OF WASTEWATER FED, FOR A PERIOD OF 24 DAYS, WHICH WAS FOLLOWED BY A 2.5 MONTH RECOVERY PERIOD



The experiments were conducted in three consecutive phases, namely the stabilization, dosing, and recovery phases. Reactors were operated without nitrite dosing to achieve similar sulphide and CH₄ production activities during the stabilization phase. During the dosing phase, R2-R4 received nitrite, while R1 was used as the control reactor (no nitrite dosage). Reactors R2-R4 were injected with nitrite to reach concentrations of 40, 80, and 120 mg-N/l of wastewater, respectively.

After the 24-day dosing phase, nitrite dosing to R2 - R4 was stopped and the reactors were allowed to recover for two months (Recovery phase).

Methane production in dosed reactors was reduced to negligible levels within 2 days after the initiation of nitrite dosage (Figure 43). Note that the methanogenic activity was measured in the absence of nitrite through batch tests using fresh sewage without nitrite addition in all cases. The different levels of nitrite addition did not cause any difference in inhibition. Complete methanogenic inhibition was achieved with the lowest nitrite concentration used in the experiment, i.e. 40 mg-N/l. The recovery of CH₄ production proceeded at very similar rates in all cases. The recovery process was almost linear during the whole recovery phase. Only less than 60% of recovery was achieved after two-month of recovery. Suppression of sulphide production was also achieved with nitrite dosage at 80 mg-N/l and 120 mg-N/l. However, the recovery of sulphide production was considerably faster.

A field trial was conducted to test intermittent nitrite dosing as a means for sulphide and CH₄ control in a real sewer line, UC09, located in Gold Coast, Australia. As described in a previous section, UC09 has a length of 1080 m and a diameter of 150 mm. It receives primarily domestic wastewater with an average daily flow of approximately 200 m³. The hydraulic retention time of sewage in the rising main varied between 1.7 and 5.7 hours during the period of the study.

Nitrite solution was dosed into the UC09 wet well manually during daytime only (8:00 a.m. to 7:00 p.m.) over three consecutive days. Before each pumping event, nitrite was added to the wet well, resulting in a concentration in sewage of 100 mg-N/l, a level determined based on the lab-scale reactor test results (Jiang et al., 2010a, see also Figure 43). The dosage was stopped after three days. Therefore, the actual dosage time was 33 hours over a 3-day period.

Prior to nitrite dosing, three measurement campaigns were conducted to monitor CH₄ production in the rising main over a period of 7 days. Each campaign involved sampling wastewater hourly at both the pumping station wet well and 828 m downstream of the pumping station, for a period of 3 – 6 hours. The samples were analyzed for dissolved CH₄ concentrations using the method described in a previous section (5.5.1). Similar measurement campaigns were also conducted 1, 4, 5, 10 and 13 weeks after nitrite dosing.

FIGURE 43

NORMALIZED CH₄ PRODUCTION RATE IN NITRITE-DOSING REACTORS: R2 (○), R3 (□), AND R4 (△), RELATIVE TO THE CONTROL REACTOR R1 (I.E. THE ACTIVITY MEASURED FOR R1 ON EACH MEASUREMENT DAY WAS CONSIDERED 100%, TO ELIMINATE THE RATE VARIATION CAUSED BY CHANGES IN WASTEWATER QUALITY). NITRITE ADDITION WAS INITIATED ON DAY 0 AND FINISHED ON DAY 24

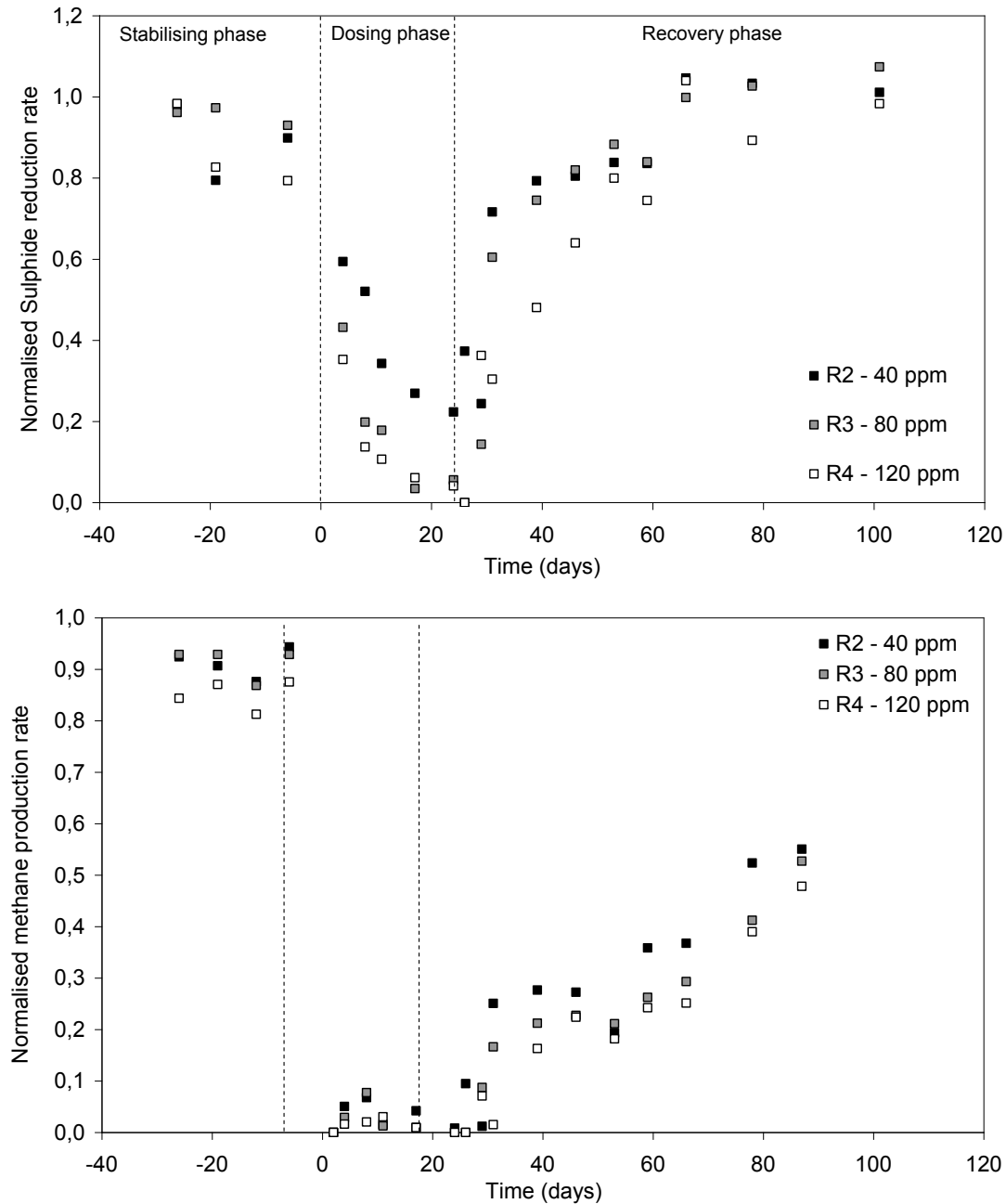
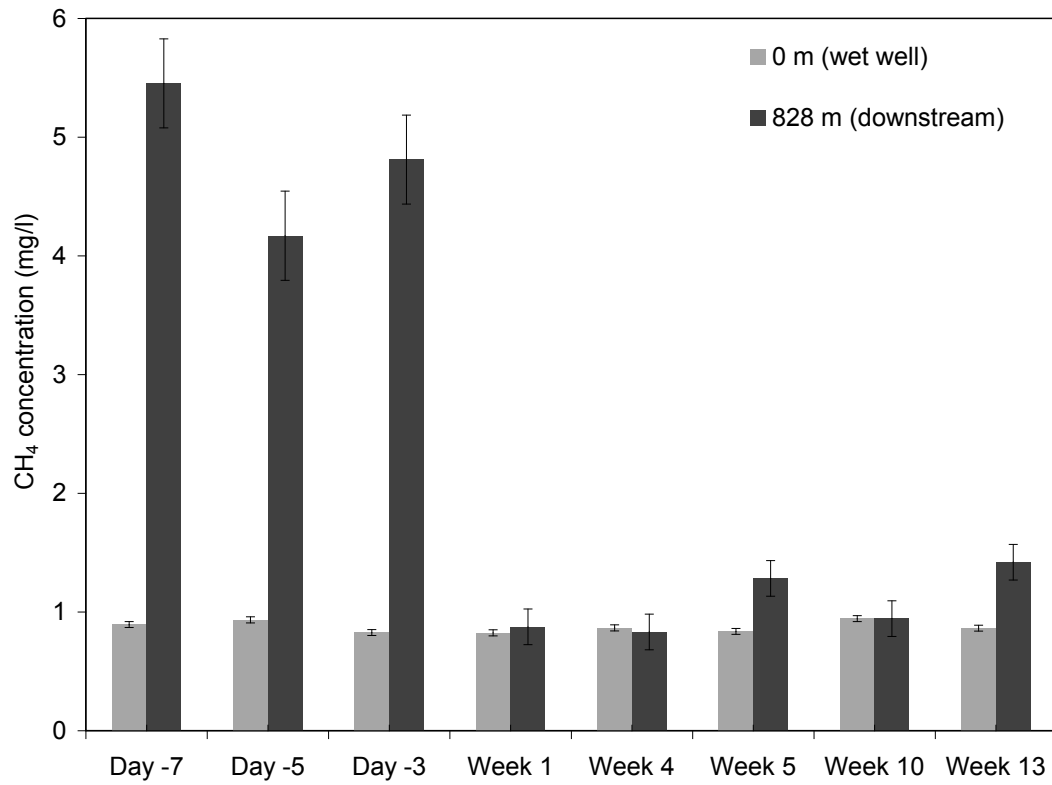


Figure 44 shows the CH₄ concentrations at the pumping station wet well and 828m downstream, where a sampling point was installed. Complete suppression of CH₄ production by nitrite addition was observed, as indicated by the nearly identical CH₄ concentrations in the pumping station and at 828m. One month after terminating nitrite dosage, CH₄ concentration at 828m remained at a level similar to that measured in the wet well, indicating that the sewer biofilm ceased to produce CH₄ in this period. The activity increased only marginally in the following two months.

FIGURE 44

DAILY AVERAGE CH₄ CONCENTRATION AT 828M AND AT THE PUMPING STATION WET WELL. DAY -7, -5, AND -3 WERE DURING THE BASELINE PERIOD. NITRITE WAS ADDED INTERMITTENTLY AT 100 MG/L DURING DAY 0 – 2 FOR A TOTAL DOSING TIME OF 33 HOURS OVER A 3-DAY PERIOD. WEEK 1 TO 13 IMPLY 1 – 13 WEEKS AFTER THE DOSAGE. METHANE CONCENTRATIONS WERE BASED ON MANUAL SAMPLING AND OFFLINE MEASUREMENTS WITH THE METHOD PREVIOUSLY DESCRIBED (3 – 6 SAMPLES WERE TAKEN ON EACH SAMPLING DAY). THE ERROR BARS SHOWN ARE STANDARD ERRORS (N = 3 – 6)



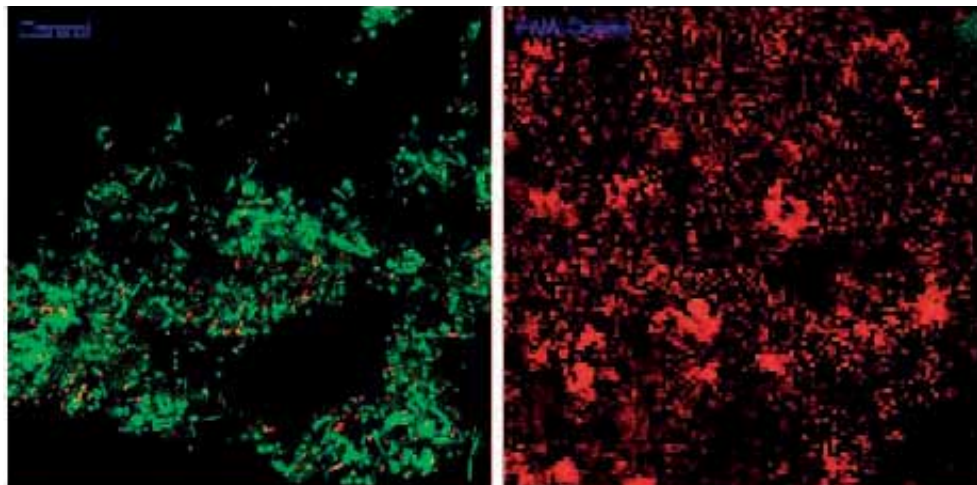
In general, the field trial confirmed lab study results that nitrite has a long-term toxic effect on methanogens (and sulphate reducing bacteria – data not shown) in anaerobic sewer biofilms. Both the field and laboratory results collectively suggest that nitrite could be applied intermittently to achieve sulphide and CH₄ control in sewers. However, given that nitrite is one of the factors that correlate with N₂O production, the impact of upstream nitrite addition and dosages on the potential on downstream N₂O emissions needs to be determined.

Jiang et al. (2010b) revealed that the suppression of CH₄ (and sulphide) production was due to a strong biocidal effect of free nitrous acid (FNA), the protonated form of nitrite, on microorganisms. The viable cells in sewer biofilms were found to decrease substantially from approximately 80% prior to nitrite dosage to 5 – 15% after 6 – 24 hour treatment at FNA levels above 0.2 mg-N/l (equivalent to 80 mgNO₂⁻-N/l at pH 6.0).

An example of the live and dead staining prior to and after FNA treatment is given in Figure 45.

FIGURE 45

CONFOCAL LASER SCANNING MICROSCOPY IMAGES OF HOMOGENIZED SEWER BIOFILMS STAINED WITH BACLIGHT® LIVE/DEAD STAINING KIT PRIOR TO (LEFT) AND AFTER (RIGHT) FNA TREATMENT. GREEN AND RED CELLS ARE INTACT AND DAMAGED CELLS, RESPECTIVELY



IRON SALT ADDITION

With a similar approach, Zhang et al. (2009) studied the impact of ferric chloride addition on microbial activities of anaerobic sewer biofilms.

Two lab-scale rising main sewer systems fed with real sewage were operated for 8 months. One received Fe³⁺ dosage at 15 mg per l of wastewater (experimental system) and the other was used as a control (no Fe³⁺ dosage). In addition to precipitating sulphide from bulk water, Fe³⁺ dosage was found to significantly inhibit sulphate reduction and CH₄ production by sewer biofilms, measured through batch tests. The sulphate reduction rate was found to be inhibited by 40 – 60%, while the methanogenic activity was inhibited by 50 – 80%. The rate data were supported by significantly lower CH₄ concentrations at the end of the experimental rising main system (for more details, see Zhang et al., 2009). The mechanisms responsible for the inhibition observed are yet to be fully understood.

Oxygen and nitrate addition

The injection of pure oxygen/air and nitrate, which is also commonly used for sulphide control in sewers, may also reduce CH₄ production due to elevated redox conditions. Oxygen may also be toxic to methanogens. However, the detailed effect is yet to be revealed despite that Mohanakrishnan et al. (2009) produced some preliminary evidence showing the absence of CH₄ production in a nitrate-receiving sewer system. Detailed studies on the effect of oxygen and nitrate addition on CH₄ production in sewers are currently underway at The University of Queensland.

6.6 METHANE EMISSION FRANCE

6.6.1 EMISSION OF CH₄

The emission of CH₄ that was measured at the four plants is presented in Table 20.

TABLE 20 EMISSION OF CH₄ AT FOUR WWTPS IN FRANCE

WWTP	% CH ₄ / of COD _{influent}
WWTP 1	<0.01%
WWTP 2	<0.01%
WWTP 3	0.04%
WWTP 4	0.03

In the four WWTPs, CH₄ represented 2 - 10% of the total direct emissions from biological reactors.

6.6.2 ORIGIN CH₄ EMISSION

The emission of CH₄ was very low in activated sludge reactors. Higher emissions can be found at pre- treatment or primary settling tanks but they were not quantified.

6.7 METHANE EMISSION UNITED STATES OF AMERICA

6.7.1 COLLECTION SYSTEM PHASE 1: EMISSION FACTORS

Figure 46 shows the winter and summer CH₄ concentration readings at the lift stations from the phase 1 data collection.

Table 21 provides CH₄ mass emissions from all three of the LCC series lift stations for comparison.

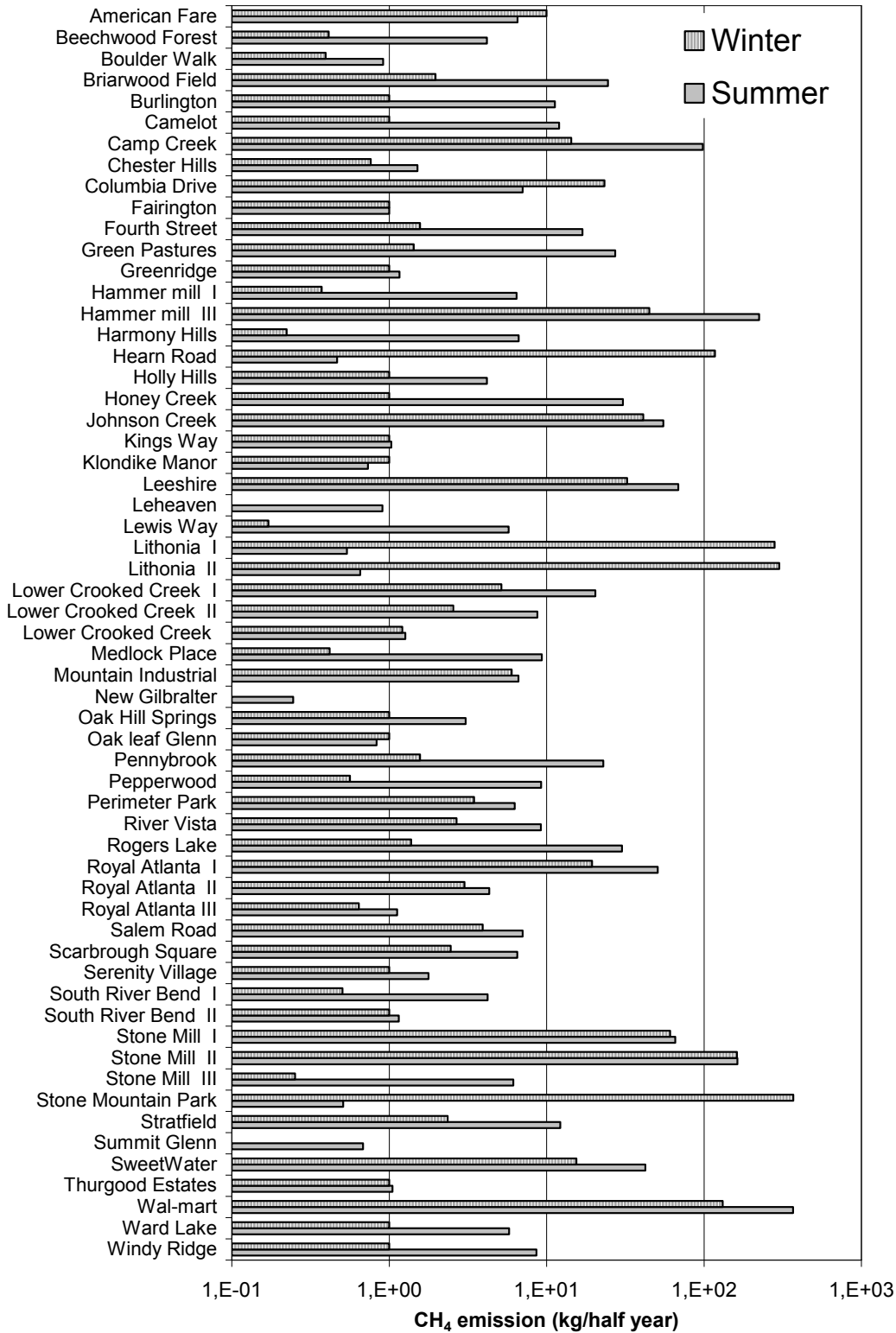
TABLE 21 COMPARISON OF CH₄ EMISSIONS FROM LCC PUMPING STATIONS

Lift station name	Estimated CH ₄ emissions (kg CH ₄ /year)	Calculation method	Notes / Comments
LCC-1	4,264	Mechanical ventilation rate and concentration in the air discharge header	Enclosed wetwell with positive ventilation
LCC-2	1,884	"breathing cycle" method	Covered wetwell with no ventilation, some CH ₄ could freely escape through openings for the screens
LCC-3	413	"breathing cycle" method	Completely open wetwell

The following observations are of interest:

- The reported emissions for LCC-1 are by far the largest of these three stations that convey approximately the same flow. This high value is likely a function of the continuous ventilation/flux and continuous ventilation methodology than any other factor.
- LCC-3, on the other hand, has the lowest reported emissions while receiving all of its flow from a force main (unlike the other two stations which are gravity fed) which is understood to increase the anticipated CH₄ production. The lower value is likely due to the uncovered fore bay and significant dilution; the other two stations have covered fore bays which limit dilution/dispersion of produced CH₄.

FIGURE 46 INSTANTANEOUS CH₄ READINGS DURING WINTER AND SUMMER MONITORING AT 59 LIFT STATIONS IN DEKALB COUNTY, GEORGIA, USA (NOTE X-AXIS IS LOGARITHMIC)



6.8 METHANE EMISSION THE NETHERLANDS

6.8.1 EMISSION OF CH₄

The emission of CH₄ as found during the measurements at three Dutch WWTPs is summarised in Table 22.

TABLE 22 EMISSION OF CH₄ FROM THREE WWTPS

WWTP	Emission (kgCH ₄ /kgCOD _{influent})
Papendrecht	0.0087
Kortenoord	0.0053
Kralingseveer October	0.012
Kralingseveer February	0.008
Emission factor NIR*	0.007 without sludge digestion 0.0085 with sludge digestion

*National Inventory Report

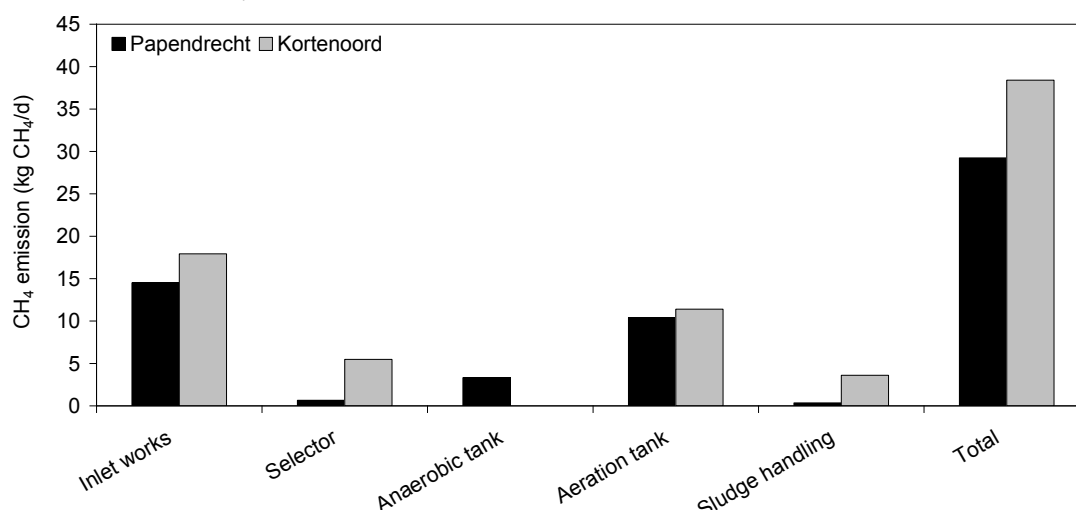
Based on Table 22 the following observations can be made:

- The highest emission of CH₄ occurred at WWTP Kralingseveer in October. This could be related to the presence of a sludge digester at this WWTP where at Papendrecht and Kortenoord a sludge digester is absent.
- In February the emission at Kralingseveer was lower than in October. This could be related to the temperature of the water which was 19°C in October and around 10 °C in February. At lower temperatures less CH₄ will (probably) be produced and more CH₄ will be dissolved. For this reason CH₄ could have left the WWTP via sludge or the effluent. However, the concentration of soluble CH₄ in both streams was not determined.
- The emission factors found for CH₄ are in some case higher in other cases lower than the emission factor currently used to estimate the emission of CH₄ from WWTPs.

6.8.2 ORIGIN OF CH₄ EMISSION

The total CH₄ emission was based on measurements at several locations at the three WWTPs. For Papendrecht and Kortenoord the origin of the CH₄ emission is presented in Figure 47. Based on Figure 47 it can be observed that in Papendrecht the inlet works and aeration tank contributed most to the emission of CH₄. The contribution from the inlet works was 45%, the aeration contributed 32%.

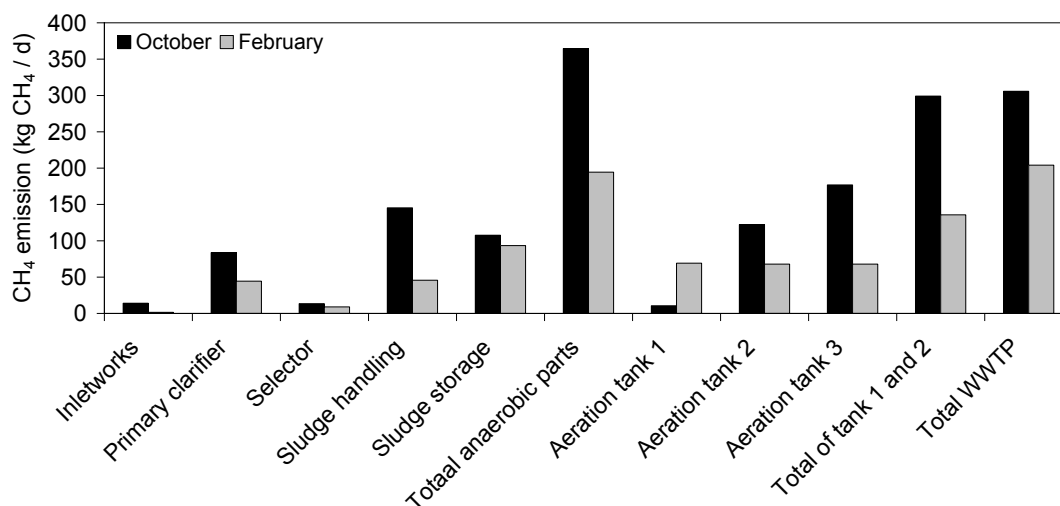
FIGURE 47 OVERVIEW OF THE CH₄ EMISSION AT WWTP PAPENDRECHT AND KORTENOORD



The emission of CH₄ at WWTP Kortenoord primarily occurred from the inletworks, selector and aeration tank, the contribution from the sludge handling was minimal. The contribution from the inlet works varied between 34 and 57%, the contribution from the selector varied between 21 – 41%, the contribution from the sludge handling remained limited with a maximum of 11%.

For both measurement periods at Kralingseveer the origin of the CH₄ emission is presented in Figure 48.

FIGURE 48 OVERVIEW OF THE CH₄ EMISSION AT WWTP KRALINGSEVEER DURING THE MEASUREMENT CAMPAIGN IN OCTOBER (2008) AND FEBRUARY (2009)



At WWTP Kralingseveer the ventilated air from all anaerobic parts is treated separately in a compost filter. After this treatment the air is reused in the aeration tanks 1 and 2. For this reason the total CH₄ emission after the anaerobic parts is presented as well in Figure 48.

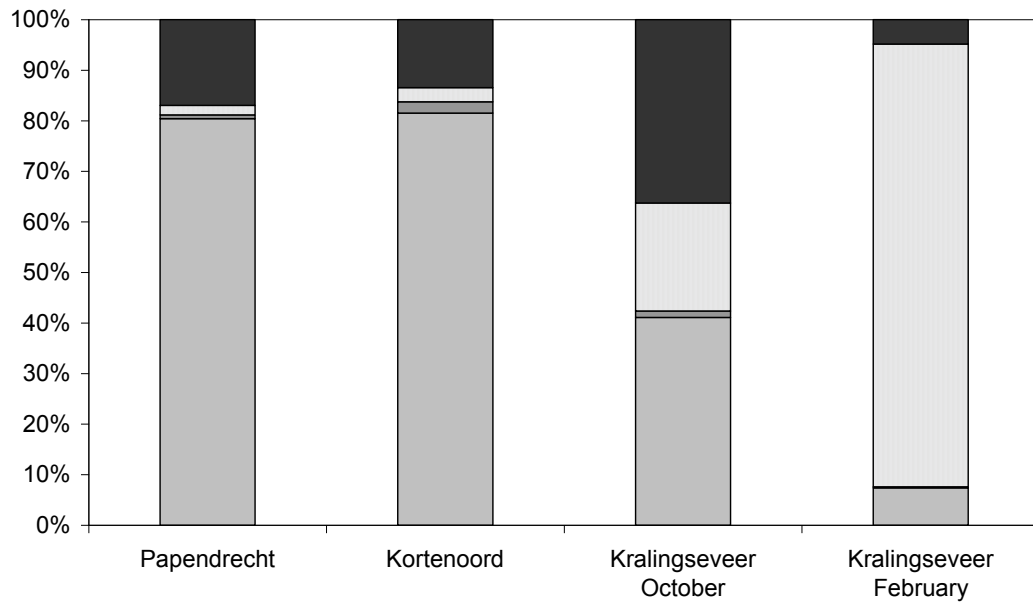
From Figure 48 it can be observed that the emission of CH₄ from all anaerobic parts is larger than the total CH₄ emission from aeration tanks 1 and 2. This implies that some CH₄ is lost or converted during treatment in the compost filter or in the old aeration tanks. The loss of CH₄ in October was 92 kg CH₄/d and for February the loss was 59 kg CH₄/d.

The main parts that contributed to the emission of CH₄ were the primary clarifier, the sludge thickeners, handling and storage. The contribution of the primary clarifier was 23% in both periods. For the sludge thickeners and sludge handling the contribution was 45% in October and 23% in February. Finally the sludge storage contributed 28% in October and 48% in February.

6.9 TOTAL CARBON FOOTPRINT

The contribution of each greenhouse gas to the total carbon footprint of the three WWTPs is presented in Figure 49.

FIGURE 49 TOTAL CARBON FOOTPRINT OF WWTP KORTENOORD (A; THE NETHERLANDS) AND WWTP KRALINGSEVEER OCTOBER (B; THE NETHERLANDS)



From Figure 49 it can be observed that:

- Electricity forms the major contributor to the total carbon footprint at WWTPs with a low N-sludge load (Papendrecht and Kortenoor).
- At the WWTP (Kralingseveer) with a higher N-sludge load the contribution of N₂O can be substantial.
- The contribution of CH₄ to the total carbon footprint depends on the presence of a sludge digester. In the last case (Kralingseveer) the contribution of CH₄ can be substantial.

7

DISCUSSION

7.1 METHODOLOGY

7.1.1 NITROUS OXIDE EMISSION

The research presented in this report used different methodologies to determine the emission of N₂O from WWTPs. The weak and strong points of these monitoring technologies are summarised in Table 23.

TABLE 9 STRONG AND WEAK POINTS METHODOLOGIES TO DETERMINE N₂O EMISSION FROM WWTPS REGARDING VARIABILITY AND ORIGIN
(+ REFERS TO BE SUITABLE, +/- REFERS TO BE LESS SUITABLE, - REFERS TO BE NOT SUITABLE)

Methodology	Variability	Origin / process parameters of influence
Liquid phase measurements / mass balance (Australia)	-	+
Gas hood (France / USA ¹)	+/-	+
Gas phase covered WWTPs (Netherlands)	+	+/-

1) Liquid phase measurements were taken as well.

To determine the N₂O emission from uncovered WWTPs different methodologies were developed. In Australia a method was developed based on liquid phase measurements for individual reactor zones. Based on mass transfer coefficients a complete mass balance over the different zones and the complete WWTP could be made. In this way the emission of N₂O was determined, but it was also possible to differentiate between N₂O emission and generation. However, this method does not allow for sufficient insight in the variation of N₂O emission as the method is based on grab samples.

The methodologies developed in the USA and France were based on gas hoods measurements. Those gas hoods were placed at the different zones of a WWTP and in case of the USA, the gas hoods measurements were combined with liquid phase measurements. By placing the gas hoods at the different zones of a WWTP and combine those with the liquid phase measurement it was also in the USA possible to differentiate between N₂O emission and generation. On the other hand, the gas-hood measurements are limited by the dimensions of the hood itself. Additionally, heterogeneities in the emission gas flow rate could also contribute to the variability in the actual emissions, which needs to be considered. To calculate the total emissions load from a plant, both the N₂O concentrations and advective gas flow must linearly scaled up from the hoods to the entire activated sludge bioreactor. Alternately, multiple measurements need to be done at different locations in the activated sludge reactor to capture the spatial variability in emissions.

The methodology used in the Netherlands was based on the analyses of the total off gas of covered WWTPs. The advantage of this method is that it captures the total emissions given the variability of N₂O emission in time and space, but with this methodology it is more difficult to differentiate between N₂O emission and generation.

To finally estimate and control the emission of N₂O from WWTPs, the use of all available methods will be required.

7.1.2 METHANE EMISSION

In the research performed in Australia, no CH₄ emission data were calculated. In the USA, the emission of CH₄ was calculated from unventilated lift stations. For this gas flow needed to be determined. This was done via the active volume of each lift station in combination with the pumping rate of each of the pumps. It is hereby assumed that the amount of air breathed in and exhausted out during each pumping cycle is approximately equal to the active volume inside the wet well or the volume of raw sewage pumped during each cycle. However, the methodology used to determine the gas flow is subject to a number of limitations, namely:

- Dispersion of CH₄, which is lighter than air, is not accounted for. Many stations cycle only a few times per day and those stations likely lose a considerable portion of the evolved CH₄ to the atmosphere in between cycles. This phenomenon would result in under-reporting of actual emissions.
- Dispersion effects can be very significant at some of the larger pumping stations, like LCC-3. LCC-3 has a large, uncovered forebay with significant interchanges of outside air. On calm days the forebay air changes could be one to five times per hour and could increase to as much as 10 or 30 times per hour on windy days. This limitation, combined with the fact that these stations have the highest volume and mass throughput which should produce more CH₄, could represent a very significant source for under-reporting.
- During a pumping cycle, a significant volume of outside air is drawn into the wet well. If a sampling event occurred soon after such a dilution, the applied concentrations could under-report the actual CH₄ emissions.

These limitations show that there is a need for a good strategy to measure loads of CH₄ emitted to the air from sewerage systems.

7.2 NITROUS OXIDE EMISSION

7.2.1 EMISSION

The emission of N₂O as measured at all the different WWTPs in Australia, France, USA and the Netherlands is summarised in Table 24.

TABLE 24 OVERVIEW N₂O EMISSION AT THE SURVEYED WWTPS AROUND THE WORLD (VALUES PRESENTED IN BOLD ARE REPORTED OUTLIERS).

Country	WWTP	Sample round	Emission (kg N ₂ O/kg TKN _{influent})	Emission (kg N ₂ O-N/kg TKN _{influent})
Australia	1 – Ox. Ditch	1	0.005	0.003
		2	0.003	0.002
		3	0.006	0.004
		4	0.003	0.002
	2 – Johannesburg	1	0.016	0.010
		2	0.027	0.017
		3	0.011	0.007
	3 – SBR	1	0.009	0.006
		2	0.011	0.007
		3	0.050	0.032
	4 – MLE (1)	1	0.019	0.012
	5 – MLE (2)	1	0.014	0.009
		2	0.047	0.030
		3	0.006	0.004
		4	0.055	0.035
	6 – MLE (3)	1	0.176	0.112
		2	0.008	0.005
		3	0.005	0.003
	7 – A ² O	1	0.011	0.007
		3	0.006	0.004
France	WWTP 1		<0.0002	<0.0001
	WWTP 2		<0.0002	<0.0001
	WWTP 3		0.003	0.0017
	WWTP 4		0.002	0.0011
USA	Separate-stage BNR	1 (15°C)	0.0005	0.0003
		2 (23°C)	0.0002	0.0001
	Four-stage Bardenpho	1 (14°C)	0.0025	0.0016
		2 (23°C)	0.0094	0.006
	Step-feed BNR 1	1 (19°C)	0.025	0.016
		2 (25°C)	0.0097	0.0062
	Step-feed non-BNR	1 (17°C)	0.0028	0.0018
		2 (26°C)	0.028	0.018
	Separate centrate ¹⁾	1 (30°C)	0.0038	0.0024
		2 (34°C)	0.0085	0.0054
	Plug-flow 1	1 (11°C)	0.0063	0.004
		2 (23°C)	0.0064	0.0041
	Plug-flow 2	1 (11°C)	0.0097	0.0062
		2 (22°C)	0.0014	0.0009
	MLE 1	1 (26°C)	0.0011	0.0007
	MLE 2	1 (26°C)	0.0009	0.0006
	Step-feed BNR 2	1 (29°C)	0.024	0.015
	Oxidation ditch	1 (19°C)	0.0005	0.0003
	Step-feed BNR 3	1 (24°C)	0.0008	0.0005
Netherlands	Papendrecht	1 (19°C)	0.00063	0.00040
	Kortenoord	1 (19°C)	0.00075	0.00048
	Kralingseveer	1 (18°C)	0.0066	0.0042
		2 (10°C)	0.096	0.061

1) Not a typical WWTP

The results of the emission of N₂O from WWTPs (see Table 24) exhibit a great variability among different WWTPs and at the same WWTP during different seasons or during the day. This great variability could be understood from the fact that the formation of N₂O is very complex and can be executed by both autotrophic and heterotrophic bacteria.

Those bacteria show a dynamic response to changing influent loads and process conditions resulting in a variable formation of N₂O in time and place. Because of the observed variability in N₂O emission the use of a generic emission factor to estimate the emission from a specific WWTP is inadequate. To determine the emission of N₂O from a specific WWTP measurements are required.

7.2.2 ORIGIN

The results presented in this report showed that the majority of the N₂O emission occurred from aerated zones. This emission was observed at the transition point between anoxic and aerobic zones (see Figure 22 and Figure 28) and in complete aerobic zones (see Figure 22 and Figure 28). The emission of N₂O at the transition point between anoxic and aerobic zones can be the result of:

- Stripping of N₂O formed during denitrification
- N₂O formation by heterotrophic denitrification
- N₂O formation by autotrophic nitrification

Stripping of N₂O formed during denitrification can primarily occur at the transition of anoxic to aerobic zones an example of this can be seen from Figure 28. At this transition point between anoxic and aerobic conditions formation of N₂O is possible by both heterotrophic denitrification and autotrophic nitrification. Heterotrophic N₂O emissions can be related to oxygen inhibition as was shown by several authors (Knowles, 1982; Korner and Zumft, 1989; Zumft, 1992; Hanaki et al., 1992). Autotrophic N₂O emissions can be related to the fact that in the presence of non limiting DO and NH₄⁺-N concentrations (as occurs at the transition point between anoxic and aerobic zones) N₂O can be formed by autotrophic nitrification as was recently proven by Yu et al. (2010). This process is most likely also responsible for the formation of N₂O under complete aerobic conditions, although the possibility of heterotrophic denitrification within sludge flocs cannot be discounted (Grady et al., 1999).

Although both heterotrophic denitrification and autotrophic nitrification can be responsible for the formation of N₂O, autotrophic nitrification seems to be the major contributor to the total N₂O emission from a WWTP based on the results reported by the USA (Figure 28) and France (Figure 22). This is supported by the found positive correlations between NO₂⁻-N, NH₄⁺-N and DO concentrations and N₂O emission (see Table 12).

7.2.3 PROCESS PARAMETERS INFLUENCE

Process parameters that influenced the emission and formation of N₂O from aerobic zones were found to be NO₂⁻-N, NH₄⁺-N and DO (isolated effect) and NH₄⁺-N – NO₂⁻-N (interactive effect) as could be observed from Table 12. All these parameters showed a positive correlation with the emission of N₂O, which means that the emission of N₂O increased as the concentration of the mentioned components increased.

Nitrite is one of the most important factors contributing to N₂O emission from nitrification as was proven by Beaumont et al. in several papers (2004a, 2004b and 2005).

From this the correlation of NO₂⁻-N with aerobic zone N₂O emissions is consistent with basic biochemical principles. However, it should be noted that the inhibitory effect of nitrite does not further increase at levels above ~2 mg NO₂⁻-N/l.

The positive correlation between NH₄⁺-N and DO with N₂O emission are in congruence with work of Yu et al. (2010) on N₂O formation by autotrophic nitrification. In this work it was proven that abrupt increases in DO concentrations in the presence of NH₄⁺-N can lead to transient accumulation of NO₂⁻-N, which in turn result in autotrophic NO and N₂O generation (Yu et al., 2010). Furthermore, it was shown that the magnitude of autotrophic nitrification driven N₂O emission also directly depends on the NH₄⁺-N concentrations. The positive interactive correlation between NH₄⁺-N and NO₂⁻-N concentrations and N₂O emissions suggests high N₂O emissions from aerobic zone locations, with simultaneously high concentrations of both NH₄⁺-N and NO₂⁻-N. This interactive correlation also points to autotrophic N₂O generation mechanisms, since both NH₄⁺-N and NO₂⁻-N are co-substrates in autotrophic denitrification by nitrifying bacteria (Beaumont et al., 2005; 2004a, 2004b).

Formation and emission of N₂O from anoxic zones was only found to be influenced by the NO₂⁻-N – DO concentration (interactive effect). This positive correlation of NO₂⁻-N – DO concentrations with N₂O emissions is consistent with known mechanisms of denitrification-related N₂O production. These mechanisms were proven by several authors. DO inhibition during denitrification was proven by Tallec et al. (2008) and Park et al. (2000); NO₂⁻-N inhibition was proven by Korner and Zumft (1989), von Schultesss et al. (1995) and Zumft (1997). Inhibition of denitrification by high DO concentrations could also lead to NO₂⁻-N build up, indirectly leading to N₂O emissions (Hanaki et al., 1992; Zhou et al., 2008).

7.2.4 IMPLICATIONS OF GAINED KNOWLEDGE

Based on the above, it becomes clear that at increasing levels of NH₄⁺-N, NO₂⁻-N and DO, the formation of N₂O increases in aerobic zones. This implies that if high concentrations of these constituents can be avoided in practice, the risk of N₂O emission will be reduced. These high concentrations can be avoided in:

- systems that approach “ideal” well-mixed conditions (i.e. high recycle rates). In this way concentrations of intermediates from nitrification-denitrification including NO₂⁻-N and NO are diluted, thereby reducing their inhibitory effect (Casey et al., 1999a,b). This is confirmed by the general results from all studies that WWTPs with the highest recycle rates and with the lowest TN effluent concentrations belonged to the WWTPs with the lowest N₂O emission.
- systems that avoid over – aeration i.e. have a rapidly responding DO control systems. In this way high levels of DO in aerobic and anoxic zones can be avoided
- systems that are equipped with flow equalization. Such systems minimize the peaking factor of influent nitrogen (ammonium) loading to the activated sludge system.

With the presented insight, it has been possible to make a first estimate on the risk level of N₂O emissions from a specific WWTP. This estimation can be based on the risk matrix presented in Table 25.

TABLE 25

RISK MATRIX TO DETERMINE RISK LEVEL OF N₂O EMISSION

Parameter	Risk on N ₂ O		
	High	Medium	Low
Effluent total organic nitrogen (mg/l)	> 10	5 - 10	< 5
Range in N-concentration in plant	H	M	L
Load variations	H	M	L
Maximum NO ₂ ⁻ -N conc. anywhere in plant	> 0.5*	0.2 – 0.5	0.2

* Risk does not increase at NO₂⁻-N concentrations higher than 2 mg N₂O-N/l.

In summary, it can be concluded that in systems with extensive nitrogen removal the formation of N₂O is greatly minimized. In other words there is no conflict between water quality and air quality; rather they go hand in hand. This implies that systems that are not designed for (complete) nitrogen removal exhibit a high risk of N₂O emission. At temperatures of 20 °C, an aerobic SRT of 2-3 days is already sufficient to achieve nitrification. In these high-loaded systems, nitrite easily accumulates resulting in greater N₂O formation.

7.2.5 FUTURE RESEARCH

The results of the presented research performed in Australia, France, the USA and the Netherlands already clearly identified key process parameters that influence the emission of N₂O from WWTPs. This knowledge presents a good starting point for defining mitigation strategies to reduce the emission of N₂O. Future research should first give better insight in the variability of N₂O emissions over prolonged times in order to develop proper sampling protocols for emissions of N₂O from treatment plants. A better understanding of the processes leading to nitrite formation in treatment plants can lead to mitigation strategies for N₂O emissions. Besides activated sludge systems, biofilm processes should be monitored for potential emissions. A better insight in N₂O production in natural systems can improve decisions on implementing full nitrogen removal as a mitigation strategy for N₂O emissions from treatment systems as well as natural systems.

7.3 METHANE EMISSION

7.3.1 SEWERS

The measurements performed in Australia and the USA show that substantial amounts of CH₄ can be formed and emitted from sewer systems. In addition to this substantial amounts of CH₄ were measured from the inlet works at the WWTPs in the Netherlands. With this knowledge, the contribution of the sewer system can not be neglected as it is dismissed in the current IPCC guidelines (IPCC 2006a). However, to be able to make a good estimate of the contribution of sewers to the total CH₄ emission from wastewater collection and treatment systems more data are required and a good strategy to measure CH₄ loads is necessary (refer to section 7.1.2). The data collected to date are far from adequate for the development of reliable accounting guidelines, and for the development, calibration and validation of CH₄ emission models. It is of utmost importance that such data are collected from sewer networks (rising mains and gravity sewer) around the world under different climate conditions, and from networks collecting and conveying various types of wastewaters. Both liquid and gas phase data are urgently needed.

The data obtained in Australia suggest that the wastewater composition, HRT and the dimensions (Area / Volume ratio) are parameters that can influence the level of CH₄ formation from a rising main.

The results from the lift stations in the USA (Figure 46) showed that the concentration of CH₄ in the gas phase was in almost all cases (80%) higher in summer than in winter. This indicates that the temperature is an important parameter determining the formation of CH₄ in sewers. This might also explain relative high CH₄ levels in Australia (although this could also result from easy degradable nature of wastewater).

Finally the gas phase measurements (%LEL) in Australia indicate that the composition of wastewater is another parameter that determines the formation of CH₄ in sewers. The results of these field measurements were confirmed by results from laboratory measurements performed by Sudarjanto et al. (2010b). In these laboratory experiments the addition of a certain volume by percent of brewery wastewater to raw sewage lead to an increase in the CH₄ production rate compared to a control reactor. Interestingly in another experiment described by Sudarjanto et al. (2010b) it was found that the addition of a certain volume by percent of dairy waste to raw sewage did not lead to any increase in CH₄ production. These results suggest that each trade waste should be assessed separately, but that the composition of the wastewater is a parameter to taken into account for the formation of CH₄ in sewers.

7.3.2 MITIGATION STRATEGIES

Several strategies used for sulphide control in sewers have been found to reduce CH₄ formation (see section 6.5.5). However these strategies may not be the most cost-effective methods for CH₄ control. Given the high sensitivity of methanogens to environmental changes, more cost-effective strategies may be developed. The addition of nitrite (free nitrous acid) is a very promising technology in this respect, but might easily lead to N₂O formation.

7.3.3 WASTEWATER TREATMENT PLANTS

Based on the results from the Netherlands it was found that the current emission factors used in the Netherlands are for the present useful to estimate the emission from WWTPs. However, the amount of measurements was limited and in general measurements at several WWTPs should be performed to confirm existing emission factors used per country.

The results from the measurements at the WWTPs without sludge digestion (Figure 47) showed that CH₄ emission primarily occurred from the inlet works and the aeration tanks. The high emission from the inlet works strongly indicates that the CH₄ emitted originates from the sewer as due to the very short HRT formation of CH₄ in the inlet works can be excluded. Furthermore, the wastewater to both WWTPs is transported via rising mains in which under anaerobic conditions CH₄ formation is possible as was shown by the results from Australia. The rather large contribution of the aeration tank to the total CH₄ emission is rather strange as under anoxic and aerobic conditions CH₄ formation is very unlikely. Part of the CH₄ emitted from the aeration tank could have been formed in the anaerobic tank or anoxic tank upstream the aeration tanks. However, this could not be confirmed due to the lack of measurements from the liquid phase. Furthermore, it is known that limited amounts of methanogens are present in activated sludge (Gray, et al., 2002 and Lens et al., 1995). Based on these considerations it is thought that the CH₄ emitted from the aeration tank mainly originates from the sewer system.

At the WWTP with sludge digestion the highest relative CH₄ emission was found from the sludge handling and storage sites (in total ~50%). This could be understood from the fact that these sites handled sludge from the digester. The contribution from the sewer was in this case smaller (~25%).

From the results at the WWTP it is shown that process units that are known for their sulphur related odours (inlet works, sludge digesters) also emit CH₄.

7.3.4 TOTAL CARBON FOOTPRINT WWTP

As a first indication on the possible contribution of N₂O and CH₄ emission to the total carbon footprint of a WWTP, the result in the Netherlands could be used as an example.

In the case studies in the Netherlands, the specific emissions of N₂O and CH₄ were determined at the same time. Together with the data on the related consumption of electricity and natural gas, it was possible to calculate a carbon footprint of three WWTPs. To determine the carbon footprint, all sources were converted to CO₂ equivalents¹⁵. The results in the Netherlands indicated that the emission of CH₄ and N₂O can significantly contribute to the total carbon footprint of a WWTP. This contribution can vary from 2% to almost 90% of the carbon footprint under extreme conditions for N₂O and 5 – 40% for CH₄. One should be aware that these numbers are specific for the Netherlands. In any other country, these numbers can differ greatly as there exist a great variation in the way wastewater and sludge is handled as well as the specific composition of the energy mix used.

Furthermore these numbers can significantly differ depending on how the boundaries are set around the analysis. In case of the analysis performed for the three Dutch WWTPs the contribution of e.g. chemical use, and sludge incineration were not accounted for.

¹⁵ It should be noted that the conversion numbers are country specific and do depend on the used energy mix (i.e. brown coal versus wind or solar energy), which is of influence on the total carbon footprint of a WWTP.

8

CONCLUSIONS AND FUTURE RESEARCH

8.1 CONCLUSIONS

8.1.1 NITROUS OXIDE EMISSION

Based on the research presented in this report the following conclusions can be drawn:

- The emission of N₂O is highly variable. For this reason the use of generic emission factor to estimate the emission from a specific WWTP is inadequate.
- To determine the emission of a specific WWTP, measurements are required. These measurements should be performed with online monitors and over the operational range of the WWTP.
- Emission of N₂O originates mainly from nitrification.
- Accumulation of nitrite leads to the formation of N₂O in mainly aerobic zones.
- High NH₄⁺-N concentrations can lead to the emission of N₂O if nitrification occurs.
- The risk level of N₂O emission can be determined based on the following risk matrix:

TABLE 26

RISK MATRIX TO DETERMINE RISK LEVEL OF N₂O EMISSION

Parameter	Risk on N ₂ O		
	High	Medium	Low
Effluent total organic nitrogen (mg/l)	> 10	5 - 10	< 5
Range in N-concentration in plant	H	M	L
Load variations (daily)	H	M	L
Maximum NO ₂ ⁻ -N concentration (mg N/l) anywhere in plant	> 0.5*	0.2 – 0.5	0.2

* Risk does not increase at higher NO₂⁻-N concentrations

- In addition to the risk matrix, a rapidly responding DO control also contributes in minimising the risk of N₂O emission.
- A good effluent quality (TN < 5 mgN/l) goes hand in hand with a low risk of N₂O emission.
- Non BNR systems with unintentional nitrification will have a high risk of N₂O emission.

8.1.2 METHANE EMISSION

Based on the research presented in this report the following conclusions can be drawn:

- Formation and emission of CH₄ from sewers can be substantial.
- Sulphur related odours are good indicators for CH₄ formation.
- Odour mitigation strategies in sewers likely also support a reduced CH₄ formation.
- Methane emission from WWTPs mainly originates from sewers and sludge handling.
- Emissions of CH₄ and N₂O significantly contribute to the total greenhouse gas footprint of WWTPs.

8.1.3 TOTAL CARBON FOOTPRINT

- Emissions of CH₄ and N₂O may significantly contribute to the total greenhouse gas footprint of WWTPs as was based on the results from the Netherlands.

8.2 FUTURE RESEARCH

Based on the outcomes of the research valuable knowledge is gained to estimate and reduce the emission of N₂O and CH₄ from wastewater collection and treatment systems. Future applied research should focus on:

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- Variability of N₂O emission such that proper sampling and monitoring programmes can be developed.
- Mitigation strategies (including insight in the relative contribution of autotrophic and heterotrophic processes to N₂O generation) to reduce emission via process design and control;
- Evaluate emission from biofilm based processes;
- Evaluate emission from various receiving aquatic environments.

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- Development of a strategy to measure loads emitted from sewers;
- Build a database for emission measurements from sewers;
- Cost effective mitigation strategies¹⁶;
- Evaluate emission from sludge treatment lagoons.

¹⁶ The addition of nitrite seemed to be a strategy that could reduce the formation of methane, however, given that nitrite is one of the factors that correlate with N₂O production, the impact of upstream nitrite addition and dosages on the potential on downstream N₂O emissions needs to be determined as well.

9

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APPENDIX 1

OVERVIEW APPLIED EMISSION FACTORS CH₄

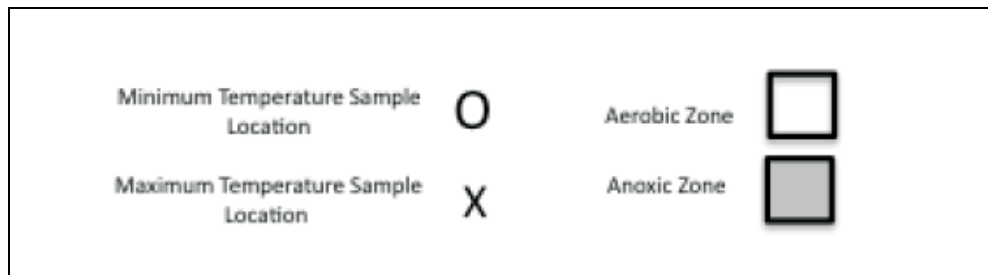
Country	Details of methodology	Ref. in NIR																				
UK	Estimates based on Hobson <i>et al.</i> (1996) who estimated emissions for 1990-1995. Subsequent years extrapolated on the basis of population. Empirical EFs have been calculated for 14 different treatment routes. The method complies with the IPCC Good Practice Guidance as a national model (IPCC, 2000).	Hobson <i>et al.</i> (1996)																				
Australia	IPCC default methodology. Tier 2 methodology developed for industrial wastewater.																					
Canada	Based on IPCC methodology; uses EF of 4.015 kgCH ₄ /person/year for CH ₄ emissions from wastewater sector. No evaluation of emissions for industrial wastewater due to lack of activity data.	ORTECH Corp. (1994)																				
Denmark	IPCC Tier 2 country-specific methodology. According to IPCC GL (1996), domestic and industrial wastewaters (and respective sludge) should be accounted separately. In Denmark, most industrial wastewater is treated at centralized WWTWs and the data available for the Total Organics in Wastewater (TOW) does not differentiate between industrial and municipal sewage sludge. Therefore it was assumed that the characteristics of TOW in domestic and industrial wastewater are similar and the division between the two types of sewage sludge was derived by subtracting the %age industrial contribution to the total emissions. The IPCC default methodology was applied from 1990 to 1998, by accounting and correcting for the industrial influent load. Since 1999 onwards actual data from the industry sector has been collected and the TOW calculated using country-specific data. The EF is calculated by the IPCC methodology (1996) using the default value of 0.6 kg CH ₄ /kg BOD but country-specific values for the MCF. The following constant values have been extrapolated: 0.26 kg CH ₄ /kg BOD based on the dry weight fraction of anaerobically treated sludge and 0.15 kg CH ₄ /kg BOD based on the wet weight fraction. The fractions that are used for biogas, combustion or reuse including combustion include methane potentials that are recovered or emitted as CO ₂ . The estimated CH ₄ potentials of these fractions have been estimated and subtracted from the calculated gross emission of CH ₄ .	Thomsen and Lick (2005)																				
Germany	Municipal wastewater treatment uses aerobic procedures, therefore it produces no CH ₄ emissions (Methane Correction Factor: MCF=0). CH ₄ emissions from cesspools and septic tanks (which is exceptional in Germany) are determined following the IPCC methodology.	FEA (2008)																				
France	Minor variation on IPCC default methodology.																					
Japan	Country specific EFs were developed: 1) From sewage treatment plants: 528.7 mg CH ₄ /m ³ (to secondary treatment) & 348.0 mg CH ₄ /m ³ (from sludge); uncertainty in emissions 33% (95%ile). 2) From in-situ domestic sewage treatment (septic tanks): <table><tr><th>Domestic wastewater treatment facilities</th><th>CH₄ EF (kg CH₄/person-year)</th></tr><tr><td>Community plants</td><td>0.195</td></tr><tr><td>Gappei-shori johkasou</td><td>1.106</td></tr><tr><td>Tandoku-shori johkasou</td><td>0.197</td></tr><tr><td>Vault toilets</td><td>0.197</td></tr></table> 87% uncertainty in emissions estimation (95%ile) 3) From collected domestic sewage (septic tanks) <table><tr><th>Treatment method</th><th>CH₄ EF (kg CH₄/m³)</th></tr><tr><td>Anaerobic treatment</td><td>0.543</td></tr><tr><td>Aerobic treatment</td><td>0.00545</td></tr><tr><td>Standard de-nitrification treatment</td><td>0.0059</td></tr><tr><td>High load de-nitrification treatment</td><td>0.005</td></tr></table>	Domestic wastewater treatment facilities	CH ₄ EF (kg CH ₄ /person-year)	Community plants	0.195	Gappei-shori johkasou	1.106	Tandoku-shori johkasou	0.197	Vault toilets	0.197	Treatment method	CH ₄ EF (kg CH ₄ /m ³)	Anaerobic treatment	0.543	Aerobic treatment	0.00545	Standard de-nitrification treatment	0.0059	High load de-nitrification treatment	0.005	GIO (2008)
Domestic wastewater treatment facilities	CH ₄ EF (kg CH ₄ /person-year)																					
Community plants	0.195																					
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Vault toilets	0.197																					
Treatment method	CH ₄ EF (kg CH ₄ /m ³)																					
Anaerobic treatment	0.543																					
Aerobic treatment	0.00545																					
Standard de-nitrification treatment	0.0059																					
High load de-nitrification treatment	0.005																					

Country	Details of methodology			Ref. in NIR
	Membrane separation			
	Other	0.00545	0.00545	
Netherlands	101% uncertainty in emissions estimation (95%ile) IPCC Tier 2 method. Anaerobic industrial wastewater treatment: EF = 0.056 t CH ₄ /t DOC design capacity. Urban wastewater treatment (and anaerobic sludge handling): EF = 0.085 t CH ₄ /t DOC influent. Septic tanks: EF = 0.0075 t CH ₄ /year/person (MCF=0.5; B ₀ =0.25)			
New Zealand	Refinement of the IPCC methodology (1996). Characteristics EFs have been calculated for 9 typical on-site treatment processes (largely used in the Country). They have been calculated from the proportion of BOD to the plant which is anaerobically degraded, multiplied by the CH ₄ conversion factor MCF. IPCC default methodology also used for industrial wastewater.			SCS Wetherhill Environmental, (2002); Beca (2007)
Sweden	Not estimated (either recovered as heat and bio-energy, or insignificant).			
USA	A slightly modified IPCC 1996 methodology was used. CH ₄ emissions from septic systems, WWTW's and sludge digestion are calculated separately. The IPCC default factors (B ₀ and MCF) are used. Emissions from industrial wastewater were developed according to the IPCC (1996) methodology (four big industries producing methane have been identified and activity data collected)			USEPA (2008)
Austria	IPCC method with MCF for septic tanks = 0.27 (calculated from average temperatures in Austria: 20°C for 2/3 of the year with MCF = 35% and 10°C for 1/3 of the year with a MCF = 10%)			Steinlechner <i>et al.</i> (1994)

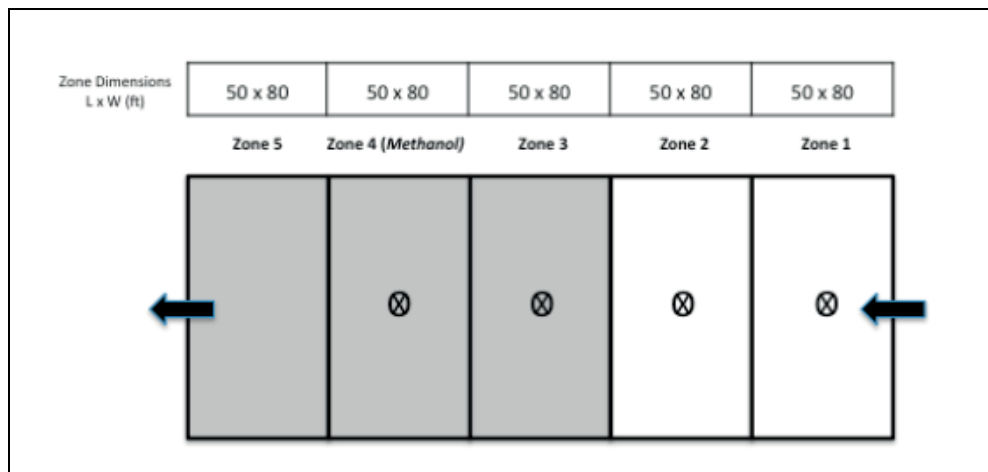
APPENDIX 2

DIMENSION WWTPS AND SAMPLING POINTS USA

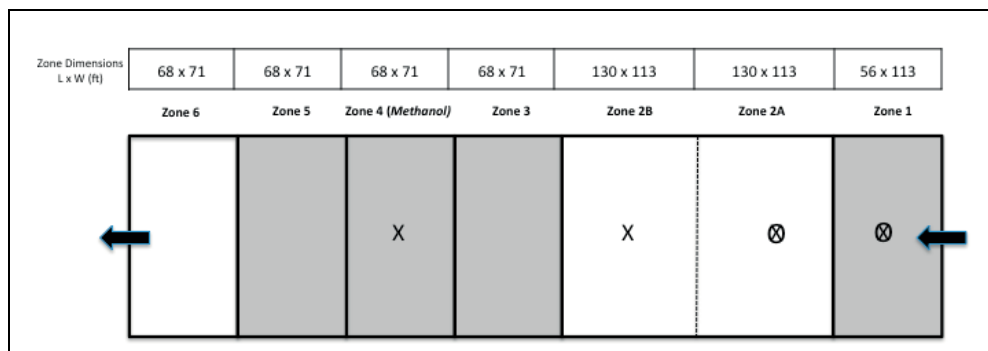
LEGEND



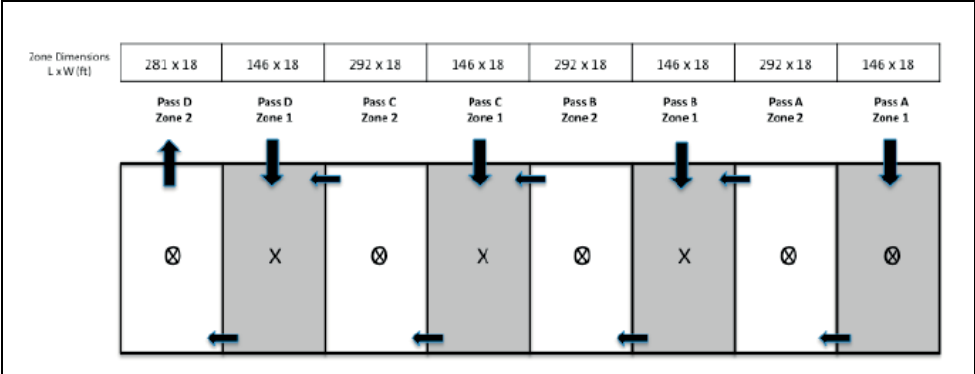
SEPARATE-STAGE BNR



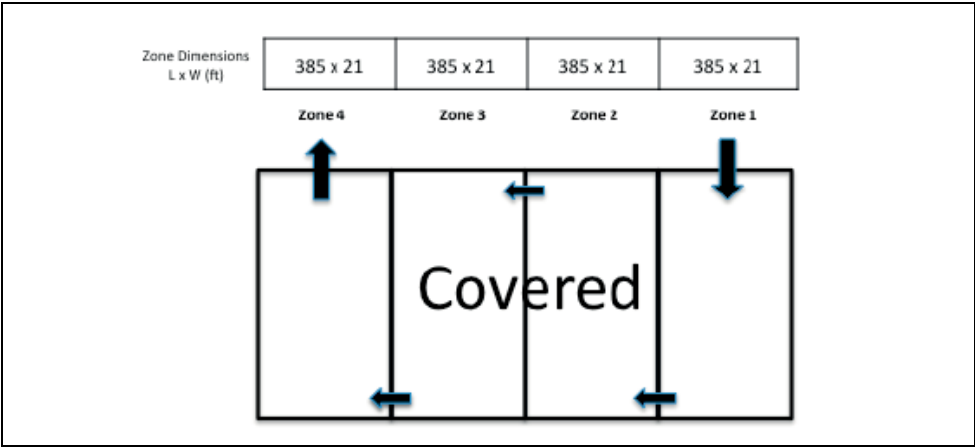
FOUR-STAGE BARDENPHO



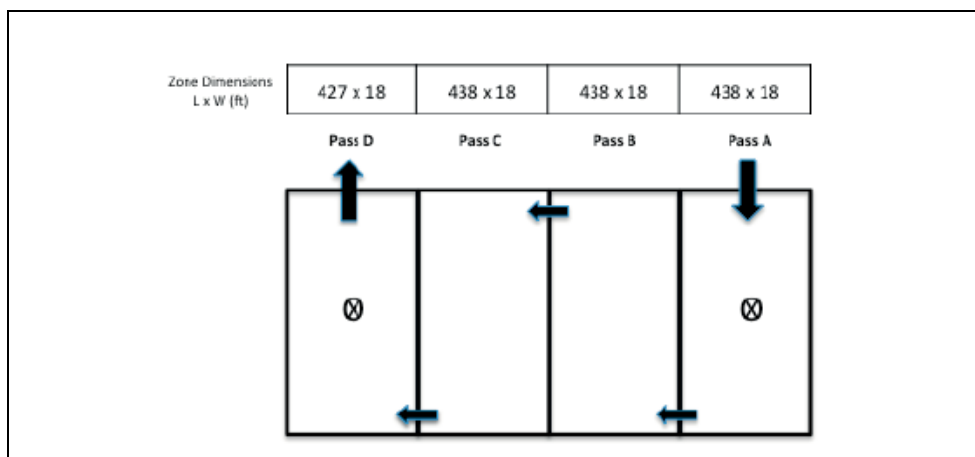
STEP-FEED BNR 1



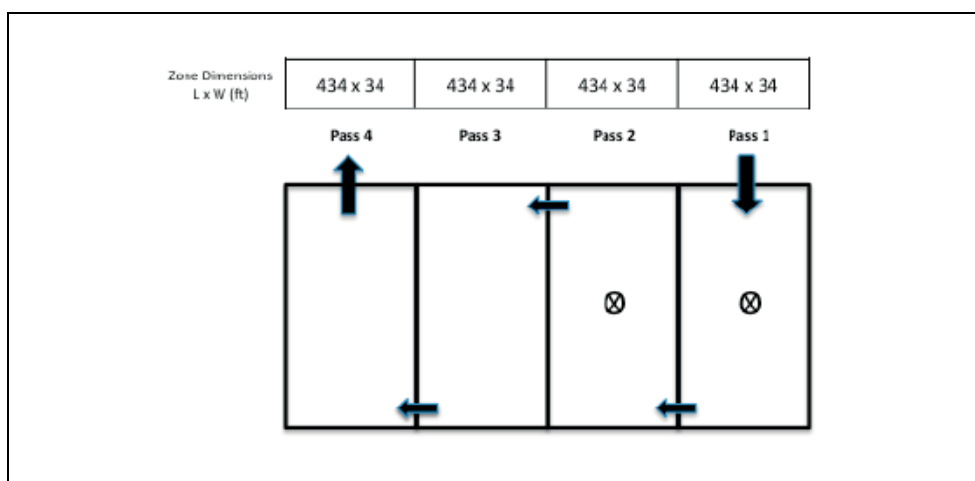
STEP-FEED NON BNR



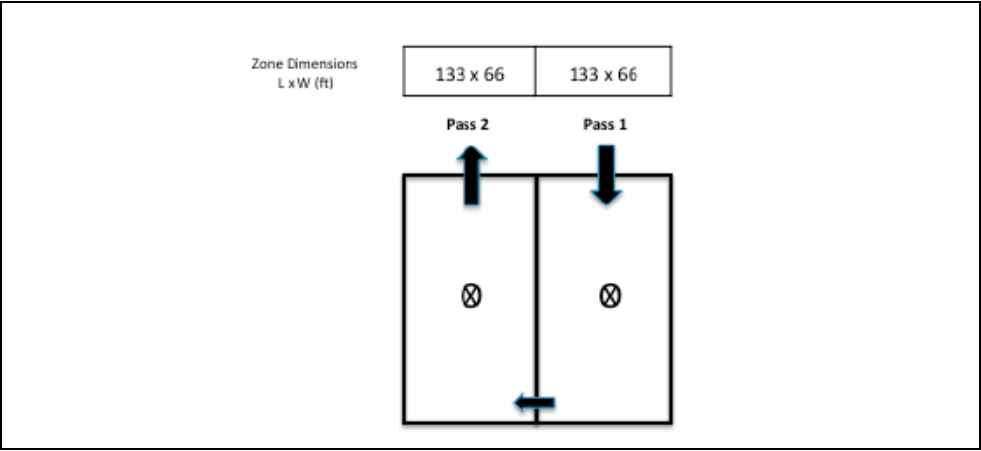
SEPARATE CENTRATE



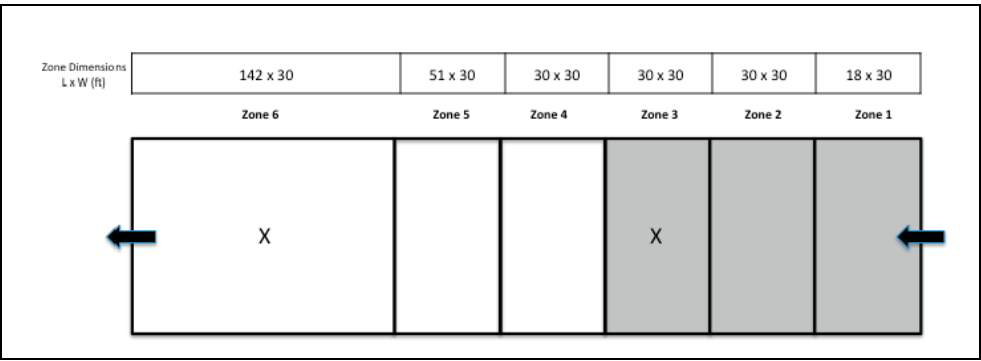
PLUG-FLOW 1



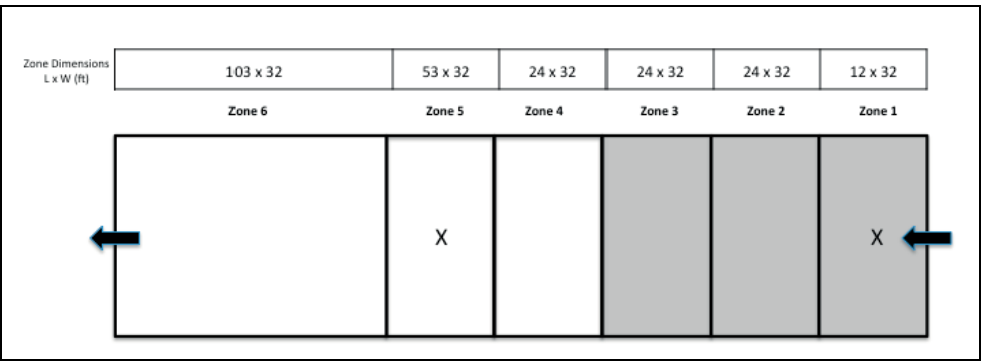
PLUG-FLOW 2



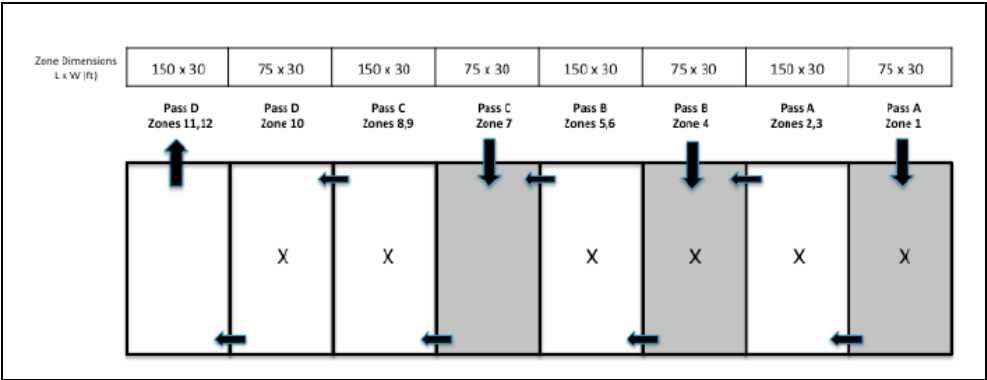
MLE 1



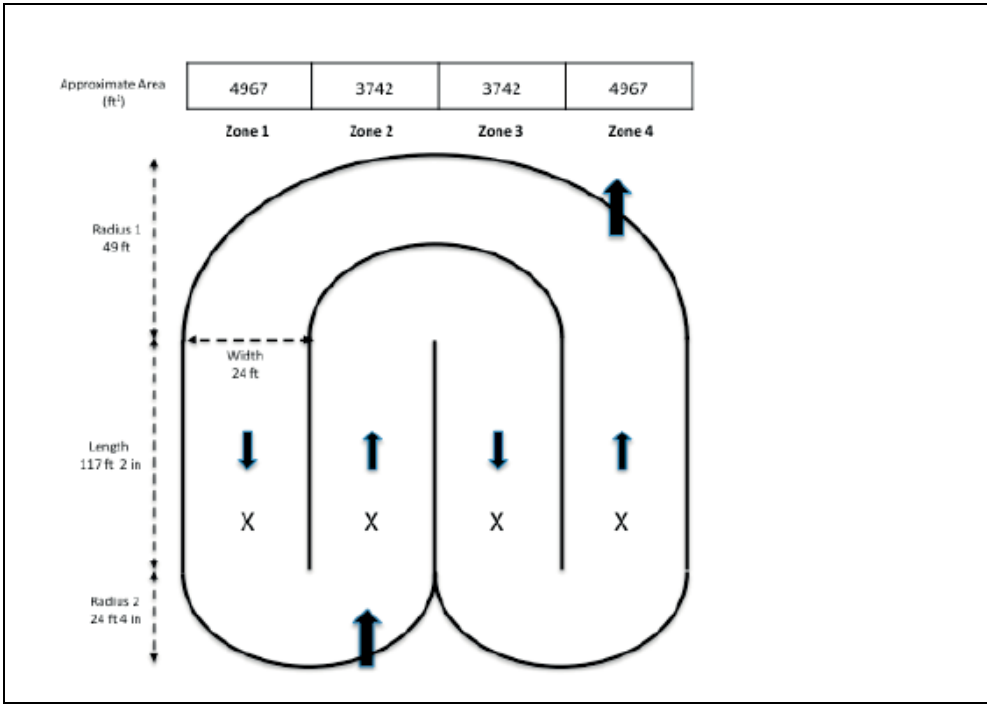
MLE 2



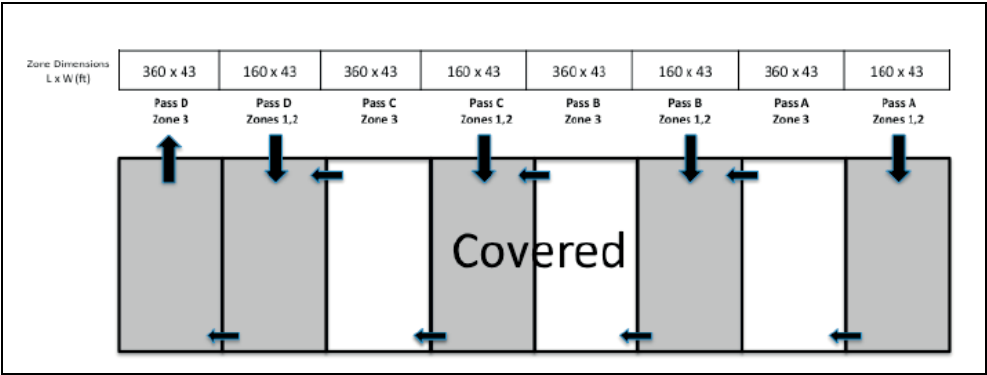
STEP-FEED BNR 2



OXIDATION DITCH



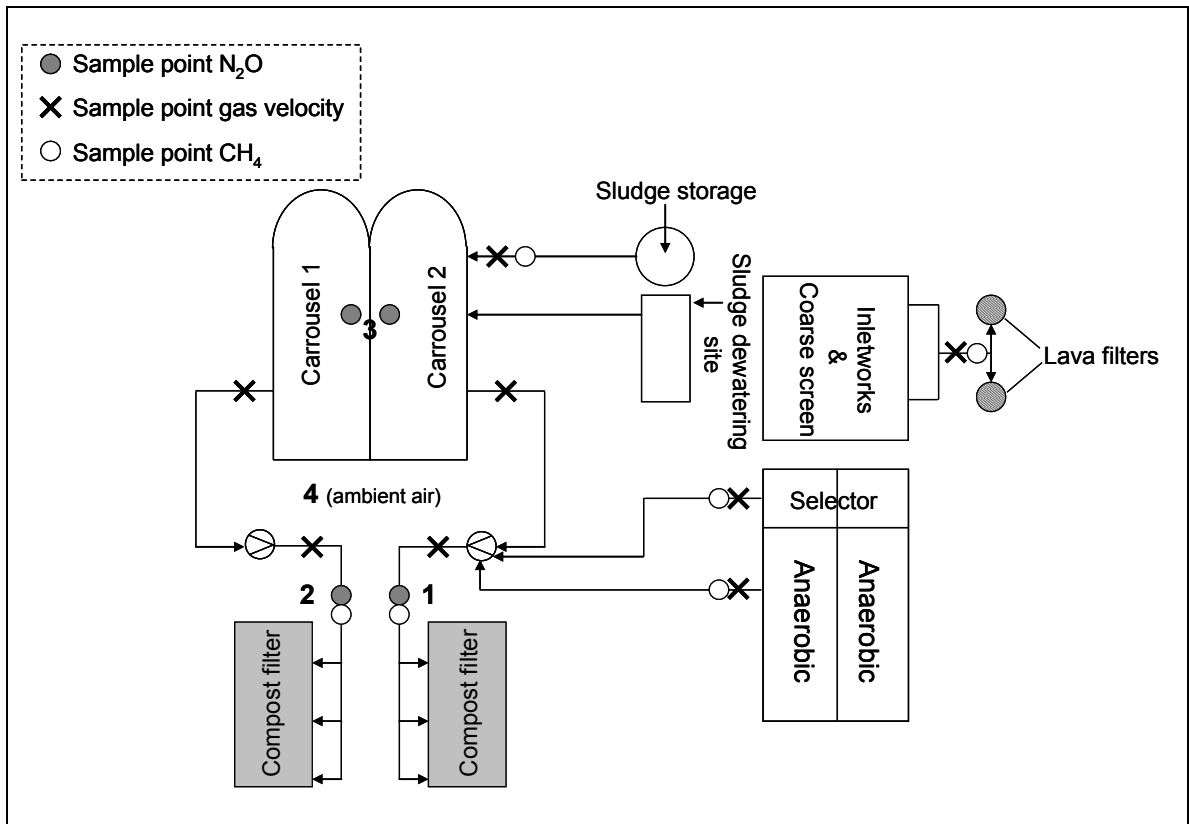
STEP-FEED BNR 3



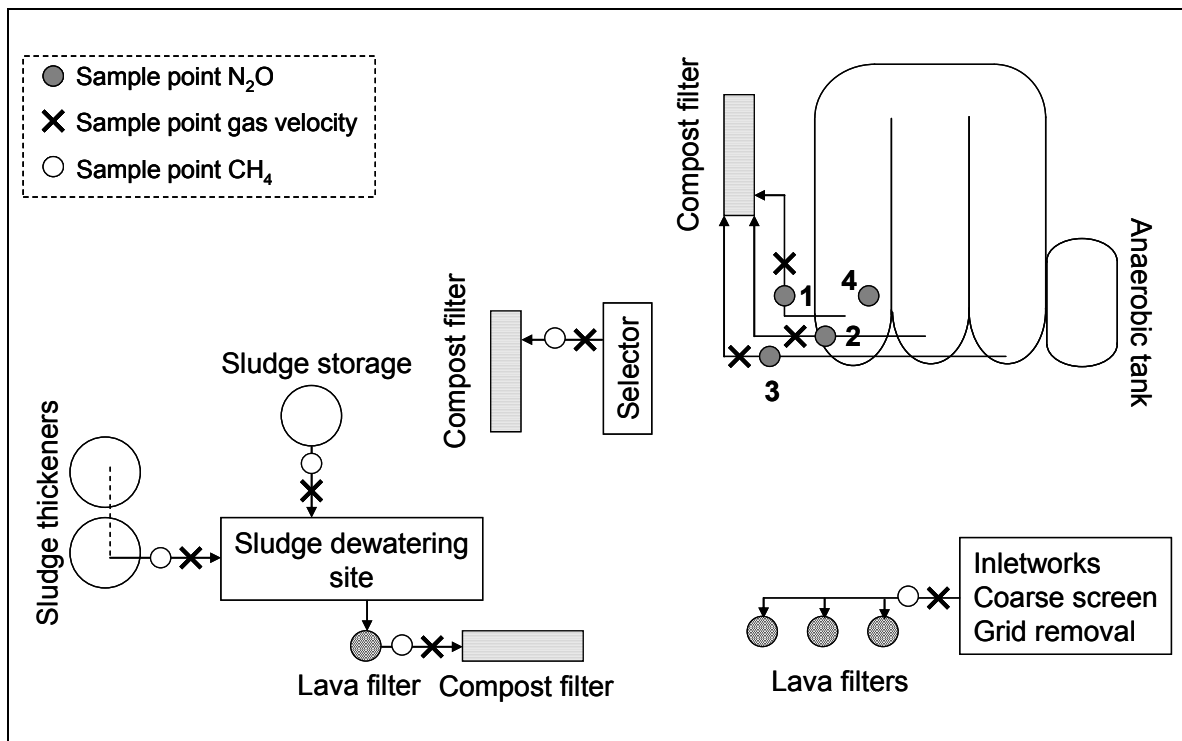
APPENDIX 3

LOCATIONS OF N₂O AND CH₄ MEASUREMENTS IN THE NETHERLANDS

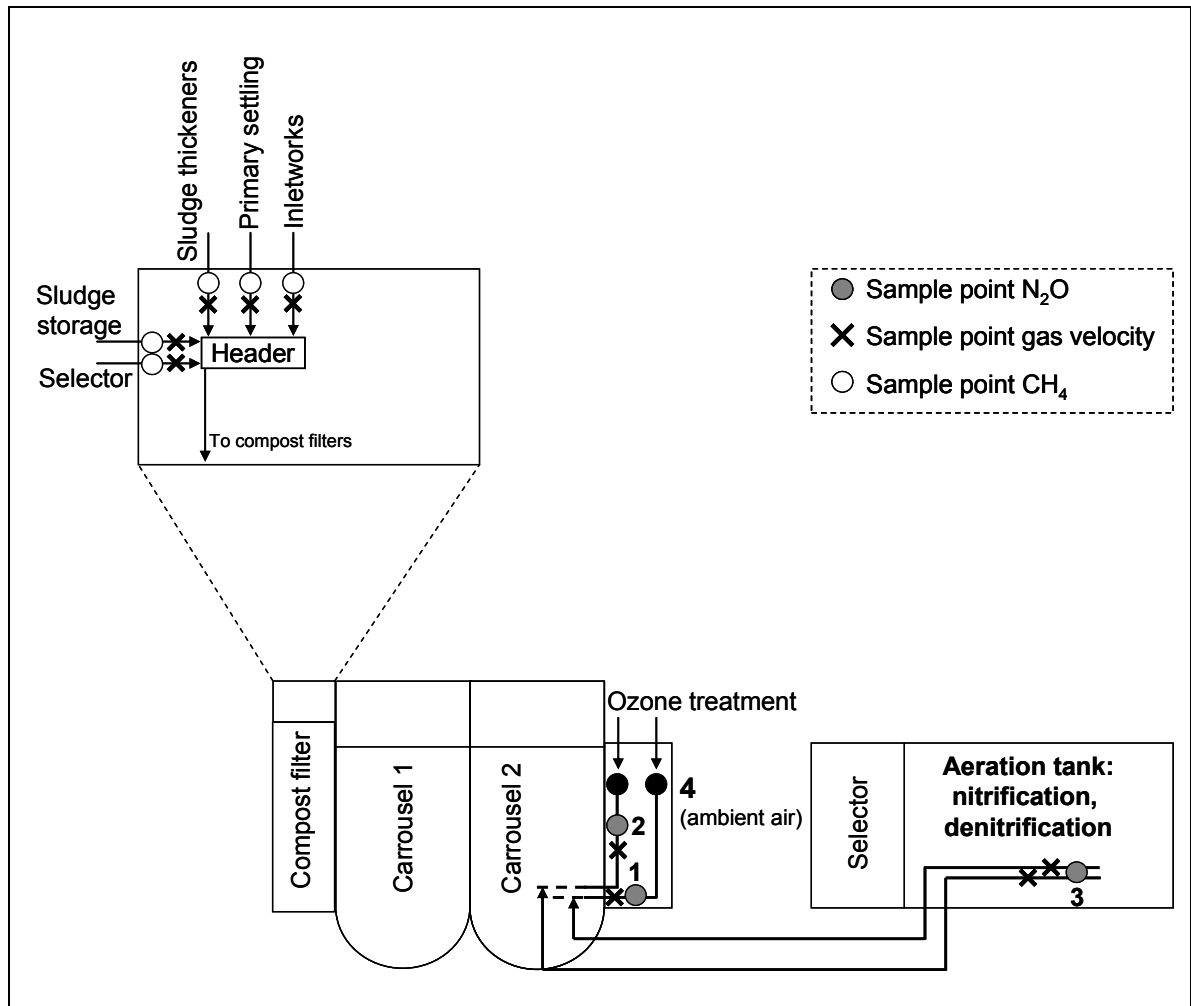
WWTP PAPENDRECHT



WWTP KORTENOORD



WWTP KRALINGSEVEER



APPENDIX 4

ABBREVIATIONS AND GLOSSARY

Abbreviation / term	
A/O and A ² /O	Type of WWTP configuration for the biological removal of phosphate ¹⁾
ADEME	French Environment and Energy Management Agency
Bardenpho	Type of WWTP configuration for the biological removal of phosphate ¹⁾ Can be designed with different amount of stages
BNR	Biological Nitrogen Removal
CH ₄	Methane
CO ₂	Carbon Dioxide
COD	Chemical Oxygen Demand
DO	Dissolved Oxygen
EPA	Environmental Protection Agency
FB	Fermentative Bacteria
Fe ^{2+/3+}	Iron
FID	Flame Ionization Detector
GC	Gas Chromatograph
GF _{WWTP}	Generation factor (N ₂ O) for whole WWTP
GHG	Greenhouse Gas
H ₂ S	Hydrogen Sulphide
HRT	Hydraulic Retention Time
IPCC	Intergovernmental Panel on Climate Change
Johannesburg	Type of WWTP configuration ¹⁾
K _L a	Mass transfer coefficient (liquid → gas)
LCC - PS	Lower Crooked Creek Pump Station
MBR	Membrane Bioreactor
MLE	Modified Ludzack-Ettinger (i.e. a configuration type)
NH ₄ ⁺ (-N)	Ammonium
NGERS	National Greenhouse and Energy Reporting System
NO ₂ ⁻ (-N)	Nitrite
NO ₃ ⁻ (-N)	Nitrate
N ₂ O	Nitrous Oxide
POTW	Public Owned wastewater treatment
SBR	Sequenced Batch Reactor
SRB	Sulphate Reducing Bacteria
SRT	Sludge retention time
STOWA	Foundation for Applied Water Research
TKN	Total Kjeldahl Nitrogen
Ton (USA)	US ton is 2,000 pounds (907 kg)
USEPA	United States Environmental Protection Agency
UKWIR	UK Water Industry Research
VFA	Volatile Fatty Acids
WERF	Water Environment Research Foundation
WSAA	Water Service Association of Australia
WWTP	Wastewater Treatment Plant

1) For more information see: Tchobanoglous, G., 2003

APPENDIX 5

NATIONAL GREENHOUSE AND ENERGY REPORTING SYSTEM, AUSTRALIA

FACILITY DEFINITION

Of particular relevance to water industry is the definition of a “facility”, with respect to water supply, sewerage or drainage services. The provision from the NGER Regulations (see the [box](#) below) would treat all of the activities undertaken as part of a water supply, sewerage and drainage services network as a single undertaking or enterprise and therefore one facility for the purposes of the Act’s threshold.

In 2009, the Water Services Association of Australia (WSAA) requested a determination from the DCCEE on an alternative definition of the boundaries of “facilities” for the water supply, sewerage and drainage services sector. This submission, at a minimum, sought to separate “water facilities” into two separate continuous systems:

- 1 From water source and extraction, through treatment and distribution; and
- 2 Sewage collection, sewage treatment and effluent discharge.

These sub-systems would then be further disaggregated, based on separate physical catchments or distribution zones. Allocation of emissions to different corporate entities will then depend on who has “operational control” of the various elements of the “facility”. Presently, the DCCEE has advised that they will not make a formal response to WSAA’s request.

2.20 *Forming part of a single undertaking or enterprise — electricity, gas etc activities*

(1) *If:*

- (a) *an activity in a series of activities is in one of the industry sectors mentioned in sub-regulation (2); and*
- (b) *the activity and any ancillary activities to it are under the overall control of the same corporation;*

then all of the activities will form part of the same single undertaking or enterprise.

(2) *For sub-regulation (1), the industry sectors are as follows:*

- (a) *Electricity transmission (262);*
- (b) *Electricity distribution (263);*
- (c) *Gas supply (270);*
- (d) *Water supply, sewerage and drainage services (281);*

CHOICE OF CALCULATION METHOD

The NGERS *Technical Guidelines 2009* provide Methods that broadly allow for both direct emissions monitoring and the estimation of emissions through the tracking of observable, closely-related variables. Four Methods have been described which provide a framework for emissions estimation for a range of purposes:

- **Method 1:** the National Greenhouse Accounts default method:
 - Method 1 provides a class of estimation procedures derived directly from the methodologies used by the Department of Climate Change for the preparation of the *National Greenhouse Accounts* ⁽¹⁷⁾. This is the most basic of the available methods.
- **Method 2:** a facility-specific method using industry sampling and Australian or international standards listed in the Determination to provide more accurate estimates of emissions at the facility level:
 - Method 2 enables corporations to undertake additional measurements in order to gain more accurate estimates for emissions for that particular facility.
- **Method 3:** a facility-specific method using Australian or international standards listed in the Determination or equivalent standards for both sampling and analysis:
 - Method 3 is very similar to Method 2, except that it requires, additionally, reporters to comply with Australian or international documentary standards for sampling, as well as documentary standards for the analyses.
- **Method 4:** direct monitoring of emission systems, either on a continuous or periodic basis:
 - Method 4 provides approaches to enable direct monitoring of greenhouse gas emissions arising from an activity. This can provide a higher level of accuracy in certain circumstances, depending on the type of emission process although it is more data intensive than other approaches.
 - There is presently no Method 4 defined for wastewater handling emissions.

The provision to select a Method for the estimation of emissions allows organisations to make their own judgement on the balance of costs of using the higher-order methods, with the benefits of potentially improved emission estimates. Experience suggests that the use of Method 2 for estimating emissions related to wastewater treatment generally results in approximately 15% lower results when compared to results calculated using Method 1. Therefore, where sufficient activity data is readily available, it is generally advisable to use Method 2, as it is less likely to over-estimate emissions than Method 1.

METHODOLOGY FOR ESTIMATING CH₄ EMISSIONS

Presently, there is no NGERS methodology to estimate fugitive greenhouse gas emissions arising from methane generation in the sewerage network, upstream of wastewater treatment facilities. The NGERS *Technical Guidelines 2009* promulgates the following equations for the estimation of fugitive CH₄ emissions arising from wastewater treatment and handling:

¹⁷ This involves estimation of COD and nitrogen loads into the plant based on the following default factors: A default quantity of COD per capita of 0.0585 tonnes per person per year; and a default nitrogen load of $0.16 \times 0.036 = 0.00576$ tonnes per person per year.

EQUATION 1

(NGERS TECHNICAL GUIDELINES, DIVISION 5.2.3 SECTION 5.25 SUB-SECTION (1)):

$$E_j = CH_4^* - \gamma(Q_{cap} + Q_{flared} + Q_{tr})$$

where:

- E_j is the emissions of CH₄ released by the plant during the year, measured in CO₂-e tonnes;
- CH_4^* is the estimated quantity of CH₄ in biogas released by the plant during the year, measured in CO₂-e tonnes;
- γ is the factor $6.784 \times 10^{-4} \times 21$ converting cubic metres of CH₄ at standard conditions (15°C, 1 atm) to CO₂-e tonnes;
- Q_{cap} is the quantity of CH₄ in biogas captured for combustion for use by the plant during the year, measured in cubic metres;
- Q_{flared} is the quantity of CH₄ in biogas flared during the year by the plant, measured in cubic metres; and
- Q_{tr} is the quantity of CH₄ in biogas transferred out of the plant during the year, measured in cubic metres.

The intent of Equation 1 is to reconcile the estimated **theoretical** amount of CH₄ generated at the wastewater treatment plant (CH_4^*) with the actual amount of CH₄ **measured** in the captured biogas (Q_{cap} , Q_{flared} and Q_{tr}). Any difference between these two figures, and assuming $CH_4^* > \gamma(Q_{cap} + Q_{flared} + Q_{tr})$, is hence assumed to equal the amount of “fugitive” CH₄ emitted to the atmosphere.

EQUATION 2

(NGERS TECHNICAL GUIDELINES, DIVISION 5.2.3 SECTION 5.25 SUB-SECTION (5)):

$$CH_4^* = (COD_w - COD_{sl} - COD_{eff}) \times F_{wan} \times EF_{wij} + (COD_{sl} - COD_{trl} - COD_{tro}) \times F_{slan} \times EF_{slij}$$

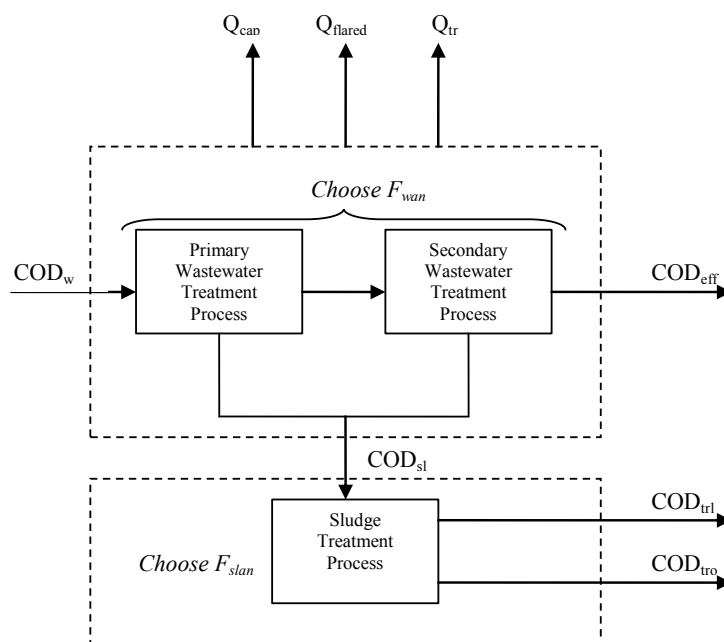
where:

- COD_w is the quantity of COD in wastewater entering the plant during the year, measured in tonnes;
- COD_{sl} is the quantity of COD removed as sludge from wastewater and treated in the plant, measured in tonnes of COD;
- COD_{eff} is the quantity of COD in effluent leaving the plant during the year, measured in tonnes;
- F_{wan} is the fraction of COD anaerobically treated by the plant during the year;
- EF_{wij} is the default CH₄ emission factor for wastewater with a value of 5.3 CO₂-e tonnes per tonne COD;
- COD_{trl} is the quantity of COD in sludge transferred out of the plant and removed to landfill, measured in tonnes of COD;
- COD_{tro} is the quantity of COD in sludge transferred out of the plant and removed to a site other than landfill, measured in tonnes of COD;
- F_{slan} is the fraction of COD in sludge anaerobically treated by the plant during the year.
- EF_{slij} is the default CH₄ emission factor for sludge with a value of 5.3 CO₂-e tonnes per tonne COD (sludge).

A simple process flow diagram is included in Figure 50 to help clarify the definition of terms in Equation 2.

FIGURE 50

DEFAULT NGERS COD MASS BALANCE



The intent of Equation 2 is to estimate the theoretical quantity of CH₄ generated at the plant, based on a simple COD mass balance. The equation is divided into two parts. The first part is a simple COD mass balance over the main wastewater treatment processes (i.e. “liquid train” primary and secondary treatment). The second part is a simple COD mass balance over the sludge treatment processes (e.g. anaerobic digesters, sludge lagoons, dewatering, etc.).

Each COD mass balance determines how much COD is consumed in the treatment process by converting it to a gas (i.e. CO₂ or CH₄). Multiplying by the maximum CH₄ generation factor (EF) calculates how much CH₄ would be generated if **100%** of the COD consumed was converted to CH₄. Then, multiplying by the process-specific F_{wan} or F_{slan} factor calculates the expected fraction of CH₄ emissions for that particular type of process.

CLARIFICATION ON FWAN AND FSLAN FACTORS

The NGERS definition of F_{wan} as “the fraction of COD anaerobically treated by the plant during the year” and of F_{slan} as “the fraction of COD in sludge anaerobically treated by the plant during the year” can be misleading and confusing.

To be consistent with Intergovernmental Panel on Climate Change (IPCC) terminology and to provide a better description of its true meaning, F_{wan} can be re-defined as:

“MCFw – the CH₄ correction factor for wastewater treatment, or fraction of the maximum CH₄ emission factor”.

The NGERS definition of EF_{wij} can also be redefined, to be consistent with IPCC terminology, as “the maximum CH₄ generation factor for wastewater”. This value is defined by the chemical stoichiometry of CH₄ production. In practice however, this maximum is not achieved (even in 100% anaerobic systems) due to some (or all) of the following losses:

- Some COD may be consumed via other biochemical pathways (e.g. sulphide reduction, volatile acids production);
- Some CH₄ that is generated may be consumed by aerobic methanotrophic microorganisms and converted to carbon dioxide (i.e. through surface layer oxidation in uncovered lagoons); and
- Some CH₄ that is generated may remain dissolved in the liquid phase (NOTE: dissolved CH₄ is not accounted for in standard COD analysis ⁽¹⁸⁾).

These losses are accounted for by the F_{wan} factor. For example, the default NGERS F_{wan} value of 0.8 for anaerobic deep lagoons accounts for the typical CH₄ losses in open anaerobic lagoon systems. This equates to 80% of consumed COD being converted to gaseous CH₄ as an end product, with the balance of 20% consumed via other pathways or remaining in its soluble form (and therefore not adding to overall atmospheric CH₄ emissions).

Similarly, to be consistent with IPCC terminology and to provide a better description its true meaning, F_{slan} can also be redefined as:

“MCFsl – the CH₄ correction factor for sludge treatment, or fraction of the maximum CH₄ emission factor”.

EF_{slj} can also be redefined as “the maximum CH₄ generation factor for sludge”.

Based on verbal discussions with the DCCEE, the formal definitions of F_{wan} and F_{slan} are likely to change, in line with these recommendations, in the next major revision of the NGERS *Determination and associated Technical Guidelines* in 2011.

The NGERS *Technical Guidelines* include IPCC default F_{wan} and F_{slan} factors (equivalent to IPCC MCFs) for various types of treatment systems. These factors have been compiled by the IPCC and are based on “expert judgement by lead authors of this section” ⁽¹⁹⁾.

The IPCC Guidelines also include ranges for each MCF to account for the variation that is likely to be seen in practice for individual treatment systems. These ranges (shown in Table 27 below) are not promulgated under NGERS.

TABLE 27

NGERS DEFAULT CORRECTION FACTORS AND IPCC RANGES FOR SOME TREATMENT SYSTEMS

Treatment System	F_{wan} or F_{slan} (NGERS)	IPCC Range
Managed Aerobic Treatment	0	0 – 0.1
Unmanaged Aerobic Treatment	0.3	0.2 – 0.4
Anaerobic Digester / Reactor	0.8	0.8 – 1.0
Anaerobic Shallow Lagoon (< 2 m deep)	0.2	0 – 0.3
Anaerobic Deep Lagoon (> 2 m deep)	0.8	0.8 – 1.0

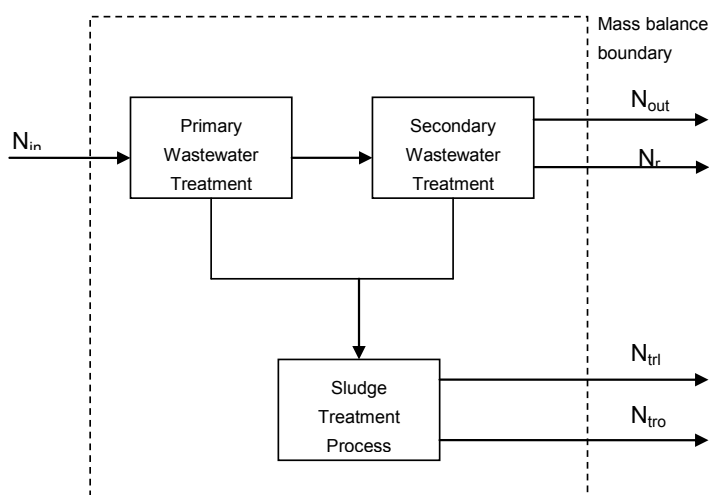
The general approach to calculating N₂O emissions from wastewater treatment processes, under the NGERS Technical Guidelines 2009 is outlined below in Table 28.

- 18 Hartley, K., Lant, P. (2006). Eliminating non-renewable CO₂ emissions from sewage treatment: An anaerobic migrating bed reactor pilot plant study, *Biotechnology and Bioengineering*, 95(3): 384-398.
- 19 IPCC, (2006). *2006 IPCC Guidelines for National Greenhouse Gas Inventories*, Ch.6 – Wastewater Treatment and Discharge, Table 6.3 p. 6.13.

TABLE 28 NGERS METHODOLOGY FOR ESTIMATING NITROUS OXIDE EMISSIONS.

NGERS Technical Guidelines 2009 Part 5.3, Division 5.3.6 – Method 2
reference

Simplified generic mass balance diagram



Typical activity data required (units)

- Raw wastewater volume, Q (ML)
- Effluent reused by third parties, Q_r (ML)
- Mass of biosolids sent to landfill, M_{trl} and/or “other”, M_{tro} (e.g. land application) (tonnes)
- Influent total nitrogen concentration, [N]_{in} (mgN/L)
- Effluent total nitrogen concentration, [N]_{out} (mgN/L)
- Effluent reused by third parties total nitrogen concentration, [N]_r (mgN/L)
- Nitrogen content of biosolids, [N]_b (mgN/kg biosolids)

Emission factors, exclusions and assumptions

- Technical Guidelines 2009, Division 5.3.5, section 5.31 (3):
- Emission factor for secondary treatment, EF_{secij} = 4.9 tCO₂-e per tonne N removed by denitrification (1);
- Emission factor for nitrogen discharge differentiated by the discharge environment, EF_{disij} = 4.9 tCO₂-e per tonne N discharged (2);
- **One emission factor, EF_{secij}, applies for ALL processes within a treatment plant, and for ALL types of treatment plants**
- **One emission factor, EF_{disij}, applies for ALL types of nitrogen discharges**

Emissions calculation method

- Raw wastewater total N mass load, N_{in} (tonnes)
- = $Q \cdot [N]_{in} \cdot (10^{-3} \text{ t/kg})$
- Discharged effluent total N mass load, N_{out} (tonnes)
- = $(Q - Q_r) \cdot [N]_{out} \cdot (10^{-3} \text{ t/kg})$
- Total N mass load in water reuse to third parties, N_r (tonnes)
- = $Q_r \cdot [N]_r \cdot (10^{-3} \text{ t/kg})$ or $Q_r \cdot [N]_{out} \cdot (10^{-3} \text{ t/kg})$
- N mass load to landfill, N_{trl} (tonnes)
- = $M_{trl} \cdot [N]_b \cdot (10^{-3} \text{ t/kg})$
- N mass load to “other” (e.g. land application), N_{tro} (tonnes)
- = $M_{tro} \cdot [N]_b \cdot (10^{-3} \text{ t/kg})$
- Nitrogen mass balance, MBN_{ww} (tonnes) (i.e. mass of nitrogen removed by denitrification)
- = $(N_{in} - N_{out} - N_r - N_{trl} - N_{tro})$
- Greenhouse gas emissions (tCO₂-e)
- = $MBN_{ww} \cdot EF_{secij} + N_{out} \cdot EF_{disij}$

20 Derivation of EF_{secij}: 10 kg N₂O-N per tonne N removed x (44 kg N₂O per 28 kg N) x (10⁻³ tonnes / kg) x (310 tonne CO₂-e per tonne N₂O) = 4.9 tonne CO₂-e per tonne N removed

21 Derivation of EF_{disij}: 10 kg N₂O-N per tonne N discharged x (44 kg N₂O per 28 kg N) x (10⁻³ tonnes / kg) x (310 tonne CO₂-e per tonne N₂O) = 4.9 tonne CO₂-e per tonne N discharged

This nitrogen mass balance framework for NGERS calculations is theoretically quite robust, but it is presently limited by the use of a single emission factor for EF_{secij} and EF_{disij} , hence lacking the flexibility to account for different levels of N₂O emissions from different processes and/or operating conditions or effluent discharges.

NGERS UNCERTAINTY

Reporting of uncertainty associated with Scope 1 emissions has been voluntary under NGERS for the first two reporting years. However, the NGERS *Technical Guidelines 2009* now provide extensive guidance on uncertainty estimation in Chapter 8. This guidance is based on the *GHG Protocol Guidance on Uncertainty Assessment in GHG Inventories and Calculating Statistical Parameter Uncertainty*, 2003.

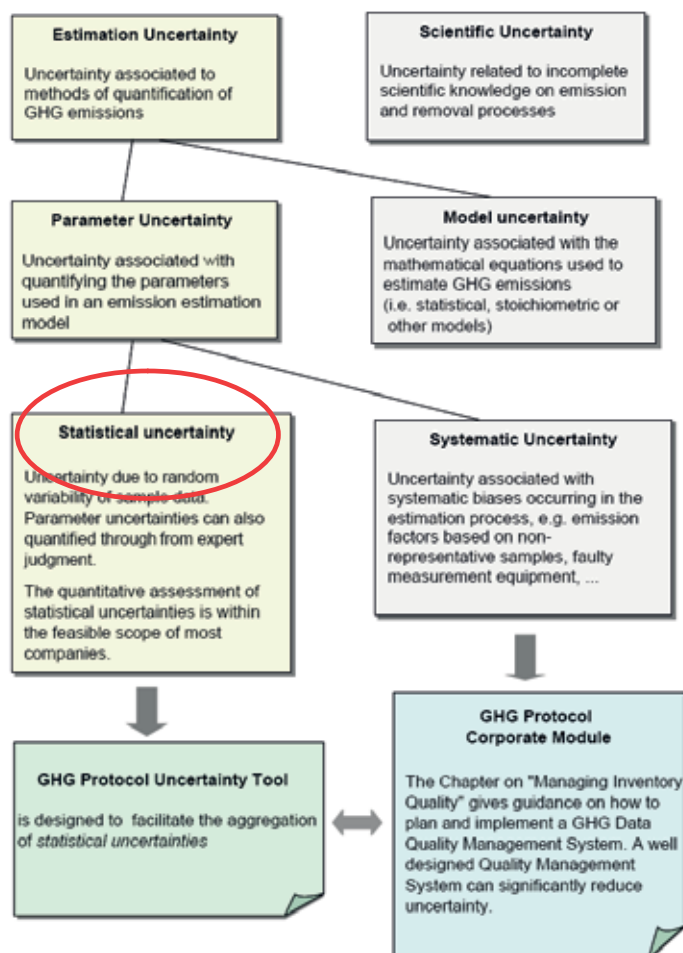
Figure 51 below shows the types of uncertainty that are associated with the estimation of greenhouse gas emissions. For the purposes of NGERS reporting, it is only necessary to calculate **statistical uncertainty** – that is, the uncertainty due to the underlying random variability of activity data. It is not necessary to account for systematic uncertainty, such as instrumental error, sampling error or analytical error. Furthermore, it is not necessary to account for the model uncertainty and scientific uncertainty that might be associated with the default emission factors. Hence, the uncertainty calculated for NGERS reporting purposes only represents a fraction of the true uncertainty associated with the results.

Wastewater treatment processes are inherently variable, both in terms of throughput quantities (i.e. flow rates) and qualities (i.e. constituent concentrations), as well as biological performance. Hence, just capturing the statistical uncertainty associated with wastewater handling processes is very difficult. Furthermore, the need to reconcile two completely independent data sets (i.e. Equation 1 above) is also statistically challenging, given that they both have high underlying variability.

FIGURE 51

TYPES OF UNCERTAINTY ⁽²²⁾

Types of Uncertainties associated with greenhouse gas inventories



Section 8.10 of the NGRS Technical Guidelines 2009 allows for a default aggregated uncertainty of 40% for domestic wastewater handling. However, by applying the statistical analyses recommended under Chapter 8, it may be possible to refine this uncertainty estimate, based on actual statistical confidence in the activity data (refer section 2.1.8 below).

22 GHG Protocol Guidance on Uncertainty Assessment in GHG Inventories and Calculating Statistical Parameter Uncertainty, 2003

CALCULATION METHOD FOR NGERS UNCERTAINTY

The NGERS *Technical Guidelines* assumes that each activity data set required for calculations in Equations 1 and 2 is normally distributed. Given this assumption, the statistical uncertainty of each data set can be represented by its arithmetic mean \pm 95% confidence interval (assuming the Student's *t*-distribution). The 95% percent confidence interval is calculated as follows:

$$\bar{x} \pm t\left(\frac{\alpha}{2}; n-1\right) \frac{s}{\sqrt{n}}$$

where:

- \bar{x} is the arithmetic mean of the sampling data;
- t is the Student's *t*-distribution;
- α is equal to 0.05 for a 95% confidence interval;
- n is the number of samples in the data set;
- s is the standard deviation of the data set.

The calculations required in Equations 1 and 2 involve addition/subtraction and multiplication/division of these various activity data sets. Combining the statistical uncertainties through these mathematical manipulations is done via a simple first order propagation technique:

- Multiplying/dividing uncertainties:
- where: $(A \pm a\%) \times (B \pm b\%) = C \pm c\%$,
- $c = \sqrt{a^2 + b^2}$
- Adding/subtracting uncertainties:
- where: $(C \pm c\%) + (D \pm d\%) = E \pm e\%$

$$e = \frac{\sqrt{(C \times c)^2 + (D \times d)^2}}{E}$$

APPENDIX 6

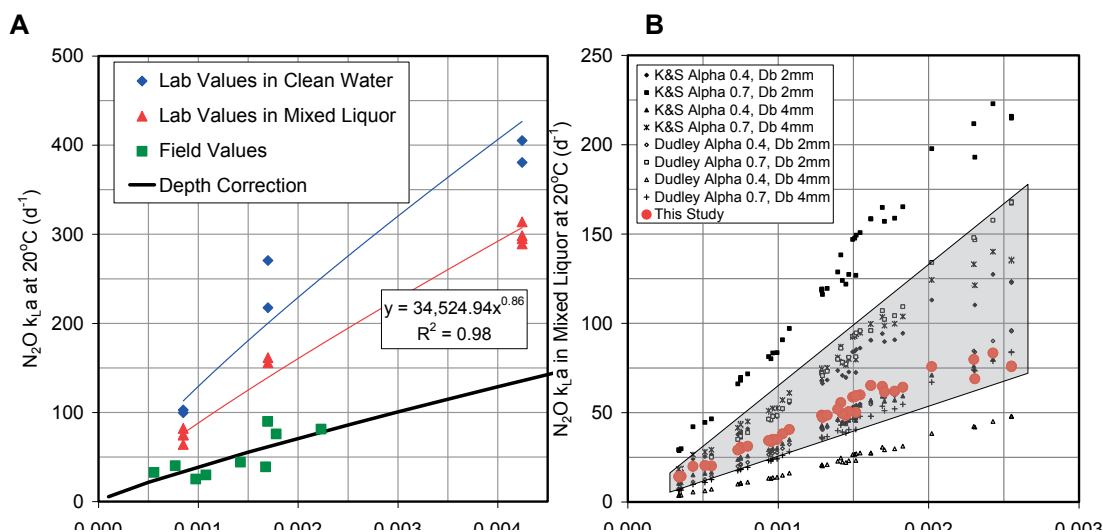
DETERMINATION OF $K_L A$ FOR N₂O EMISSION, AUSTRALIA

To determine the N₂O mass transfer coefficient ($k_L a$) for each WWTP reactor zone (refer to Equation 5 in section 5.1.3), a series of lab-scale stripping experiments was conducted. These experiments were conducted in both clean tap water and mixed liquor (from WWTP No.1, MLSS 4.45 g.L⁻¹) in a 0.05 m diameter, 0.815 m deep glass column, with a gas sparging bar at its base. Measurements of dissolved N₂O and dissolved oxygen were made at two locations in the liquid column using an on-line Membrane Inlet Mass Spectrometer (Hidden Analytical, Warrington, United Kingdom).

In each stripping experiment, the liquid column was initially saturated by bubbling through 0.51% N₂O gas (in helium), which equated to an equilibrium dissolved N₂O concentration of 6.45 mg.L⁻¹ at 20°C (Weiss and Price, 1980), and dissolved oxygen equal to zero. The sparge gas was then switched to compressed air, and the dissolved concentration of O₂ and N₂O monitored until oxygen reached saturation (9.07 mg.L⁻¹ at 20°C, corrected for chlorinity) (APHA, 1995) and N₂O approached zero. This procedure was repeated for clean water and mixed liquor at three different sparge flowrates (500 mL.min⁻¹, 200 mL.min⁻¹ and 100 mL.min⁻¹). In the mixed liquor stripping experiments, the clean water saturation concentrations were reduced by a 95% β correction factor to account for differences in solubility caused by constituents in the mixed liquor such as salts, particulates and surface-active substances (Tchobanoglous et al., 2003).

The N₂O mass transfer coefficient for each experiment was then calculated using a non-linear parameter estimation routine (Stenstrom et al., 1997). The results of these calculations are shown in Figure 52A. As expected, the mass transfer coefficient decreased with superficial gas velocity, v_g (m³.m⁻².s⁻¹), in a power law relationship. This result is similar to other empirical mass transfer modelling approaches for aerated systems, such as bubble columns (Envirosim, 2007; Garcia-Ochoa and Gomez, 2009)

FIGURE 52 A) NITROUS OXIDE VOLUMETRIC MASS TRANSFER COEFFICIENTS ($K_L A$) FROM CLEAN WATER AND MIXED LIQUOR STRIPPING EXPERIMENT IN A LAB-SCALE COLUMN, AND FROM FIELD MEASUREMENTS; B) COMPARISON OF MODELLED $K_L A$ VALUES FOR ALL WWTPS (EQUATION 9) AGAINST LITERATURE CORRELATIONS FOR A RANGE OF BUBBLE DIAMETERS, D_b AND αF FACTORS. THE SHADED AREA INDICATES THE 95% CONFIDENCE INTERVAL ADOPTED FOR THE $K_L A$ VALUES IN EQUATION 5 IN SECTION 5.1.3



However, the volumetric mass transfer coefficient is also related to several other variables, including reactor geometry (particularly aerator immersion depth), aeration bubble size, diffuser layout and liquid viscosity (Gillot et al., 2005). To better correlate the lab-scale bubble column results to full-scale reactors that are significantly deeper (i.e. 3.68 – 5.95 m), six off-gas samples were collected from the aerated zones of WWTP No.5 (depth = 4.5 m), two off-gas samples from WWTP No. 3 (depth = 3.9 m) and one sample from WWTP No. 6 (depth = 4.7 m) (refer to paragraph 5.1.1). For each sample, the mass transfer coefficient was then calculated as per Equation A1:

$$k_L a_F = \frac{\frac{Q_A \times p_{N_2O,G}}{R \times T} \times MW_{N_2O}}{V_R \times ([N_2O]_R - [N_2O]_S^*)} \quad (A1)$$

Where:

- $k_L a_F$ = volumetric mass transfer coefficient, calculated from field data (d⁻¹)
- Q_A = aeration flowrate supplied to reactor at the time of sampling (m³.d⁻¹)
- $p_{N_2O,G}$ = partial pressure of N₂O in off-gas from reactor (atm)
- MW_{N_2O} = molecular weight of N₂O (44 kg.kmol⁻¹)
- R = universal gas constant (0.08206 m³.atm.kmol⁻¹.K⁻¹)
- T = reactor liquid temperature at time of sampling (K)

The results for these field $k_L a$ estimates are also shown in Figure 52A, and are lower than the mass transfer coefficients measured for the shallower lab-scale column. Therefore, the power law estimation based on v_g only, from the lab-scale mixed liquor experiments (refer to Figure 52) was modified by the addition of a correction factor

$$\left\{ \frac{D_R}{D_L} \right\}^{\kappa}, \text{ to account for the increased depth of full-scale WWTP reactors. The value of } \kappa$$

was empirically determined using a sum-of-least-squares fitting algorithm in MS Excel. The resulting depth-corrected $k_L a$ correlation is shown in Equation A2:

$$k_L a_F^* = \left\{ \frac{D_R}{D_L} \right\}^{-0.49} \times (34,500 \times v_g)^{0.86} \quad (A2)$$

Where:

- D_R = depth of the field reactor (m)
- D_L = depth of the lab stripping column (0.815 m)
- v_g = superficial gas velocity of the field reactor (m³.m⁻².s⁻¹)

Using Equation 9, the N₂O volumetric mass transfer coefficient was calculated for all aerated reactor zones in the seven WWTPs. To determine the uncertainty associated with these field $k_L a$ values, two independent mass transfer correlation techniques were applied to the field data. The first correlation technique (Khudenko and Shpirt, 1986) empirically related the clean water oxygen mass transfer coefficient ($k_L a_{O_2}$) to v_g , reactor geometry and aeration bubble diameter, D_B . The N₂O mass transfer coefficient ($k_L a_{N_2O}$) was then estimated in

accordance with Higbie's penetration model (Capela et al., 2001):

$$k_L a_{N_2O} = k_L a_{O_2} \times \sqrt{\frac{D_{F,N_2O}}{D_{F,O_2}}} \quad (A3)$$

Where:

$$\begin{aligned} D_{F,N_2O} &= \text{Molecular diffusivity of } N_2O \text{ in water} \\ &= 1.84 \times 10^{-9} \text{ m}^2 \cdot \text{s}^{-1} \text{ at } 20^\circ\text{C} \text{ (Tamimi et al., 1994)} \\ D_{F,O} &= \text{Molecular diffusivity of oxygen in water} \\ &= 1.98 \times 10^{-9} \text{ m}^2 \cdot \text{s}^{-1} \text{ at } 20^\circ\text{C} \text{ (Ferrell and Himmelblau, 2002)} \end{aligned}$$

The second correlation (Dudley, 1995) empirically related the clean water N₂O k_La to v_G, D_{F,N₂O}, D_B and viscosity. Both correlations were also corrected for temperature, using a standard θ factor of 1.024 (Tchobanoglous et al., 2003):

$$k_L a_{T^\circ C} = k_L a_{20^\circ C} \times \theta^{T-20} \quad (A4)$$

Both correlations are sensitive to bubble diameter, which was unknown for all WWTPs. Furthermore, the clean water k_La values must also be corrected for mixed liquor conditions and diffuser fouling (i.e. αF factor), which was also unknown. Therefore, a range of likely values was evaluated for both parameters: DB = 2 – 4mm (Hasanen et al., 2006) and αF = 0.4 – 0.9 (Tchobanoglous et al., 2003). The results of these sensitivity analyses are shown in Figure 52B for the two correlations. It can be seen that the k_La model proposed for this study (Equation 9) fits well within the band of likely values and thus provides an independent validation of the adopted approach. The correlations also provide an uncertainty range for the k_La value used in Equation 5 (section 5.1.3). Using the full range of sensitivity analysis values calculated from the two correlations, a 95% confidence interval (t-dist, α = 0.05) of k_La values was calculated (i.e. shaded area in Figure 52B). This confidence interval was then applied to each k_La determined by Equation A2. From Figure 52B, it can be seen that the values from Equation A2 sit in the lower quartile of the confidence interval generated from the literature correlations.

The mass transfer coefficients for quiescent reactor zones (i.e. primary sedimentation tanks, anaerobic zones, anoxic zones, secondary sedimentation tanks) were estimated using the empirical correlation technique of Van't Riet (1979). This relates k_La to the volumetric power input (P/V) for mixing. Similar to the approach for aerated zones, a range of likely P/V values was surveyed (2 – 8 W.m⁻³) and used to construct a 95% confidence interval for each field reactor zone. However, the k_La values in these quiescent zones (indicatively 3 – 4 d⁻¹) were an order of magnitude smaller than those in the aerated zones. Therefore, the results are not especially sensitive to these values.

ANNEX 7

WWTPS FRANCE

FIGURE 53 SCHEME OF WWTP 1

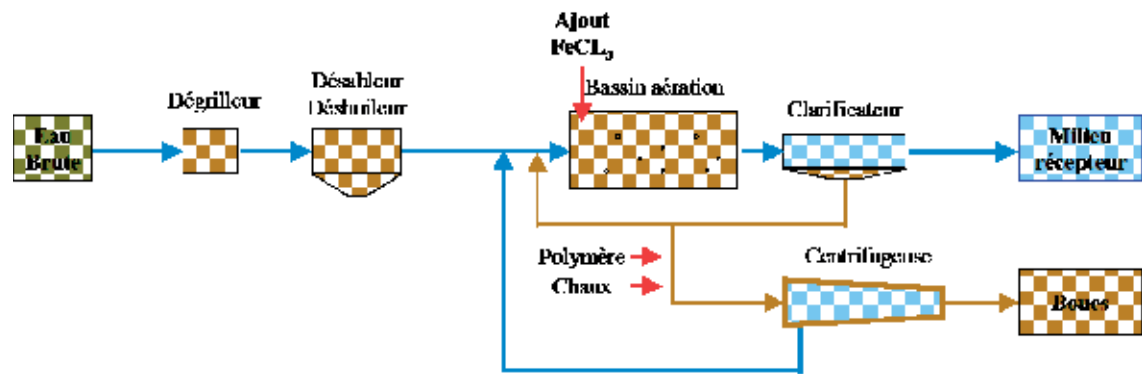


FIGURE 54 SCHEME AND PHOTO OF WWTP 2

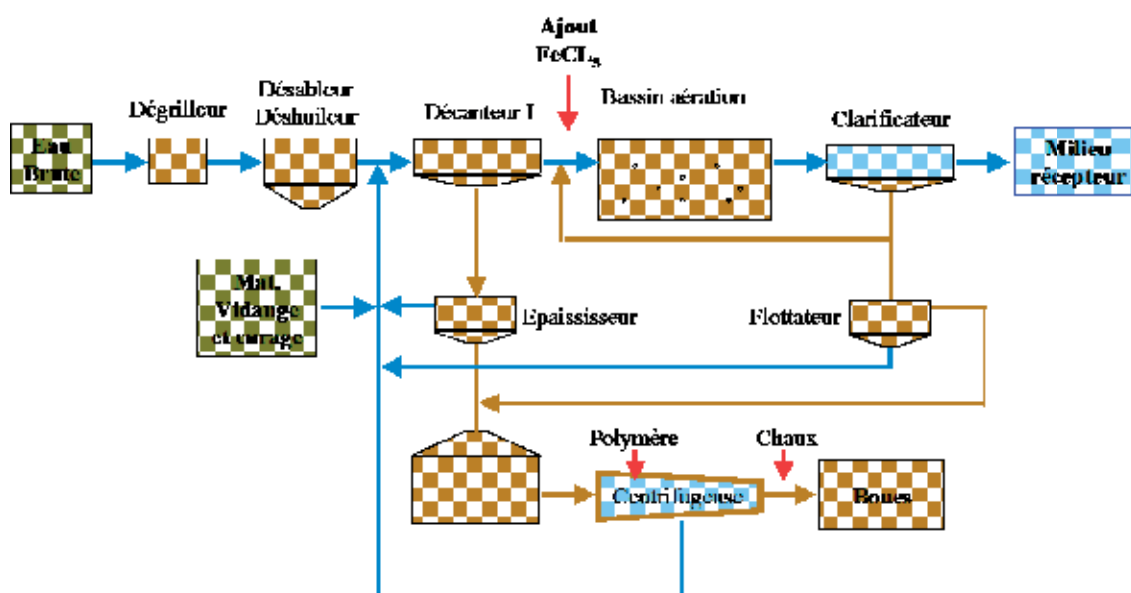


FIGURE 55 SCHEME AND PHOTO OF WWTP 3

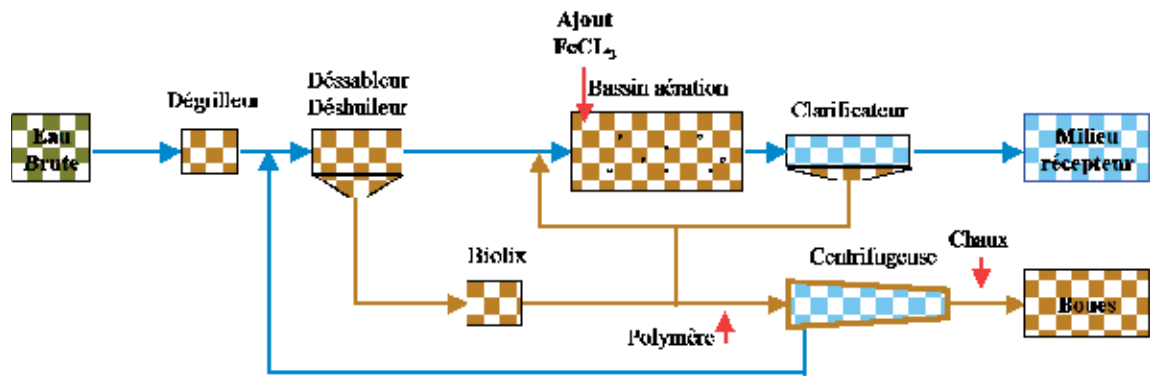
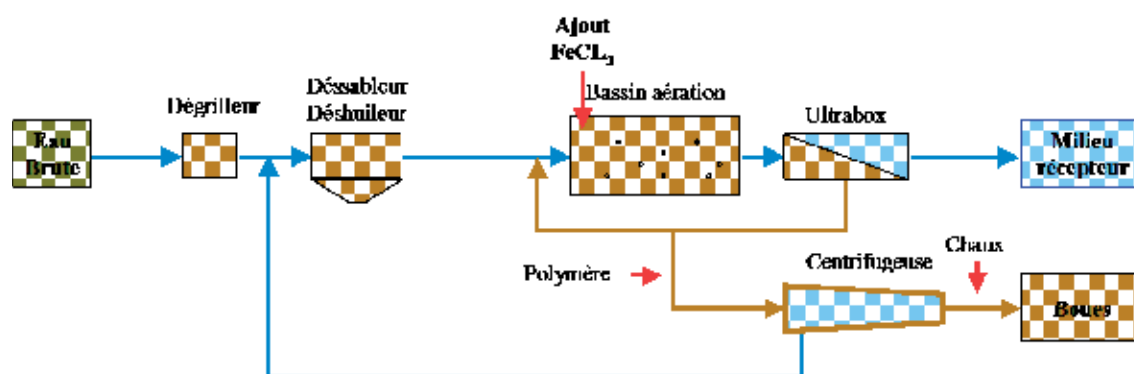


FIGURE 56 SCHEME AND PHOTO OF WWTP 4



ANNEX 8

DEKALB COUNTY'S

