

# ESSE TTT TOXICITY



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# ECOLOGICAL KEY FACTOR TOXICITY

⇒ Part 5

Background document on effect-based trigger  
values for environmental water quality



# COLOPHON

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**Abstract** STOWA developed the conceptual framework of the Ecological Key Factors for the ecological assessment of water quality issues. The key factors describe preconditions for good water quality. The key factors help to structure the available information of water quality and make it possible to pinpoint dominant processes in water system functioning. Understanding of the water quality functioning enables identification of effective restoration measures. Toxicity has been identified as one of the Ecological Key Factors. As part of water quality assessment with EKFs an evaluation should be made to assess whether the water system complies with the key factor toxicity. In a serie of five reports the methodology to assess whether the water quality complies with the ecological key factor toxicity has been described. The methodology gives insight in the effect of chemical compounds on the biology. This report is part 5 and is a background document on effect-based trigger values (EBT's) for the assessment of the ecological risks of (a combination of) compounds. The EBT's are used in the SIMONI model which can be applied to assess the chemical waterquality.

**Keywords** Ecological Key Factors, watersystemanalysis, toxicity, ecological impact assessment, bioassays, effect based trigger values, SIMONI

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## SUMMARY

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STOWA developed the conceptual framework of the Ecological Key Factors for the ecological assessment of water quality issues. The key factors describe preconditions for good water quality. The key factors help to structure the available information of water quality and make it possible to pinpoint dominant processes in water system functioning. Understanding of the water quality functioning enables identification of effective restoration measures. Toxicity has been identified as one of the Ecological Key Factors.

There are key factors for stagnant and streaming waterbodies. The Ecological Key Factor toxicity is applicable for both types.

In a serie of five reports the Ecological Key Factor toxicity is elaborated. The serie contains a main report and 4 attachments.

This report is part 5 and is a background document on effect-based trigger values for the assessment of the ecological risks of (a combination of) chemicals. The concept is part of the toxicology track described in part 1. Part 4 describes the procedures for monitoring.

Aquatic life is subjected to stress due to the release of thousands of chemicals in the water, but it remains hard to determine the ecological actual risks. A paradigm shift in the field of water quality assessment from a single-chemical approach to an effect-based approach would overcome the limitations connected with the current monitoring strategies. Chemical analyses can be conducted only on a limited set of known chemicals and it is virtually impossible to assess mixture toxicity of such complex mixtures. Bioanalytical tools, in combination with chemical analyses, can be a valid alternative to classic monitoring programs. Such an integrated approach would provide information regarding the overall impact of co-exposure to multiple chemicals (known and unknown) on different levels of biological organization.

This study aimed to design environmental effect-based trigger values (EBT) for a selection of bioassays. The aim of these EBTs is that they should provide initial hazard identification of organic micropollutants for the aquatic organisms. The EBTs will be included in a model called "SIMONI" (Smart Integrated MONItoring) that can be used to discriminate between 'low risk' sites, where no further analyses are needed, and 'potential risk' sites, where an additional risk assessment has to be conducted. The goal is to reduce monitoring costs, and meanwhile generate a more complete analysis of the chemical water quality by using a battery of bioassays covering the most relevant modes of toxicant action.

A three-step approach was used to design effect-based trigger values (EBT). A selection of compounds was made that have a known response in the bioassay, with relative effect potencies (REPs) close to the reference compound for that assay. The first step was a literature search for toxicity data on these selected compounds, and conversion to their TEQ values (toxic equivalents of the respective reference compounds). Lowest TEQs of all toxic effects found (divided by an assessment factor) will be used as 'save TEQ'. The second step was a species sensitivity distribution of all TEQ values in order to estimate the TEQ level that may cause an adverse effect to 5% of the species (HC5 TEQ). The HC5 TEQ should preferably be higher than or equal to the proposed low-risk effect-based trigger value (EBT). The final step was a benchmark with Waternet field data. The average bioassay responses at eight ecologically clean sites were considered as 'clean TEQ'. A realistic EBT was derived that should be higher than the average effect observed at eight reference sites with a good ecological status. In order to get a good discrimination between sites, the EBT should be exceeded only at a limited number of seriously polluted sites. Therefore, further validation studies will be needed to optimize the proposed trigger values in the near future.

Effect-based trigger values were derived for:

- estrogenic activity (Era CALUX): 0.5 ng EEQ/L;
- anti-androgenic activity (anti-AR CALUX): 25 µg F1EQ/L;
- glucocorticoid activity (GR CALUX): 100 ng DEQ/L;
- dioxin-like activity (DR CALUX): 50 pg TEQ/L;
- PPAR $\gamma$  receptor activity (PPAR $\gamma$  CALUX): 10 ng REQ/L;
- toxic PAHs activity (PAH CALUX): 150 ng BEQ/L;
- oxidative stress (Nrf2 CALUX): 10 µg CEQ/L;
- pregnane X receptor activity (PRX CALUX): 3 µg N1EQ/L;
- five classes of antibiotics activity (RIKILT WaterSCAN):
  - aminoglycosides activity: 500 ng N2EQ/L;
  - macrolides &  $\beta$  lactams activity: 50 ng PEQ/L;
  - sulphonamides activity: 100 ng SEQ/L;
  - tetracyclines activity: 250 ng OEQ/L;
  - quinolones activity: 100 ng F2EQ/L.

It is essential to propose better monitoring strategies in order to preserve the quality of freshwater environment. The introduction of bioassays in monitoring programs can lead to a more sustainable water quality assessment. This study may provide a reference for further measures and implementation of risk assessment of micropollutants in the water cycle.

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# 1 INTRODUCTION

## 1.1. ECOLOGICAL KEY FACTORS

STOWA designed the conceptual framework of the Ecological Key Factors for the ecological assessment of water quality issues. The key factors describe preconditions for good water quality. The key factors help to structure the available information of water quality and make it possible to pinpoint dominant processes in water system functioning. Understanding of the water quality functioning enables identification of effective restoration measures. Toxicity has been identified as one of the Ecological Key Factors.

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## 1.2. EFFECTS OF CHEMICALS

Since the beginning of the industrialized era a constantly increasing number of anthropogenic chemicals have been released in the environment. The thousands of substances that reach the water cycle such as pharmaceuticals, pesticides, reproductive hormones, steroids, detergents, disinfectants, insect repellants and fire retardants (Erickson, 2002), may cause deterioration of the water quality and pose a threat to the aquatic ecosystems. In 2000, the European Parliament approved the Water Framework Directive (WFD, 2000/60/EC), in order to develop good monitoring programs and to classify the water bodies based on their chemical and ecological quality. The WFD represents a milestone in the field of water monitoring and management, but it is far from conclusive (Dworak et al., 2005). According to the directive, chemical quality of the water should be investigated trough chemical analysis of selected priority compounds, whose concentrations should not exceed established Environmental Quality Standards or EQSs (Environmental Quality Standards Directive, 2008/105/EC). However, this single-chemical approach has been proven insufficient and can lead to an underestimation of the potential toxic hazard caused by thousands of compounds present in the water, since “as any analytical chemist knows, what you see depends on what you look for” (Lynn Roberts, Johns Hopkins University). Additionally, new chemicals are continuously developed to replace the current ones, and, as a consequence, the list of priority substances should be constantly updated. Moreover, chemicals that are present under detection limits can still cause significant mixture toxicity or create toxic metabolites and secondary products. Several authors (Altenburger et al., 2015; Escher and Leusch, 2012) highlight the need to development new monitoring methods that will allow a more realistic overview of the risk connected with the presence of chemicals in the water phase.

A paradigm shift in the field of water quality assessment from a single-chemical approach to an effect-based approach would overcome the present limitations (Poulsen et al., 2011; Maas et al., 2004). As previously stated, only a small fraction of the compounds that could be present in the water are monitored with current programs. The introduction of effect-based techniques will lead to a more holistic and qualitative environmental risk assessment. As a matter of fact, this shift will allow us to detect toxicity of mixtures of (un)known bioavailable chemicals and to better link the cause (presence of chemicals) to the effect (toxicity for the aquatic communities). The first step of a risk assessment is a hazard identification to establish if a certain situation is of concern or not. The combination of bioassays with chemical analysis in water assessment appears to be a more suitable tool than only chemical analysis, which was demonstrated in several studies investigating the reliability of an effect-based methodology. Among others, Chapman et al. (2011) investigated water



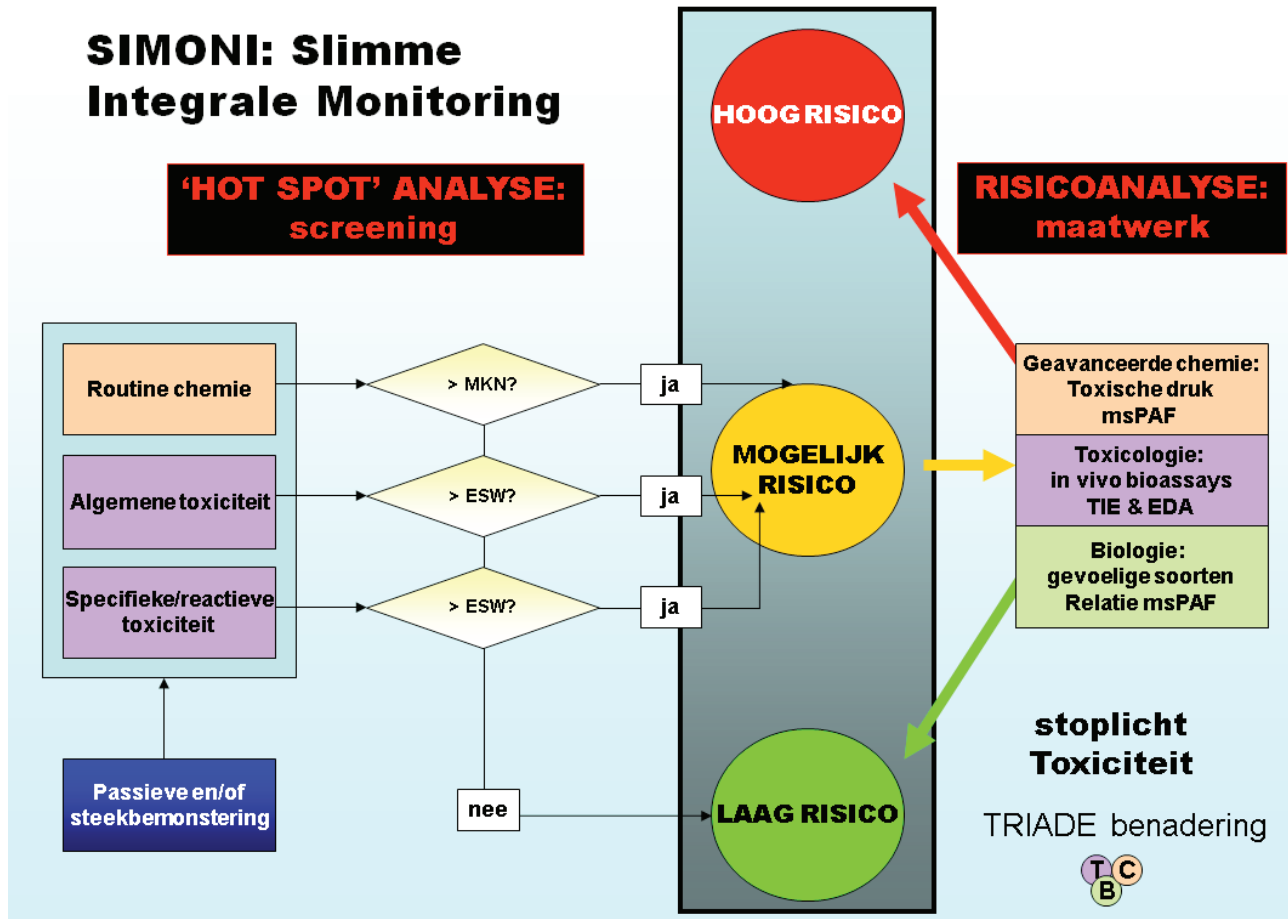
samples from nine water reclamation plants in Australia with a combination of chemical analysis and bioassays in order to characterize human and environmental risks. One of the main conclusion of this investigation was that bioassays and chemical analysis were complementary and in agreement. Additionally, bioassays were able to provide a more complete analysis of the water quality, since they detected activity at concentrations below the detection limit of chemical analyses. Maas and co-workers (2004) showed that bioassays are sensitive enough to evaluate the effects of all WFD priority pollutants, in particular PAHs, herbicides and insecticides.

Effect-based tools or bioassays are techniques that investigate the toxicity of samples, using biological systems. They can be classified in two categories based on the level of organization of the biological system: *in vivo* bioassays (conducted on organisms, population, or ecosystem level) and *in vitro* bioassays (conducted on isolated cells or tissues). The biggest limitation of the effect-based techniques is their inability to determine the identity of the compounds causing the observed toxic effects. Certain *in vitro* bioassays, however, allow the identification of groups of chemicals with a similar mode of toxicant action (MOA). Generally, *in vivo* bioassays assess non-specific toxicity while *in vitro* bioassays will measure specific or reactive toxicity. Several types of *in vitro* bioassays are available to detect a variety of toxic activities, such as estrogenic, androgenic, dioxin-like effects, genotoxicity and neurotoxicity (Escher and Leusch, 2012). A battery test is defined as a selection of bioassays targeting different toxic endpoints. The endpoint selection is based on the protection goal or the chemical activity that need to be targeted. A battery test selected for water quality assessment should cover all relevant MOAs, since the goal is to get a wide comprehensive picture of the toxic potency of the available micropollutants. Since *in vitro* bioassays are able to detect specific toxic activity of groups of chemicals, a broad selection of toxic endpoints will gain a more complete picture of chemical risks, including the effects of unexpected or unknown toxic substances (Escher and Leusch, 2012).

Several countries are already using bioassays to monitor the microchemical water quality (see de Zwart, 1995 and Wernersson et al., 2014 for an overview). Battery tests have also been used for water quality assessment, e.g., to investigate the efficiency of sewage treatment plants and the chemical quality of drinking water (Kienle et al., 2011; Macova et al., 2010; Pablos et al., 2009; Macova et al., 2011; Zęgura et al., 2009). In the Netherlands, bioassays have been applied for many years in surveillance monitoring of the Meuse, Scheldt and Rhine river basin. The use of bioassays, however, is restricted to research programs since there are no official guidelines to incorporate effect-based tools in regulatory monitoring programs. One of the main reasons for this is the lack of tools for a clear interpretation of bioassay results, such as effect-based triggers values (ETVs). The definition of EBTs will help water managers to define the bioassay responses that indicate that levels of micropollutants can be considered a “low risk” or a “potential risk” for the ecosystem. Several international projects like DEMAU (<http://demeau-fp7.eu>) and SOLUTIONS (<http://www.solutions-project.eu>), investigate and promote implementation of effect-based tools in current regulations for water quality assessment. In 2010, Waternet started a project called “Smart monitoring”. The main goal of this project is to combine traditional chemical analysis with effect-based tools in a tiered screening approach, in order to obtain a more efficient and cost-effective environmental risk assessment (Van der Oost et al., in prep). Bioassays will represent a powerful screening tool that will allow the classification of sites by a model called “SIMONI” (Smart Integrated Monitoring toxicity traffic light, Figure 1). According to the model, sites will be classified as low risk (green), potential risk (orange) and high risk (red), based on the responses of a battery of selected bioassays and target chemical analyses. Only when a potential risk is indicated an investigation with chemical analysis will be needed, in order to identify the chemicals causing the observed toxic effects.

**FIGURE 1**

Schematic representation of the SIMONI model (Van der Oost, in preparation a). Sites will be classified as low risk (green) or potential risk (orange) based on the responses of a bioassay battery. Advanced chemical analyses will only be needed at potential risk, to identify if the chemicals causing the toxic effects are a high ecological risk (red).



### 1.3. IN VITRO BIOASSAY BATTERY

The present study focusses on EBT for *in vitro* bioassays for specific and reactive effects, and bioassays for the determination of antibiotics activities. As stated above, *in vitro* bioassays are able to detect specific activities caused by unknown mixtures of compounds with the same MOAs (Sonneveld et al., 2005). The specific activity is expressed in toxic equivalents (TEQ), i.e., the amount of a reference compound (see Table 1) that would cause the same effect as all compounds of the unknown mixture the bioassay is exposed to. CALUX (Chemical Activated LUciferase gene eXpression) techniques are *in vitro* bioassays performed with modified cell lines that contain luciferase reporter genes and specific receptor sites. The binding of chemicals to a specific receptor will induce luciferase gene expression. After addition of the luciferine substrate, the intensity of the luciferase induction can be measured as an increased luminescence, which is a measure of the activity of all micropollutants with this specific MOA. Detection of antibiotics activities was performed with the Water SCAN bioassay, developed by RIKILT (Netherlands). This bioassay uses agar plates inoculated with five species of microorganisms that are sensitive to different classes of antibiotics with the same MOAs. After exposure to a sample, the growth inhibition of the microorganisms is measured as a clear area in the agar. The surface area of the clear spot, which is proportional to the total activity of specific antibiotics in the sample, was used to quantify the response. Different versions and improvements of the bioassay have been suggested and detailed information can be found in Pikkemaat et al. (2008).

#### ER CALUX

Estrogen receptors (ER) are a group of proteins found inside cells that are activated by the hormone estrogen (17 $\beta$ -estradiol). Once activated by estrogen, the ER is able to translocate into the nucleus and bind to DNA to regulate the activity of different genes (i.e. it is a DNA-binding transcription factor). Estrogen is the primary female sex hormone that is responsible for the development and regulation of the female reproductive system and secondary sex characteristics. Estrogen may also refer to any substance, natural or synthetic that mimics the effects of the natural hormone.

#### Anti-AR CALUX

The androgen receptor (AR) is a nuclear receptor that is activated by binding the androgenic hormones, testosterone or dihydrotestosterone in the cytoplasm and then translocating into the nucleus. The androgen receptor is most closely related to the progesterone receptor, and progestins in higher dosages can block the androgen receptor. The main function of the androgen receptor is as a DNA-binding transcription factor that regulates gene expression. Androgen regulated genes are critical for the development and maintenance of the male sexual phenotype.

#### DR CALUX

Dioxin Responsive bioassays, such as DR CALUX, are used to detect dioxins and dioxin-like compounds. It is based on the mechanisms of the arylhydrocarbon receptor (AhR) pathway (Murk et al., 1996). Chronic activation of the AhR-pathway by these compounds has been shown to cause cancer in the predominantly the liver and can cause developmental defects in vertebrates. DR CALUX is also used to screen food for dioxins and dioxin-like compounds in order to guarantee food safety. Dioxins are considered to be the most toxic man-made chemicals.

#### GR CALUX

The glucocorticoid receptor (GR) is the receptor to which cortisol and other glucocorticoids bind. The primary mechanism of GR action is the regulation of gene transcription. After the receptor is bound to glucocorticoids, the receptor-glucocorticoid complex can take either of two paths (Rhen and Cidlowski, 2005). The activated GR complex up-regulates the expression of anti-inflammatory proteins in the nucleus or represses the expression of pro-inflammatory proteins in the cytosol (preventing the translocation of other transcription factors from the cytosol into the nucleus).

#### NRF2 CALUX

Oxidative stress reflects the imbalance between the manifestation of reactive oxygen species and the biological ability to detoxify the reactive intermediates or to repair the resulting damage (Sies, 1985). Disturbances in the normal redox state

of cells can cause toxic effects through the production of peroxides and free radicals that damage all components of the cell, including proteins, lipids, and DNA. Base damage is mostly indirect and caused by reactive oxygen species (ROS) generated, e.g. superoxide radical, hydroxyl radical and hydrogen peroxide. The detection of oxidative stress with the *in vitro* Nrf2 CALUX ('Nuclear-factor-E2-related factor') is based on the activation of the Nrf2 pathway, which regulates cytoprotective enzymes in response to oxidants and electrophilic compounds through binding to the antioxidant response element (ARE) (Nguyen et al., 2009).

### PAH CALUX

Polycyclic aromatic hydrocarbons (PAHs) are one of the most widespread organic pollutants. The toxicity of PAHs is structure-dependent. Isomers (PAHs with the same formula and number of rings) can vary from being nontoxic to extremely toxic. One PAH compound, benzo[a]pyrene (BaP), is notable for being the first chemical carcinogen to be discovered. Certain PAHs are well known for their carcinogenic, mutagenic, and teratogenic properties. The PAH CALUX is designed for the detection of the activity of the most toxic PAHs, like BaP.

### PPAR $\gamma$ CALUX

The peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ) regulates fatty acid storage and glucose metabolism. The genes activated by PPAR $\gamma$  stimulate lipid uptake and adipogenesis by fat cells. PPAR $\gamma$  knockout mice fail to generate adipose tissue when fed a high-fat diet (Jones et al., 2005). Many naturally occurring agents directly bind with and activate PPAR $\gamma$ .

### PXR CALUX

PXR is a nuclear receptor whose primary function is to sense the presence of foreign toxic substances and in response up regulate the expression of proteins involved in the detoxification and clearance of these substances from the body (Kliewer et al., 2002). Receptors such as PXR recognize such xenobiotics and to control the expression of a large series of phase I, phase II, and phase III metabolizing enzymes and transporters. The encoded protein is a transcriptional regulator of the cytochrome P450 gene CYP3A4, binding to the response element of the CYP3A4 promoter as a heterodimer with the retinoic acid receptor (RAR). PXR is activated by a range of compounds that induce CYP3A4, including dexamethasone and rifampicin (Bertilsson et al., 1998).

## ANTIBIOTICS ACTIVITIES

### *Aminoglycosides*

Aminoglycosides bind irreversibly to different subunits of bacterial ribosomes and therefore interfere with protein synthesis (Brain et al., 2004). Streptomycin is one of the most commonly used aminoglycosides and, apart from binding to the 30S ribosomal subunit of bacteria, slows down the already initiated protein synthesis and produces misreading of mRNA (Van der Grinten et al., 2010).

### *Macrolides & $\beta$ -lactams*

Within the macrolides and  $\beta$ -lactams antibiotics, an important number of compounds have been tested on different organisms. Macrolides bind to the bacterial 50S subunit of the ribosome, inhibiting translocation of tRNA during translation (Van der Grinten et al., 2010).  $\beta$ -Lactams are another group of compounds, which often appear classified together with macrolides. They inhibit the synthesis of cell wall in bacteria by targeting the transpeptidase enzymes of these organisms (Wilke et al., 2005). Additionally, other compounds such as carbacephems (for example, cephalexin), pose a similar mode of action as  $\beta$ -lactams, inhibiting the synthesis of bacterial cell walls (Brain et al., 2004).

### *Sulfonamides*

Sulfonamides are synthetic compounds which act as folate antagonists, avoiding the production of coenzyme dihydrofolic acid by blocking the conversion of paminobenzoic acid in microorganisms (Brain et al., 2004). They pose a broad

spectrum of action against bacteria and coccidian and their presence in the environment is mainly due to the excretion of active forms and the transformation of their inactive metabolites back into the active form by bacteria, which explains their persistency and resistance in the environment (De Liguoro et al., 2009). Diaminopyrimidines are a group of antibiotic commonly presented together with sulfonamides because of their synergistic effect. The most common diaminopyrimidine is trimethoprim, which inhibits the enzyme dihydrofolate reductase by reversible binding, therefore interfering with the synthesis of folate (Van der Grinten et al., 2010).

### Tetracyclines

Tetracyclines bind irreversibly to the 30S ribosomal subunit, inhibiting the protein synthesis by blocking the binding of aminoacyl transfer to DNA (Brain et al., 2004). This group of antibiotic is most commonly used in veterinary applications. Oxytetracycline, one of the most commonly used tetracyclines, inhibits the protein synthesis by avoiding the interaction between aminoacyl-tRNA and the bacterial ribosome (Kołodziejska et al., 2013; Van der Grinten et al., 2010). Florphenicol and derivatives from thiamphenicol act on protein synthesis of Gram-negative and Gram-positive bacteria by inhibiting transpeptidation (Christensen et al., 2006; Kołodziejska et al., 2013).

### Quinolones

Quinolones are DNA gyrase and topoisomerase IV inhibitors, which present higher affinities for the bacterial enzyme than for the vertebrates' enzyme (Brain et al., 2004; Carlsson et al., 2009). These antibiotics have evolved from the first generation of quinolones, such as oxolinic acid, to the second and third generation, the fluoroquinolones. In these last ones, a fluorine atom was added to the structure, which improved the efficiency of these compounds (Robinson et al., 2005). One of the most commonly used quinolones is enrofloxacin, from which 11% is estimated to be transformed into its metabolite, ciprofloxacin (Rico et al., 2014a). Therefore, these compounds are sometimes estimated or tested for toxicity together.

## 1.4 AIM OF THE STUDY

As part of the 'Smart monitoring' (Waternet) and 'Ecological Key Factor Toxicity' (STOWA) projects, the present study aims to derive "low risk" effect-based trigger values (EBTs) for eight bioassays, targeting estrogenic activity, anti-androgenic activity, glucocorticoid activity, PAH- and dioxin-like activity, lipid metabolism (PPAR $\gamma$ ), oxidative stress, pregnane X receptor (PXR) activity and five antibiotics activities. The selected endpoints and the corresponding bioassays are presented in

**TABLE 1**

*Target activities and in vitro bioassays selected for environmental hazard identification, with reference compounds for each bioassay.*

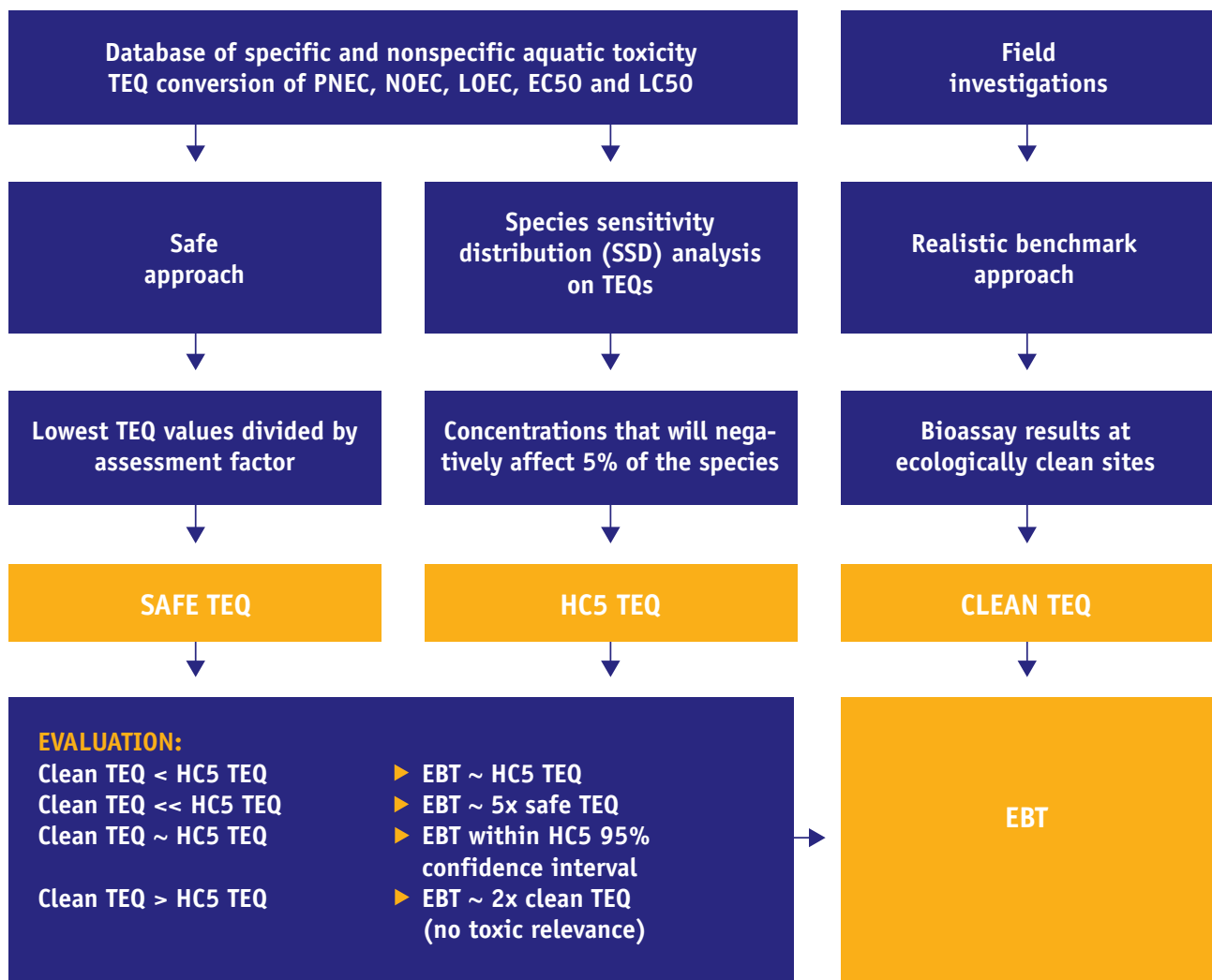
Activity detected	Bioassay	Reference Compound	(CAS)
Estrogenic	ERa CALUX	17- $\beta$ estradiol	50-28-2
Anti-androgenic	antiAR CALUX	Flutamide	13311-84-7
Dioxin and dioxin-like	DR CALUX	2,3,7,8-TCDD	1746-01-6
Glucocorticoid	GR CALUX	Dexamethasone	50-02-2
PPAR $\gamma$ receptor	PPAR $\gamma$ CALUX	Rosiglitazone	122320-73-4
Toxic PAHs	PAH CALUX	Benzo[a]pyrene	50-32-8
Oxidative stress	Nrf2 CALUX	Curcumin	458-37-7
Pregnane X receptor	PXR CALUX	Nicardipine	54527-84-3
Antibiotics activities	Aminoglycosides	Neomycin	1404-04-2
	Macrolides & $\beta$ -Lactam	Penicillin	61-33-6
	Sulphonamides	Sulfamethoxazole	723-46-6
RIKILT WaterSCAN	Tetracyclines	Oxytetracycline	79-57-2
	Quinolones	Flumequine	42835-25-6

The EBTs developed in the present study will be incorporated in a model called “SIMONI”, which will be used to investigate the micro-chemical quality of the water (Figure 1). Within this model, the hazard identification (left site of the schedule in Figure 1) will be based on the responses of a battery of selected bioassays. The comparison between the bioassay responses and the EBTs will allow the classification of the investigated sites in low risk or potential risk for adverse ecological effects. Additional investigations will be conducted only on sites for which the bioassay responses indicate a potential ecological risk, and will be directed towards the identification of the toxic compounds in the samples through chemical analysis (van der Oost, in preparation a).

## 2 METHODS

The derivation of EBTs for the selected CALUX bioassays and antibiotic activities will be based on a three-step approach. The first step is a selection of compounds that are known to trigger a response in the bioassay, with relative effect potencies (REPs) close to the reference compound, a literature search for toxicity data on these selected compounds, and conversion to their TEQ values of the respective reference compounds. Lowest TEQs of the toxic effects found (divided by an assessment factor) will be used as safe toxic equivalents concentration (safe TEQ). The second step was a species sensitivity distribution of all TEQ values in order to estimate the TEQ level that may cause an adverse effect to 5% of the species (HC5 TEQ). The HC5 TEQ should preferably be higher than or equal to the proposed low-risk effect-based trigger value (EBT). The last step was a benchmark with Waternet field data. A realistic EBT should be higher than the average effect observed at eight reference sites with a good ecological status (clean TEQ). In order to get a good discrimination between sites, the EBT should be exceeded only at a limited number of polluted sites. The steps that were followed for the definition of EBTs are summarized in Figure 2, and described in detail in the following paragraphs.

**FIGURE 2**  
Schematic representation of the steps taken for the design of environmental EBTs.



## 2.1 TOXICOLOGICAL DATABASE

A collection of available toxicological data is the first and necessary step on which every method that aims to set quality standards is based. In a classical single-chemical approach, data are collected for the compound under study. However, due to the nature of the bioassays, it was necessary to follow a different approach. Since bioassays are effect-based tools that measure activities caused by a mixture of available compounds in a sample, the nature of the compounds that cause the observed effect remains unknown. The measured activity is expressed as *toxic equivalent* (TEQ) concentration to a reference compound, i.e. the equivalent concentration of a reference compound that would cause the same observed effect as the (un)known mixture of compounds present in the investigated sample. The reference compound is a chosen representative of a certain activity and is specific for each bioassay (see Table 1 for reference compounds of all selected bioassays). Additionally, different substances can be more or less potent in triggering a response than the corresponding reference compound. The concept of *relative effect potencies* (REPs) is used to account for these differences. REPs can be calculated by dividing the effect concentration of the reference compound by the concentration of another compound that is required to produce a similar effect. The TEQ concentration of a compound can thus be calculated by multiplying the actual concentration by its REP value.

Since a search of toxicological data for only the reference compounds is unreliable to set relevant EBTs, we included a selection of other compounds that are able to trigger a response in each bioassay. The compounds were selected based on their REPs. A complete list of the selected compounds and their REPs is presented in Appendix I. For all toxicological endpoints a search in scientific literature and toxicological databases was conducted to establish toxicity data of all selected compounds in water organisms at different trophic levels. Toxicity data were classified in five groups, i.e. PNEC (Predicted No-effect Concentration), NOEC (No Observed Effect Concentration), LOEC (Lowest Observed Effect Concentration), EC50 (50% Effect concentrations) and LC50 (concentrations lethal to 50% of the test organisms). The complete dataset of all toxicity data that were used for this study is presented in Appendix II. All toxicity data were converted to TEQ concentrations of the respective reference compounds by multiplication with the REPs. According to the precautionary principle, chronic toxicity was considered the most relevant for environmental risk assessment. In order to compare chronic with acute data some data conversion was needed.

**Assumption 1:** the focus of the trigger value design will be on chronic toxicity; in order to compare all toxicity data, acute data were converted in chronic data by dividing them by an assumed acute-to-chronic ratio of 10 (Durand et al., 2009). Since there are no strict definitions for acute and chronic exposure times, an assumption of the criteria for chronic exposure for different taxa had to be estimated.

**Assumption 2:** the estimated durations of chronic experiments for different groups of organisms are listed in Table 2; toxicity data of experiments with shorter exposure times are divided by a safety factor of 10 (i.e., assumption 1: acute-to-chronic concentration ratio).

**Assumption 3:** since chemicals with very low relative effect potencies (REPs) will give extremely low TEQ values, a certain restriction was needed for a realistic hazard identification; in order to compare the REP impact, all calculations for each EBT were performed on two chemical selections: the REP1 group included compounds with REPs > 0.1, while the REP2 group included compounds with REPs > 0.001.

The REPs for the CALUX bioassays were provided by BDS (BioDetection Systems, Amsterdam, The Netherlands), calculated from EC10 results of the different compounds. The REPs for the detection of antibiotic activity were estimated from the detection limits of the selected compounds. The RIKILT WaterScan and similar methods, such as RIKILT MeatScan or NDKT (New Dutch kidney test), are based on the growth inhibition of certain microorganisms after exposure to the samples. It was assumed that the detection limits of the antibiotics, i.e. the minimum concentrations that cause a detectable growth inhibition, corresponded to their potencies. Therefore, the REPs were estimated by dividing the detection limit of the ref-



erence compound by the detection limit of the considered compound. REPs were calculated from the detection limits of the RIKILT WaterScan method or those reported by Pikkemaat et al. (2008) for RIKILT MeatScan or NDKT. All REPs for each bioassay that were used for the present study are presented in APPENDIX I.

**TABLE 2**

*Criteria applied in the present study to estimate chronic exposure.*

Organism	Chronic exposure (days)
Protozoa	≥ 1
Bacteria	
Fungus	
Polyp	≥ 4
Algae	
Rotifer	
Crustacean	
Insect	
Mollusca	
Worm	
Plant	≥ 7
Amphibian	
Fish	

## 2.2 SAFE APPROACH (SAFE TEQ)

According to the precautionary principle it is relevant to define “safe” TEQs that indicate no risk levels of active compounds to the ecosystem. The lowest TEQs concentrations for each toxicological endpoint (PNEC, NOEC, LOEC, EC50 and LC50) were selected and divided by an assessment or safety factor (AF), which ranged for 1 to 100 according to the toxic endpoint considered (see Assumption 4: Table 3).

**Assumption 4:** assessment factors to estimate save biological activities by extrapolation of five different toxic endpoints are listed in Table 3.

**TABLE 3**

*Assessment factors (AFs) applied in the present study to convert toxicity data to assumed save levels.*

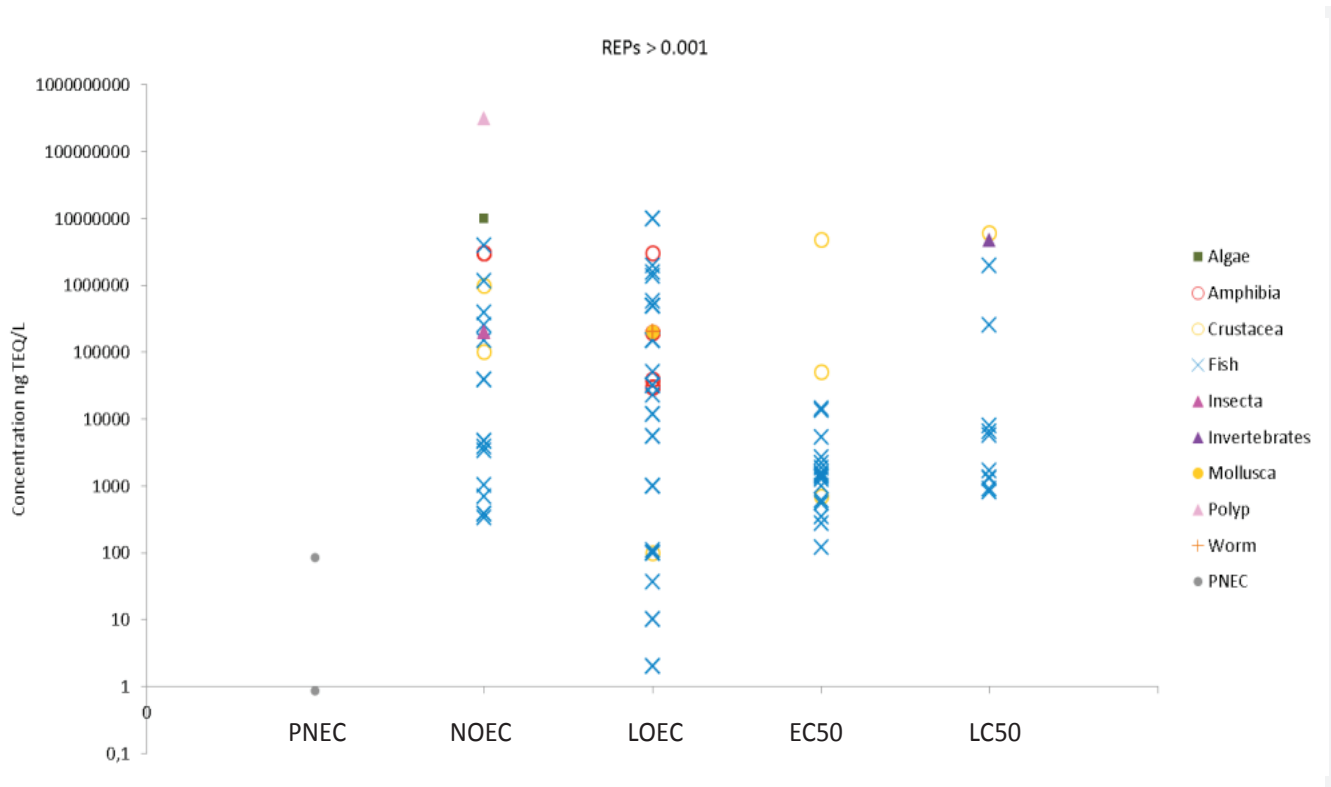
Endpoint	AF
PNEC	1
NOEC	1
LOEC	5
EC50	10
LC50	100

The lowest of all chronic toxicity values found in the literature, divided by their respective AFs, was considered as a safe value for water organisms and defined as the “safe” toxic equivalents concentration (safe TEQ). As an illustration of the ‘save approach’, all collected toxicity data for dioxin-like compounds of the REP2 group are presented in Figure 3. The lowest LOEC (divided by assessment factor 5) was used as the safe TEQ. Since these “no risk” safe TEQs will be exceeded at most moderately polluted sites, a more realistic approach was followed in order to define a “low risk” effect-based trigger value (EBT). This approach will be described in the next paragraphs.

Graphic representations of all collected toxicity data that were used for the EBT design for all bioassays (both REP1 and REP2 groups) are presented in Appendix IV.

**FIGURE 3**

*Toxicity dataset for dioxin-like compounds with REPs > 0.001.*



### 2.3 SPECIES SENSITIVITY DISTRIBUTION ANALYSIS (HC5 TEQ)

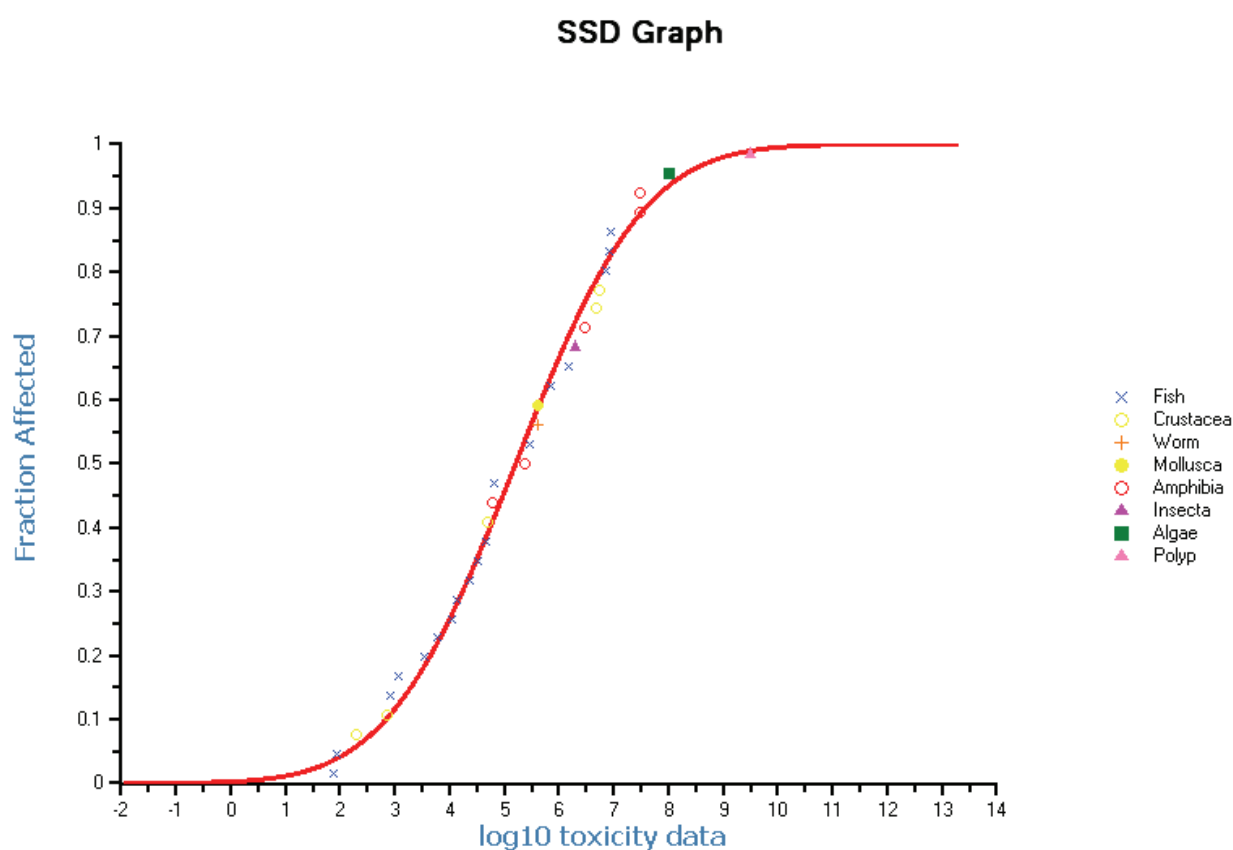
A more realistic trigger value approach ('low risk' instead of 'no risk') was based upon a Species Sensitivity Distribution (SSD) analysis (Posthuma et al., 2002). SSD curves are usually generated by fitting the distribution of log-transformed toxicological data (usually NOEC, EC50 or LC50) of several species for a single compound. When more data are available for the same compound in the same species, an average toxic concentration is used for the SSD. The output of the SSD distribution can be used to determine the 5th percentile hazard concentration (HC5 TEQ), which represents the concentration of the investigated compound that will negatively affect 5% of the species.

In the present study an unusual SSD approach was applied, since toxicological data of various compounds that trigger a response in the same bioassay had to be included. Since it is impossible to generate SSD curves with data of different substances, we converted all toxicological data to TEQ concentrations of the reference compounds of the bioassay. This conversion allowed us to generate SSD curves with the collected toxicity data for all species and for each original compound. Average TEQ values were used if different toxicity values were available for the same compound in the same species. The SSD curves were preferably generated from EC50 TEQ values with the statistical software ETX 2.0 (Vlaardingen et al., 2004).

For the DR CALUX and GR CALUX, the amount of available EC50 values was insufficient to run the SSD analysis. As a consequence, a combination of NOEC, LOEC and EC50 TEQ values were used. Prior to the analysis, NOEC and LOEC values were multiplied by 10 and 2 respectively, in order to get an AFs of 10 like the EC50 (assumption 4, Table 3), to account for the differences between these toxicological endpoints. The HC5 TEQ-values that were determined by this approach were meant to be used as upper limits of the low-risk effect-based trigger values. In some cases, however, an EBT above the HC5 TEQ had to be defined, due to higher benchmark data at sites with a good ecological status.

As an illustration of the SSD approach, the SSD curve with collected toxicity data (pg TEQ/L) for dioxin-like compounds of the REP2 group are presented in Figure 4. The TEQ-level that is hazardous to 5% of the organisms can be estimated with the SSD curve (affected fraction 0.05). SSD curves of all collected toxicity data that were used for the EBT design for all bioassays (both REP1 and REP2 groups) are presented in Appendix V.

**FIGURE 4**  
*Species sensitivity distribution (SSD) for dioxin-like compounds with REPs > 0.001.*



#### 2.4 BENCHMARK APPROACH BIOASSAYS (CLEAN TEQ)

Another approach to obtain more realistic “low risk” EBTs was a benchmark with field data. This benchmark approach was primarily carried out with the results of bioassay monitoring at eight reference sites with a good ecological status. The rationale behind this approach was that the bioassay responses that were observed at sites with a good ecological status were not considered crucial for realistic overall risk estimations and should not indicate potential ecological hazards. Therefore the benchmark data were used to indicate the lower limits for the EBTs, the “clean TEQ”.

## 2.5 DERIVATION OF EBT

In the ideal case, the clean TEQ would be somewhere in between the safe TEQ and the HC5-TEQ, determined with SSD. Further refinement of EBT derivation was based upon evaluation of the safe, HC5 and clean TEQs according to following algorithms. If the clean TEQ was lower than the HC5 TEQ of the bioassay, than an EBT around the HC5 TEQ value was proposed. If the clean TEQ was much lower than the HC5 TEQ of the bioassay, than an EBT of approximately 5 times the safe TEQ was proposed. This factor ranges between 2 and 10, depending upon the strength of the dataset (data for many substances and many species) used to determine safe and HC5 TEQs. If the clean TEQ was close to the HC5 TEQ, than a value within the 95% confidence interval of HC5 TEQ was proposed as EBT. If the clean TEQ was much higher than the HC5 TEQ, than an EBT of approximately 2 times the clean TEQ was proposed, depending upon the strength of the dataset. This latter situation was typical for bioassays that are responsive to a wide array of chemicals, such as anti-AR, oxidative stress and PXR responses. EBT derived for these assays are not considered toxicologically relevant, but are used as indicators for overall chemical stress.

The bioassay analyses for the benchmark approach were performed according to validated standard protocols, as described by Van der Oost et al. (in preparation b). The modes of action (MOA) and toxicological relevance of the various bioassays are described in the introduction.

### 3 RESULTS

The complete dataset of the collected toxicity data is presented in Appendix II. The lowest toxic concentrations found for five endpoints (PNEC, NOEC, LOEC, EC50 and LC50) are summarized for each bioassay in Appendix III.

#### 3.1 ESTROGENIC ACTIVITY

Release of endocrine disrupting compounds (EDC) in the water received much attention in the last decades, due to their ability to negatively affect aquatic populations. It was possible to find numerous studies in literature investigating the toxicity of estrogenic compounds, including biomarker endpoints (e.g. production of vitellogenin and changes in gene expression). The collected toxicity data for substances with estrogenic activity are presented in Appendix II A. The lowest toxic concentrations found for the five endpoints are presented in Appendix III.

##### Safe approach

Graphic representations of the collected toxicity values for the REP1 and REP2 groups are presented in Appendix IV as estradiol equivalents per liter water (EEQ). In the REP1 group, the lowest value found was a PNEC of 0.035 ng/L for 17 $\alpha$ -Ethinyl estradiol (James et al., 2014). However, 17 $\alpha$ -Ethinyl estradiol presents a relative potency of 1.56, and the PNEC, once transformed in EEQ, was equal to 0.055 ng EEQ/L. The lowest LOEC found was 0.5 ng/L of 17 $\alpha$ -Ethinyl estradiol for zebrafish (*Danio rerio*) after acute exposure (Colman et al., 2009). This value was multiplied by the acute to chronic ratio of 0.1 and transformed in EEQ. This resulted in a final value of 0.078 ng EEQ/L. After application of a safety factor of 5, a safe TEQ of 0.016 ng EEQ/L was proposed. In the REP2 group the safe TEQ was based on the lowest LOEC of 3.3 ng/L for rainbow trout (*Oncorhynchus mykiss*) after chronic exposure to estrone (Thorpe et al., 2003) with a REP of 0.01. This resulted in a TEQ value of 0.033 ng EEQ/L that was divided by a safety factor of 5, which lead to a proposed safe TEQ value of 0.007 ng EEQ/L.

##### SSD

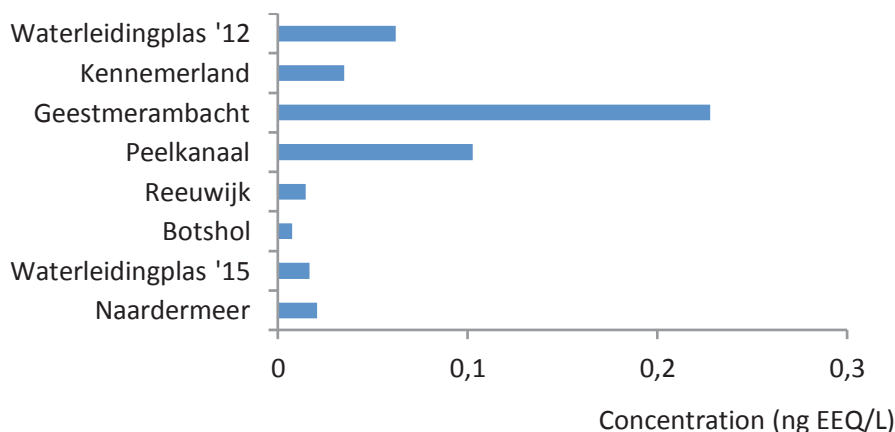
The SSD curves for the REP1 and REP2 groups are presented in Appendix V. The SSD analysis for the REP1 group resulted in a HC5 TEQ equal to 0.47 ng EEQ/L (95% confidence interval from 0.009 to 6.2 ng EEQ/L). The SSD analysis for the REP2 group resulted in a HC5 TEQ equal to 0.52 ng EEQ/L (95% confidence interval from 0.019 to 5.4 ng EEQ/L).

##### Benchmark approach

The ER CALUX responses for estrogenic activity at 8 sites with good ecological status are presented in Figure 5. The clean TEQ was 0.06 ng EEQ/L.

**FIGURE 5**

Bioassay responses for estrogenic activity (ng EEQ/L) at eight sites with good ecological status.



Based on the benchmark values and after evaluation of bioassays responses at clean, moderately polluted and heavily polluted sites (Table 4B), we propose a low risk EBT for overall estrogenic activity of 0.5 ng EEQ/L that resembles the HC5 TEQ values of both REP groups. This trigger value is exceeded at sites affected by effluents from waste water treatment plants (wwtp), two moderately polluted and eleven heavily polluted sites.

### 3.2 ANTI-ANDROGENIC ACTIVITY

The complete set of collected toxicity data of anti-androgenic substances is presented in Appendix II B. The lowest toxic concentrations found for the five endpoints are presented in Appendix III. The group of compounds that can inhibit the human androgen receptor and block its action (anti-androgenic response) is very heterogeneous (see Appendix I). It includes estrogenic compounds (e.g. 17 $\alpha$ -ethinylestradiol and estradiol), pesticides (e.g. alachlor, triclosan, vinclozolin), synthetic materials (e.g. bisphenol A and phthalates) and non-ionic surfactants (alkylphenoles).

#### Safe approach

Graphic representations of the collected toxicity values for the REP1 and REP2 groups are presented in Appendix IV as flutamide equivalents per liter (F1EQ/L). The values used for the definition of a safe TEQ did not differ in the REP1 and REP2 groups. The lowest value found in literature was a PNEC for benzo[a]pyrene equal to 0.0017  $\mu$ g/L (OSPAR Agreement, 2014-05). Since benzo[a]pyrene has a REP of 1 in the antiAR CALUX, this PNEC is equal to 0.0017  $\mu$ g FEQ/L. However, the safe TEQ was set based on the lowest LC50 of 0.016  $\mu$ g/L for endosulfan for the copepod *Mesocyclops longisetus* after acute exposure (Gutierrez et al., 2013). The transformed chronic value was equal to 0.005  $\mu$ g F1EQ/L. Assuming a safety factor of 100 for the LC50 endpoint, the final proposed safe TEQ is equal to 0.05 ng F1EQ/L.

#### SSD

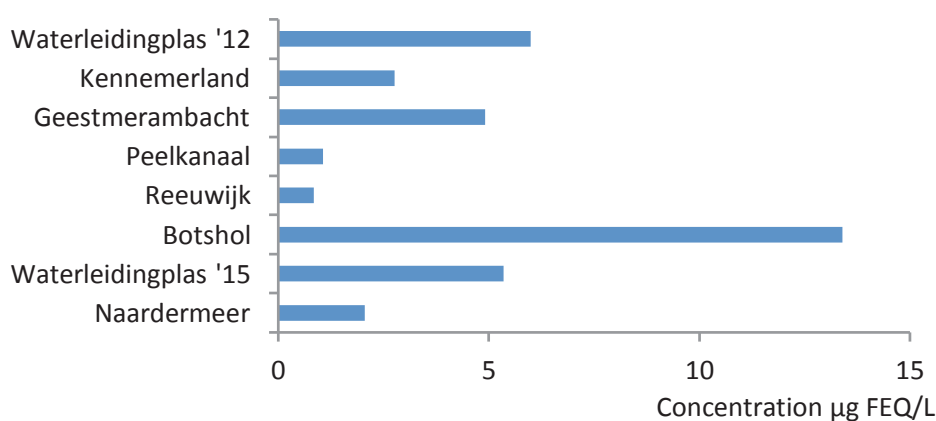
The SSD curves for the REP1 and REP2 groups are presented in Appendix V. The SSD analysis for the REP1 group resulted in a HC5 TEQ equal to 0.29  $\mu$ g F1EQ/L (95% confidence interval from 0.1 to 0.6  $\mu$ g F1EQ/L). The SSD analysis for the REP2 group resulted in a HC5 TEQ equal to 0.13  $\mu$ g F1EQ/L (95% confidence interval from 0.05 to 0.27  $\mu$ g F1EQ/L).

#### Benchmark approach

The anti-AR CALUX responses for estrogenic activity at 8 sites with good ecological status are presented in Figure 6. The mean anti-AR response was equal to 4.55  $\mu$ g F1EQ/L.

**FIGURE 6**

Bioassay responses for anti-androgenic activity ( $\mu$ g FEQ/L) at eight sites with good ecological status.



The range of toxic concentrations collected for compounds that exhibit anti-androgenic activity was very broad, reflecting the different nature of the substances taken into consideration. In this contest, the benchmark approach was considered even more important, since an approach based on the HC5 TEQ value of the SSD analysis would lead to an insufficient discrimination between sites (EBT would be exceeded at all sites). Based on the reference benchmark values and after evaluation of Waternet bioassays responses at polluted sites (Table 4B), we propose a low risk EBT for overall anti-androgenic activity of 25 µg F1EQ/L, which is much higher than the predicted HC5 TEQ and the proposed safe TEQ. This trigger value was exceeded at one of the reference sites, four moderately polluted sites and five heavily polluted sites.

### 3.3 DIOXIN AND DIOXIN-LIKE ACTIVITY

A complete set of collected toxicity data of substances with dioxin-like activity is presented in Appendix II C. The lowest toxic concentrations found for the five endpoints are presented in Appendix III. Dioxin and dioxin-like compounds are poorly water-soluble. They tend to accumulate in organism due to bioaccumulation or biomagnification. Most of the studies reported nominal concentrations of exposure, which may lead to an underestimation of the risk connected to the exposure of aquatic organisms to this group of compounds.

#### Safe approach

Graphic representations of the collected toxicity values for compounds in the REP1 and REP2 groups are presented in Appendix IV as 2,3,7,8-tetrachloro dibenzodioxin (TCDD) equivalent per liter (TEQ/L). In both REP1 and REP2 group, the lowest value found was a LOEC of 2 pg/L for rare minnow (*Gobiocypris rarus*) after chronic exposure to 2,3,7,8-TCDD (Wu et al., 2001), which is equal to the TEQ concentration, since 2,3,7,8-TCDD is the reference compound for the DR CALUX. A safety factor of 5 for LOEC was applied to this value, so the proposed safe TEQ is equal to 0.4 pg TEQ/L.

#### SSD

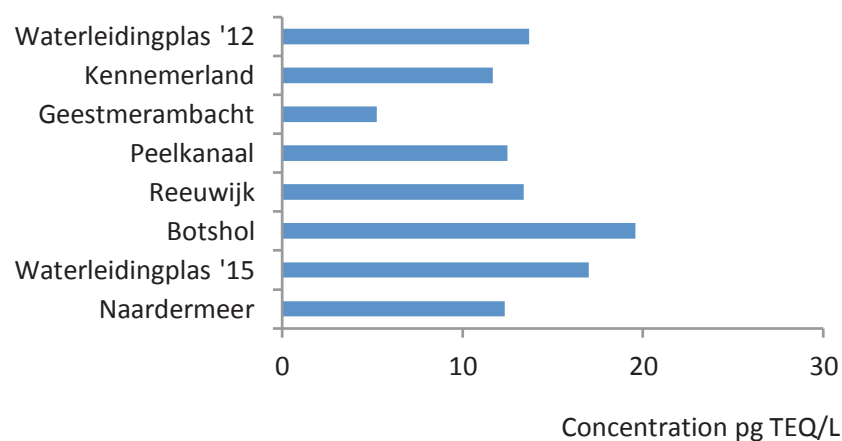
The SSD curves for the REP1 and REP2 groups are presented in Appendix V. The SSD analysis for the REP1 group resulted in a HC5 equal to 36 pg TEQ/L (95% confidence interval from 1.7 to 308 pg TEQ/L). The SSD analysis for the REP2 group resulted in a HC5 equal to 137 pg TEQ/L (95% confidence interval from 15 to 736 pg TEQ/L).

#### Benchmark approach

The DR CALUX responses for dioxin and dioxin-like activity at 8 sites with good ecological status are presented in Figure 7. The clean TEQ was equal to 13.2 pg TEQ/L.

**FIGURE 7**

*Bioassay responses for dioxin and dioxin-like activity (pg TEQ/L) at eight sites with good ecological status.*



Based on this values and after evaluation of Waternet bioassays responses at polluted sites (Table 4C), we propose a low risk EBT for overall dioxin-like activity of 50 pg TEQ/L, which is slightly higher than the HC5 TEQ of the REP1 group, but lower than the HC5 TEQ of the REP2 group. This trigger value is exceeded at seven polluted sites, six of which were considered to be moderately polluted.

### 3.4 GLUCOCORTICOID ACTIVITY

A complete set of collected toxicity data of compounds with glucocorticoid activity is presented in Appendix II D. The lowest toxic concentrations found for the five endpoints are presented in Appendix III. The toxic effects of glucocorticoids for the aquatic community have been poorly investigated. The dataset for both the REP1 and REP2 groups is limited if compared to others activities investigated in the present study. Most studies were conducted on fish, while information for other trophic levels is scarce or inexistent.

#### *Safe approach*

Graphic representations of the collected toxicity values for compounds in the REP1 and REP2 groups are presented in Appendix IV as dexamethasone equivalents per liter (DEQ/L). In both REP1 and REP2 group, the lowest value found was a LOEC of 100 ng/L for fathead minnow (*Pimephales promelas*) after chronic exposure to dexamethasone (Lalone et al., 2012), which is equal to the TEQ concentration, since dexamethasone is the reference compound for the GR CALUX. A safety factor of 5 for LOEC was applied to this value, so the proposed safe TEQ is equal to 20 ng DEQ/L. This safe TEQ is 3 orders of magnitude lower than the only PNEC found in literature for prednisone of 27,800 ng DEQ/L (Escher et al., 2011).

#### *SSD*

The SSD curves for the REP1 and REP2 groups are presented in Appendix V. The SSD analysis for the REP1 group resulted in a HC5 TEQ equal to 3236 ng DEQ/L (95% confidence interval from 80 to 29965 ng DEQ/L). The SSD analysis for the REP2 group resulted in a HC5 TEQ equal to 2145 ng DEQ/L (95% confidence interval from 116 to 14311 pg TEQ/L).

#### *Benchmark approach*

The GR CALUX at eight sites with good ecological status did not show any glucocorticoid activity above the detection limit of 1.2 ng DEQ/L.

Since no GR activity was observed at the clean reference sites, the bioassays responses at polluted sites (Table 4) were leading for the benchmark. We propose low risk EBT for overall glucocorticoid activity of 100 ng DEQ/L, which is five times higher than the safe TEQ of 20 ng DEQ/L. This EBT is only exceeded at three sites heavily affected by wwtp effluents.

### 3.5 OXIDATIVE STRESS

It was not possible to find any toxicity data for aquatic community for the reference compound of this bioassay (i.e. curcumin). However, information was available on the many other compounds that cause oxidative stress to cells and trigger a response in Nrf2 CALUX. The complete set of collected toxicity data of compounds causing oxidative stress is presented in Appendix II E. The lowest toxic concentrations found for the five endpoints are presented in Appendix III.

#### *Safe approach*

Graphic representations of the collected toxicity values for the REP1 and REP2 groups are presented in Appendix IV as curcumin equivalent per liter water (CEQ/L). In the REP1 group, the safe TEQ was based on the lowest EC50 of 0.42 µg/L for zebrafish (*Danio rerio*), after acute exposure to retinoic acid (Selderslaghs et al., 2012). This value was multiplied for an acute to chronic ratio of 0.1 and multiplied by the corresponding REP. The final TEQ value was equal to 0.007 µg CEQ/L. This value was divided by a safety factor of 10 for EC50, which resulted in a proposed safe TEQ of 0.0007 µg CEQ/L. This safe TEQ is one order of magnitude lower than the lowest PNEC of 0.023 µg CEQ/L for carbenzadim (Oekotoxzentrum website, EAWAG).



The lowest values found for all the toxic endpoints in the REP2 group were for estradiol, due to the fact that this compound has a relative potency of 0.06. The proposed safe TEQ of 0.006 ng CEQ/L was based on the lowest NOEC of 0.001 µg/L for *Oryzias latipes* (*Japanese Medaka*) after acute exposure to estradiol (Lee et al., 2012).

### SSD

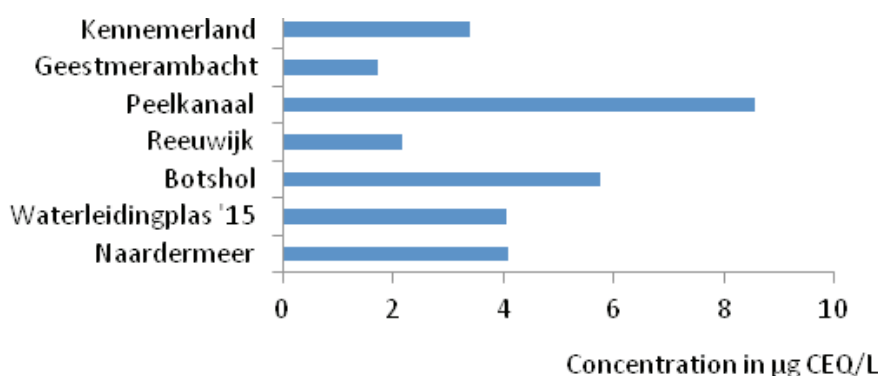
The SSD curves for the REP1 and REP2 groups are presented in Appendix V. The SSD analysis for the REP1 group resulted in a HC5 TEQ equal to 0.7 µg CEQ/L (95% confidence interval from 0.2 to 2.2 µg CEQ/L). The SSD analysis for the REP2 group resulted in a HC5 TEQ equal to 0.034 µg CEQ/L (95% confidence interval from 0.008 to 0.11 µg CEQ/L).

### Benchmark approach

The Nrf2 CALUX responses for oxidative stress at 7 sites with good ecological status are presented in Figure 8. The clean TEQ was equal to 4.25 µg CEQ/L.

**FIGURE 8**

*Bioassay responses for Nrf2 oxidative stress (µg CEQ/L) at seven sites with good ecological status.*



The range of toxic concentrations collected for compounds that cause oxidative stress was (like anti-AR compounds) very broad, reflecting a different nature and toxicity. The low HC5 TEQ value that was found was not useful for designing a realistic EBT, because it would be exceeded at all unpolluted sites. Based upon the mean benchmark value at the reference sites, we propose a low risk EBT for overall oxidative stress activity of 10 µg CEQ/L, which is much higher than the predicted HC5 TEQ values. This EBT was not yet performed at many polluted sites, and no exceedances have been observed thus far (Table 4C).

### 3.6 TOXIC PAHS

The complete set of collected toxicity data of toxic PAHs is presented in Appendix II F. The lowest toxic concentrations found for the five endpoints are presented in Appendix III. As in the case of dioxin and dioxin-like compounds, PAHs are lipophilic compounds that tend to accumulate in soil, organic particulate and tissues rather than in water. For this reason, the concentration of this class of pollutants should be measured during and/or at the end of the exposure period. However, the majority of the reviewed studies reported nominal concentrations of exposure, which may lead to an underestimation of the risks.

### Safe approach

Graphic representations of the collected toxicity values for the REP1 and REP2 groups are presented in Appendix IV as benzo[a]pyrene equivalents per liter water (BEQ/L). In the REP1 group the lowest value found was a PNEC of 0.17 ng/L for

benzo[a]pyrene (OSPAR Agreement, 2014-05). This value was used to set a safe TEQ of 0.17 ng BEQ/L. In the REP2 group the lowest value found was a LOEC of 0.02 ng/L for *Gobiocypris rarus* (Rare minnow) after chronic exposure to benzo[a]pyrene (Wu et al., 2001). This value was divided by a safety factor of 5 for LOEC, which resulted in a proposed safe TEQ of 0.008 ng BEQ/L.

### SSD

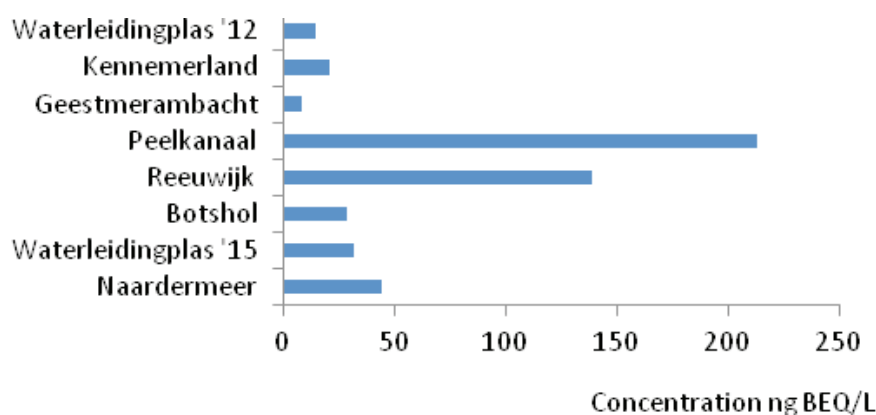
The SSD curves for the REP1 and REP2 groups are presented in Appendix V. The SSD analysis for the REP1 group resulted in a HC5 TEQ equal to 47 ng BEQ/L (95% confidence interval from 2 to 368 ng BEQ/L). The SSD analysis for the REP2 group resulted in a HC5 TEQ equal to 41 ng BEQ/L (95% confidence interval from 2.5 to 254 ng BEQ/L).

### Benchmark approach

The PAH CALUX responses at 8 sites with good ecological status are presented in Figure 9. The clean TEQ was equal to 62.7 ng BEQ/L.

**FIGURE 9**

Bioassay responses for toxic PAHs (ng BEQ/L) at eight sites with good ecological status.



Based on the clean TEQ value and after evaluation of Waternet bioassays responses at polluted sites (Table 4C), we propose a low risk EBT for overall PAH activity of 150 ng BEQ/L. This EBT was above the estimated HC5 TEQ values, but falls within the HC5 95% confidence intervals. The EBT was exceeded at four moderately polluted sites, while not many measurements were performed at heavily polluted sites.

### 3.7 PPAR $\gamma$ RECEPTOR

The complete set of collected toxicity data of PPAR $\gamma$  inducing compounds is presented in Appendix II. The lowest toxic concentrations found for the five endpoints are presented in Appendix III. The *in vitro* PPAR $\gamma$  CALUX is able to detect compounds that activate the PPAR gamma receptor, including several classes of aquatic contaminants, such as organotins and perfluorinated compounds.

### Safe approach

Graphic representations of the collected toxicity values for the REP1 and REP2 groups are presented in Appendix IV as rosiglitazone equivalent per liter water (REQ/L). For the REP1 group the safe TEQ was based on the lowest EC50 found of 424 ng/L for zebrafish (*Danio rerio*) after acute exposure to retinoic acid (Selderslaghs et al., 2012). This value was multiplied by an acute to chronic ratio of 0.1 and transformed to a TEQ value equal to 13.4 ng REQ/L. This value was then divided by

a safety factor of 10, which resulted in a safe TEQ of 1.34 ng REQ/L. In the REP2 approach the safe TEQ was based on the lowest PNEC of 0.14 ng/L for dibenzo[a,h] anthracene (OSPAR Agreement, 2014-05). After TEQ transformation a value of 0.00014 ng REQ/L was defined as the safe TEQ.

### SSD

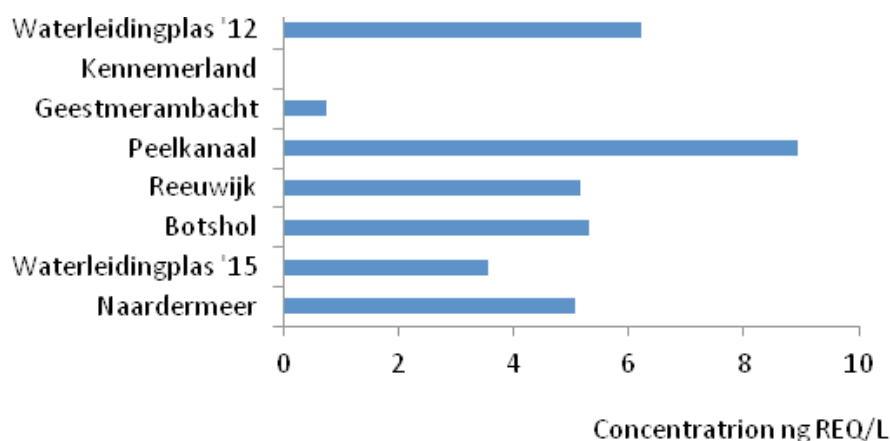
The SSD curves for the REP1 and REP2 groups are presented in Appendix V. The SSD analysis for the REP1 group resulted in a HC5 TEQ equal to 45 ng REQ/L (95% confidence interval from 0.8 to 371 ng REQ/L). The SSD analysis for the REP2 group resulted in a HC5 TEQ equal to 0.3 ng REQ/L (95% confidence interval from 0.002 to 6.9 ng REQ/L).

### Benchmark approach

The PPAR $\gamma$  CALUX responses for at 8 sites with good ecological status are presented in Figure 10. The clean TEQ was equal to 4.37 ng REQ/L.

**FIGURE 10**

*Bioassay responses of PPAR $\gamma$  lipid metabolism (ng REQ/L) at eight sites with good ecological status.*



Based upon the benchmark of unpolluted reference sites and after evaluation of Waternet bioassays responses at polluted sites (Table 4C), we propose a low risk EBT for peroxisome proliferation of 10 ng REQ/L. This value is higher than the HC5 TEQ of the REP2 group, but lower than the HC5 TEQ calculated with REP1 compounds. The ETB was exceeded at two moderately polluted sites and seven heavily polluted sites.

### 3.8 PREGNANE X RECEPTOR

The complete set of collected toxicity data of PXR inducing compounds is presented in Appendix II H. The lowest toxic concentrations found for the five endpoints are presented in Appendix III. The PXR CALUX is able to detect many WFD priority compounds, including pesticides, PAHs and alkyl phenols.

### Safe approach

Graphic representations of the collected toxicity values for the REP1 and REP2 groups are presented in Appendix IV as nicarbidine equivalent per liter water (NEQ/L). In both REP1 and REP2 groups the safe TEQ was based on the lowest LOEC found of 1 ng/L for *Daphnia magna* after acute exposure to chlorpyrifos-ethyl (Ha and Choi, 2009). This value was multiplied by an acute to chronic ratio of 0.1 and transformed to a TEQ value equal to 0.020 ng NEQ/L. This value was then divided by a safety factor of 5 for LOEC, which resulted in a safe TEQ of 0.004 ng NEQ/L. The safe TEQ

is 1 order of magnitude lower than the lowest PNEC found for benzo(k)fluoranthene, equal to 0.03 ng NEQ/L (OSPAR Agreement, 2014-05).

### SSD

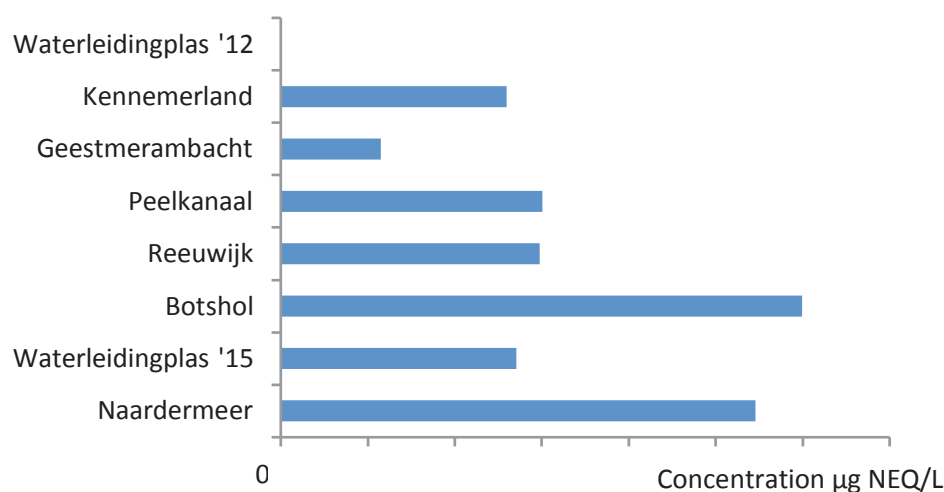
The SSD curves for the REP1 and REP2 groups are presented in Appendix V. The SSD analysis for the REP1 group resulted in a HC5 TEQ equal to 7 ng NEQ/L (95% confidence interval from 1 to 30 ng NEQ/L). The SSD analysis for the REP2 group resulted in a HC5 TEQ equal to 8 ng NEQ/L (95% confidence interval from 2 to 24 ng NEQ/L).

### Benchmark approach

The PXR CALUX responses for biotransformation activity at 7 sites with good ecological status are presented in Figure 11. The clean TEQ was equal to 1.71 µg NEQ/L.

**FIGURE 11**

*Bioassay responses of PXR biotransformation activity (µg NEQ/L) at seven sites with good ecological status.*



The range of toxic concentrations collected for compounds that cause elevated PXR biotransformation was (like for anti-AR and oxidative stress endpoints) very broad, reflecting a different nature and toxicity. The low HC5 TEQ value that was found was not useful for designing a realistic EBT, because it would be exceeded at all unpolluted sites. Based upon the mean benchmark value at the reference sites, we propose a low risk EBT for overall PXR activity of 3 µg CEQ/L, which is much higher than the predicted HC5 TEQ values. This bioassay was not yet performed at many polluted sites, and only three exceedances have been observed thus far (Table 4C).

## 3.9 ANTIBIOTICS ACTIVITIES

According to their mode of action (MOA) antibiotics are generally divided into five classes: amidoglycosides, macrolides & β-lactams, sulfonamides, tetracyclines and quinolones. Since the bioassay determines the activities of all classes of antibiotics five separate ETVs were developed.

### 3.9.1 Aminoglycosides

The complete set of collected toxicity data for compounds with aminoglycosides activity is presented in Appendix II I.1. The lowest toxic concentrations found for the five endpoints are presented in Appendix III.

#### *Safe approach*

There were no compounds within this group of antibiotics with REPs lower than 0.1. Graphic representation of the collected toxicity values for the REP1 group is presented in Appendix IV as neomycin equivalents per liter water (NEQ/L). The safe TEQ was based on the lowest PNEC found of 300 ng/L for neomycin (Park and Choi, 2008). Neomycin is the reference compound for this group of antibiotic. As a consequence, the TEQ value was also equal to 300 ng NEQ/L. This PNEC was calculated applying an assessment factor of 100 on the neomycin chronic NOEC for *Daphnia magna* (0.03 mg/L) (Park and Choi, 2008). This value is therefore the proposed safe TEQ for aminoglycosides group.

#### *SSD*

The SSD curve for the REP1 group is presented in Appendix V. The SSD analysis resulted in a very high HC5 TEQ equal to 33222 ng NEQ/L (95% confidence interval from 1546 to 219614 ng NEQ/L).

#### *Benchmark approach*

No detectable aminoglycosides activity (>90 ng NEQ/L) was found at the eighth clean reference sites. Moreover, most of the aminoglycosides activities found in the environment by Waternet in the years 2010-2014 were below detectable levels, apart from some sites that were affected by wwtp effluents (Table 4B).

Mainly based upon the benchmark, we propose a low-risk EBT of 500 ng NEQ/L, which is twice the safe TEQ of 300 ng NEQ/L. The EBT was only exceeded at two sites with a significant wwtp influence (Table 4B).

### **3.9.2 Macrolides and $\beta$ -Lactams**

The complete dataset of collected toxicity values of compounds with macrolides activity is presented in Appendix II I.2. The lowest toxic concentrations found for the five endpoints are presented in Appendix III.

#### *Safe approach*

Graphic representations of the collected toxicity values for the REP1 and REP2 groups are presented in Appendix IV as penicillin equivalents per liter water (PEQ/L). In the REP1 group the safe TEQ was based upon the lowest PNEC found of 3.7 ng/L for amoxicillin (Jones et al., 2002). The safe TEQ for this group of antibiotics was derived after transformation to a TEQ value of 2.22 ng PEQ/L. In the REP2 group, the safe TEQ was equal to 1.8 ng PEG/L, calculated from the lowest EC50 for *Microcystis aeruginosa* after chronic exposure to 18 ng/L tiamulin (Halling-Sorensen, 2000).

#### *SSD*

The SSD curves for the REP1 and REP2 groups are presented in Appendix V. The SSD analysis for the REP1 group resulted in a HC5 TEQ equal to 2615 ng PEQ/L (95% confidence interval from 96 to 23135 ng PEQ/L). The SSD analysis for the REP2 group resulted in a HC5 TEQ equal to 98 ng PEQ/L (95% confidence interval from 12.9 to 470 ng PEQ/L).

#### *Benchmark approach*

No detectable activities of macrolides and  $\beta$ -lactams (>1.4 PEQ/L) were found at the eight clean reference sites. The concentrations of macrolides and  $\beta$ -lactams antibiotics (e.g. penicillin) collected by Waternet passive sampling demonstrate that this group of antibiotics is the most commonly found in Dutch waters (Table 4B), with detectable concentrations especially at sites influenced by wwtp effluents.

Considering these data and the low safe TEQ, a low risk EBT of 50 ng/L is proposed. The proposed EBT is lower than both HC5 TEQ estimations, but the lower limit of the HC5 95% confidence interval for the REP2 group is below this EBT. The EBT was exceeded at four sites with a significant wwtp influence (Table 4B).

### 3.9.3 Sulfonamides

The complete dataset of collected toxicity values of compounds with sulfonamides activity is presented in Appendix II I.3. The lowest toxic concentrations found for the five endpoints are presented in Appendix III.

#### *Safe approach*

There were no compounds considered for this bioassay with REPs lower than 0.1. Graphic representation of the collected toxicity values for the REP1 group is presented in Appendix IV as sulfamethoxazole equivalents per liter water (SEQ/L). The safe TEQ was based on the lowest LOEC found of 1000 ng/L for zebra fish (*Danio rerio*) after acute exposure to sulfadiazine (Lin et al., in press). The TEQ value was equal to 50 ng SEQ/L. After application of a safety factor of 5 for LOEC, the proposed safe TEQ for this group of antibiotics is equal to 10 ng SEQ/L.

#### *SSD*

The SSD curve for the REP1 group is presented in Appendix V. The SSD analysis resulted in a HC5 TEQ equal to 67037 ng SEQ/L (95% confidence interval from 24675 to 148222 ng SEQ/L).

#### *Benchmark approach*

A sulfonamide activity of 37 ng SEQ/L was found in one of the reference sites, while activities at the other seven clean sites were all below the detection limit of 2 ng SEQ/L. The clean TEQ was equal to 4.6 SEQ/L.

Considering the high HC5 TEQ values, we propose a low risk EBT of 100 ng SEQ/L, which is ten times the safe TEQ. The EBT was exceeded at seven sites with a significant wwtp influence (Table 4B).

### 3.9.4 Tetracyclines

The complete dataset of collected toxicity values of compounds with tetracycline activity is presented in Appendix II I.4. The lowest toxic concentrations found for the five endpoints are presented in Appendix III.

#### *Safe approach*

Graphic representations of the collected toxicity values for the REP1 and REP2 groups are presented in Appendix IV as oxytetracycline equivalents per liter water (OEQ/L). In both REP1 and REP2 groups the safe TEQ was based upon the lowest PNEC found of 170 ng/L for oxytetracycline (Park and Choi, 2008). As for sulfonamides, these authors used the lowest value they could find, in this case the acute LC50 of 0.17 mg/L for the algae *Selenastrum capricornutum* after 3 days of exposure, reported by Nunes et al. (2005), and applied a safety factor of 1000 to calculate the resulting PNEC. The corresponding TEQ value and proposed safe TEQ for this group of antibiotics is thus equal to 170 ng OEQ/L.

#### *SSD*

The SSD curves for the REP1 and REP2 groups are presented in Appendix V. The SSD analysis for the REP1 group resulted in a HC5 TEQ equal to 32931 ng OEQ/L (95% confidence interval from 9837 to 83368 ng OEQ/L). The SSD analysis for the REP2 group resulted in a HC5 TEQ equal to 27275 ng OEQ/L (95% confidence interval from 8292 to 68544 ng OEQ/L).

#### *Benchmark approach*

No detectable tetracycline activity (>22 ng OEQ/L) was found at the eighth clean reference sites. Tetracyclines-like environmental activities were above detection limit in four Waternet samples, with a maximum of 104 ng OEQ/L close to a wwtp discharge (Table 4B). We propose a low risk EBT for tetracyclines activity of 250 ng OEQ/L, which is about twice the safe TEQ. Tetracycline activities above the detection limit were only observed at five sites, all below the proposed EBT (Table 4B).

### 3.9.5 Quinolones

The complete dataset of collected toxicity values of compounds with quinolone activity is presented in Appendix II I.5.

The lowest toxic concentrations found for the five endpoints are presented in Appendix III. Although it is not used as an antibiotic, triclosan data are also included because this substance induces a clear response in the quinolones bioassay. Triclosan is an antibacterial and antifungal substance often used in personal care products like soaps (Riva et al., 2012).

#### *Safe approach*

There were no compounds within this group with REPs lower than 0.1. Graphic representation of the collected toxicity values for the REP1 group is presented in Appendix IV as flumequine equivalents per liter water (FEQ/L). The safe TEQ was based on the lowest EC50 found of 530 ng/L for the algae *Selenastrum capricornutum* after acute exposure to triclosan (Yang et al., 2008). The corresponding TEQ value was equal to 5.3 ng FEQ/L. The safe TEQ was established as 0.53 ng SEQ/L after application of a safety factor of 10 for EC50.

#### *SSD*

The SSD curve for the REP1 group is presented in Appendix V. The SSD analysis resulted in a HC5 TEQ equal to 8759 ng FEQ/L (95% confidence interval from 2197 to 26050 ng FEQ/L).

#### *Benchmark approach*

No detectable quinolones activity (>44 ng FEQ/L) was found at any of the eighth clean reference sites. Considering the high HC5 TEQ and the low safe TEQ due to triclosan, a low risk EBT of 100 ng/L (i.e. 200x the safe TEQ value) is proposed for this group of antibiotics. Quinolones activity below the EBT was only detected three Waternet sites thus far (Table 4B), and the EBT was never exceeded.

### **3.10 Overview and evaluation of ebt development**

The effects measured in passive sampler extracts were converted to estimated water effect by a method proposed in Van der Oost et al. (in preparation b). Water TU or TEQ levels were divided by the proposed low-risk trigger values (EBT). It is demonstrated that EBTs are only exceeded at the clean reference sites, and that most EBT exceedance is observed at sites that were polluted by pesticides (Legmeerpolders) or in raw wwtp effluents (Amstelveen and Hilversum). The results of the benchmark studies at sites that were classified as clean, moderately polluted and heavily polluted, carried out by Waternet and STOWA, are presented in Table 4. This table is divided into three separate sections for A. bioassays for general toxicity (for which the EBT were not determined in this study), B. specific bioassays that were applied on the polar extracts of POCIS passive samplers, and C. specific bioassays that were applied on the non-polar extracts of silicon rubber passive samplers. All benchmark results are summarized in the heatmap of Table 5, and compared to the SIMONI scores that were calculated with the entire bioassay battery. A SIMONI score above 1 indicates a potential risk of the ecosystem due to micropollutant exposure.

The benchmark studies, together with the SSD analyses, were used to define low-risk trigger values that will be used for the environmental hazard identification of micropollutants. All relevant results for the determination of the proposed EBTs, using the compounds selected for both REP groups 1 (>0.1) and 2 (>0.001), are summarized in Table 6.

**TABLE 4A**

Benchmark results of relative bioassay responses (response/EBT) for general toxicity; orange = response above EBT, dark green = response below EBT; light green = no response; white = not measured.

General toxicity						
Sites	year	field	bacteria	algae	daphnids	cytotox
	polar	%/EBT	TU/EBT	TU/EBT	TU/EBT	TU/EBT
	non-polar					
<b>Effect-Based Triggervalue (LR-EBT)</b>		<b>20</b>	<b>0,05</b>	<b>0,05</b>	<b>0,05</b>	<b>0,05</b>
<b>clean</b>						
Waterleidingplas '12	2012	0,00	0,15	0,00	0,00	0,00
Naardermeer	2015	0,00	0,04	0,00	0,00	0,00
Waterleidingplas '15	2015	0,50	0,00	0,00	0,00	0,00
Botshol	2015	0,25	0,06	0,00	0,00	0,00
Reeuwijk	2015	0,00	0,09	0,00	0,00	0,00
Peelkanaal	2015	1,50	0,10	0,00	0,00	0,00
Geestmerambacht	2015	0,00	0,03	0,00	0,00	0,00
Kennemerland	2015	0,00	0,06	0,00	0,00	0,00
<b>LR-EBT exceedance</b>		<b>12,5%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>0,0%</b>
<b>moderately polluted</b>						
Amstelveen A1-1	2010		0,00	0,00	0,00	0,00
Amstelveen A2-1	2010		0,00	0,00	0,00	0,00
Amstelveen A1-2	2011		0,90	0,58	0,00	0,00
Amstelveen A2-2	2011		0,74	0,00	0,00	0,00
KRW spagaat Zodden	2011		0,00	0,00	0,00	0,00
KRW spagaat Strook	2011		0,00	0,00	0,00	0,00
KRW spagaat Vecht	2011		0,00	0,59	0,00	0,00
KRW spagaat WL kanaal	2011		0,00	0,00	0,00	0,00
KRW spagaat Zodden	2011		0,07	0,13	0,04	0,00
KRW spagaat Strook	2011		0,02	0,04	0,02	0,00
KRW spagaat Vecht	2011		0,06	0,05	0,04	0,00
KRW spagaat WL kanaal	2011			0,05	0,02	0,00
Amstel voor Uithoorn	2012-1	1,50				
Amstel na Uithoorn	2012-1	0,00				
Amstel voor Uithoorn	2012-2	1,50	0,08	0,04	0,15	0,00
Amstel na Uithoorn	2012-2	0,00	0,18	0,07	0,45	0,00
Vecht Utrecht	2012	0,00	0,06	0,03	0,31	0,00
Vecht Loenen	2012	0,00	0,24	0,04	0,24	0,00
Weesp near Solvay	2012	0,00	0,13	0,02	0,02	0,00
Zevenhoven	2013-1	0,00	0,14	0,10	0,00	0,00
Zevenhoven	2013-2	0,75	0,34	0,00	0,53	0,00
Horstermeer	2014	1,25	0,03	0,02	0,05	0,01
Uithoorn	2014	0,00	0,00	0,00	0,07	0,00
Ronde Venen	2014	0,00	0,00	0,02	0,07	0,01
Amstelveen	2014	0,00	0,02	0,02	0,05	0,00
Amstelveen '15	2015	0,00	0,02	0,02	0,17	0,00
Ronde Venen '15	2015	0,00	0,03	0,00	0,10	0,00
Femmeer	2015	0,50	0,03	0,00	0,07	0,00
<b>LR-EBT exceedance</b>		<b>23,1%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>0,0%</b>
<b>heavily polluted</b>						
wwtp Amstelveen A3-1	2010		0,00	0,00	0,00	0,00
wwtp Amstelveen A3-2	2011		0,75	0,97	0,00	0,00
Zuider Legmeerpolder	2012-1	5,00				
Noorder Legmeerpolder	2012-1	0,50				
Zuider Legmeerpolder	2012-2	0,25	0,26	0,08	7,63	0,00
Noorder Legmeerpolder	2012-2	3,50	0,19	0,07	1,17	0,00
Zuider Legmeerpolder 1	2013-1	3,00	0,19	0,06	0,85	0,00
Zuider Legmeerpolder 2	2013-1	2,25	0,45	0,07	0,15	0,00
Zuider Legmeerpolder 3	2013-1	3,50	0,09	0,02	0,22	0,00
Zuider Legmeerpolder 4	2013-1	3,00	0,24	0,26	1,14	0,00
Zuider Legmeerpolder 5	2013-1	0,00	0,05	0,02	0,07	0,00
Noorder Legmeerpolder 1	2013-1	2,75	0,02	0,00	0,04	0,00
Noorder Legmeerpolder 2	2013-1	1,25	0,07	0,00	0,09	0,00
Zuider Legmeerpolder 1	2013-2	1,25	0,07	0,04	0,24	0,00
Zuider Legmeerpolder 2	2013-2	2,50	0,26	0,08	0,00	0,00
Zuider Legmeerpolder 3	2013-2	0,50	0,26	0,00	1,10	0,00
Zuider Legmeerpolder 4	2013-2	5,00	0,15	0,00	0,74	0,00
Zuider Legmeerpolder 5	2013-2	0,00	0,13	0,05	0,59	0,00
Noorder Legmeerpolder 1	2013-2	2,75	0,37	0,00	0,00	0,00
Noorder Legmeerpolder 2	2013-2	0,25	0,37	0,00	0,00	0,00
wwtp Hilversum	2014	1,00	0,03	0,02	0,06	0,01
Hilversum '15	2015	1,75	0,02	0,00	0,02	0,00
Blaricum	2015	1,00	0,02	0,00	0,08	0,01
<b>LR-EBT exceedance</b>		<b>78,9%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>19,0%</b>	<b>0,0%</b>



TABLE 4B

Benchmark results of relative bioassay responses (response/EBT) for specific toxicity in polar PS extracts; orange = response above EBT, dark green = response below EBT; light green = no response; white = not measured.

Specific toxicity & antibiotics in polar PS extracts									
Sites	year	ER	anti-AR	GR	amino	macro	sulfon	tetra	quino
	polar	eq/EBT	eq/EBT	eq/EBT	eq/EBT	eq/EBT	eq/EBT	eq/EBT	eq/EBT
<b>Effect-Based Triggervalue (LR-EBT)</b>		<b>0,5</b>	<b>25</b>	<b>100</b>	<b>500</b>	<b>50</b>	<b>100</b>	<b>250</b>	<b>100</b>
clean									
Maarsseveense plassen	2010				0,00	0,50	0,00	0,00	0,00
Waterleidingplas '12	2012	0,02	0,18	0,00	0,00	0,00	0,00	0,00	0,00
Naardermeer	2015	0,04	0,08	0,00	0,00	0,00	0,00	0,00	0,00
Waterleidingplas '15	2015	0,03	0,21	0,00	0,00	0,00	0,37	0,00	0,00
Botshol	2015	0,02	0,54	0,00	0,00	0,00	0,00	0,00	0,00
Reeuwijk	2015	0,03	0,03	0,00	0,00	0,00	0,00	0,00	0,00
Peelkanaal	2015	0,21	0,04	0,00	0,00	0,00	0,00	0,00	0,00
Geestmerambacht	2015	0,55	2,36	0,00	0,00	0,00	0,00	0,00	0,00
Kennemerland	2015	0,07	0,11	0,00	0,00	0,00	0,00	0,00	0,00
<b>LR-EBT exceedance</b>		<b>0,0%</b>	<b>12,5%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>0,0%</b>
moderately polluted									
Amstelveen A1-1	2010	0,48		0,00	0,00	0,00	0,00	0,00	0,00
Amstelveen A2-1	2010	0,38		0,00	0,00	0,00	0,00	0,00	0,00
Vecht 1	2010				0,00	0,00	0,00	0,00	0,00
Vecht 2	2010				0,00	0,31	0,00	0,00	0,00
Vecht 3	2010				0,00	0,00	0,00	0,00	0,00
Vecht 4	2010				0,00	0,00	0,00	0,00	0,00
Vecht 5	2010				0,00	0,00	0,00	0,00	0,00
Vecht 6	2010				0,00	0,31	0,00	0,00	0,00
Amstelveen A1-2	2011	0,63	1,02	0,00	0,00	0,00	0,00	0,00	0,00
Amstelveen A2-2	2011	0,38	1,36	0,00	0,00	0,00	0,00	0,00	0,00
KRW spagaat Zodden	2011	0,09	0,40	0,00	0,00	0,00	0,00	0,00	0,00
KRW spagaat Strook	2011	0,00	1,31	0,00	0,00	0,00	0,00	0,00	0,00
KRW spagaat Vecht	2011	0,81	0,47	0,00	0,00	0,00	0,00	0,00	0,00
KRW spagaat WL kanaal	2011	0,87	0,23	0,00	0,00	0,00	0,00	0,00	0,00
Amstel voor Uithoorn	2012	0,07	0,63	0,00	0,00	0,00	0,00	0,00	0,00
Amstel na Uithoorn	2012	0,14	1,21	0,00	0,00	0,08	0,00	0,00	0,00
Vecht Utrecht	2012	0,35	0,67	0,04	0,25	0,23	0,00	0,00	0,00
Vecht Loenen	2012	0,11	0,30	0,00	0,00	0,00	0,00	0,00	0,00
Weesp nabij Solvay	2012	0,27	0,48	0,00	0,00	0,00	0,00	0,00	0,00
Zevenhoven	2013	0,00	0,00	0,00	0,00	0,00	0,00	0,00	0,00
Horstermeer	2014	1,21	0,70	0,00	0,00	1,40	0,24	0,06	0,00
Uithoorn	2014	0,79	0,29	0,29	1,09	1,68	1,36	0,00	0,00
Ronde Venen	2014	0,43	0,75	0,05	0,00	0,63	0,00	0,00	0,00
Amstelveen	2014	0,38	0,69	0,02	0,26	0,22	0,33	0,00	0,00
Amstelveen '15	2015	0,67	0,47	0,13	0,46	0,18	0,42	0,00	0,00
Ronde Venen '15	2015	0,36	0,26	0,10	0,40	0,11	0,30	0,00	0,49
Eemmeer	2015	3,29	0,46	0,73	0,16	0,27	1,40	0,09	0,00
<b>LR-EBT exceedance</b>		<b>9,5%</b>	<b>21,1%</b>	<b>0,0%</b>	<b>3,7%</b>	<b>7,4%</b>	<b>7,4%</b>	<b>0,0%</b>	<b>0,0%</b>
heavily polluted									
Amstelveen A3-1	2010	1,00		0,43	0,11	1,06	2,39	0,00	0,00
Amstelveen A3-2	2011	3,62	0,87	0,00	0,13	0,56	1,17	0,00	0,00
Zuider Legmeerpolder	2012	0,17	1,44	0,00	0,00	0,00	0,63	0,00	0,00
Noorder Legmeerpolder	2012	0,23	2,66	0,00	0,00	0,00	0,35	0,00	0,00
Zuider Legmeerpolder 1	2013-1	1,21	0,92	0,00	0,00	0,00	0,00	0,00	0,00
Zuider Legmeerpolder 2	2013-1	1,60	0,95	0,00	0,00	0,00	0,00	0,00	0,00
Zuider Legmeerpolder 3	2013-1	1,26	1,02	0,00	0,00	0,00	0,00	0,00	0,00
Zuider Legmeerpolder 4	2013-1	0,45	0,50	0,00	0,00	0,00	0,00	0,00	0,00
Zuider Legmeerpolder 5	2013-1	1,26	0,54	0,00	0,00	0,00	0,00	0,00	0,00
Noorder Legmeerpolder 1	2013-1	0,45	0,91	0,00	0,00	0,00	0,00	0,00	0,00
Noorder Legmeerpolder 2	2013-1	0,50	0,47	0,00	0,00	0,00	0,00	0,00	0,00
Zuider Legmeerpolder 1	2013-2	1,36	0,56	0,00	0,00	0,00	0,00	0,00	0,00
Zuider Legmeerpolder 2	2013-2	0,29	0,63	0,00	0,00	0,00	0,00	0,00	0,00
Zuider Legmeerpolder 3	2013-2	0,98	0,51	0,00	0,00	0,00	0,00	0,00	0,00
Zuider Legmeerpolder 4	2013-2	0,31	1,08	0,00	0,00	0,00	0,00	0,00	0,00
Zuider Legmeerpolder 5	2013-2	0,40	1,05	0,00	0,00	0,00	0,00	0,00	0,00
Noorder Legmeerpolder 1	2013-2	0,45	0,60	0,00	0,00	0,00	0,00	0,00	0,00
Noorder Legmeerpolder 2	2013-2	1,60	0,00	0,00	0,00	0,00	0,00	0,00	0,00
Hilversum	2014	5,52	1,73	2,70	1,83	2,29	2,29	0,42	0,00
Hilversum '15	2015	5,24	0,39	2,26	0,20	0,38	3,49	0,19	0,73
Blaricum	2015	7,80	0,54	1,83	0,10	0,37	1,40	0,14	0,24
<b>LR-EBT exceedance</b>		<b>52,4%</b>	<b>25,0%</b>	<b>14,3%</b>	<b>4,8%</b>	<b>9,5%</b>	<b>23,8%</b>	<b>0,0%</b>	<b>0,0%</b>

**TABLE 4C**

Benchmark results of relative bioassay responses (response/EBT) for specific toxicity in non-polar PS extracts; orange = response above EBT, dark green = response below EBT; light green = no response; white = not measured.

Specific toxicity in non-polar PS extracts								
	year	DR	PPARg	PAH	Nrf2	PXR	p53-	p53+
	non-polar	eq/EBT	eq/EBT	eq/EBT	eq/EBT	eq/EBT	TU/EBT	TU/EBT
<b>Effect-based trigger value (LR-EBT)</b>		<b>50</b>	<b>10</b>	<b>150</b>	<b>10</b>	<b>3</b>	<b>0,005</b>	<b>0,005</b>
<b>clean</b>								
Waterleidingplas '12	2012	0,24	0,28	0,09			0,00	
Naardermeer	2015	0,25	0,25	0,30	0,41	0,91	0,00	1,30
Waterleidingplas '15	2015	0,34	0,18	0,21	0,41	0,45	0,00	0,00
Botshol	2015	0,39	0,26	0,19	0,58	1,00	0,00	0,00
Reeuwijk	2015	0,27	0,26	0,93	0,22	0,50	0,00	0,00
Peelkanaal	2015	0,25	0,45	1,42	0,86	0,50	0,00	0,00
Geestmerambacht	2015	0,10	0,04	0,05	0,17	0,19	0,00	0,00
Kennemerland	2015	0,23	0,00	0,14	0,34	0,43	0,00	0,00
<b>LR-EBT exceedance</b>		<b>0,0%</b>	<b>0,0%</b>	<b>12,5%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>14,3%</b>
<b>moderately polluted</b>								
Amstelveen A1-1	2010	0,01		0,06			0,00	
Amstelveen A2-1	2010	0,00		0,02			0,16	
Amstelveen A1-2	2011	0,00	0,05	0,38			0,00	
Amstelveen A2-2	2011	0,00	0,00	0,81			0,00	
KRW spagaat Zodden	2011	0,00	0,10	0,23			0,00	
KRW spagaat Strook	2011	0,00	0,30	0,15			0,10	
KRW spagaat Vecht	2011	0,00	0,04	0,50			0,12	
KRW spagaat WL kanaal	2011	0,00	0,05	0,14			0,14	
Amstel voor Uithoorn	2012	0,60	0,14	1,37			0,00	
Amstel na Uithoorn	2012	0,64	0,32	1,93			0,00	
Vecht Utrecht	2012	0,39	0,17	0,94			0,00	
Vecht Loenen	2012	0,36	0,22	1,09			0,00	
Weesp nabij Solvay	2012	0,17	0,18	0,35			0,00	
Zevenhoven	2013-1	0,15	2,59				0,00	
Zevenhoven	2013-2	0,24	0,90				0,00	
Horstermeer	2014	0,38	0,00		0,31		0,28	
Uithoorn	2014	1,01	0,00		0,15		0,40	
Ronde Venen '14	2014	1,10	0,00		0,29		0,36	
Amstelveen '14	2014	0,45	0,00		0,11		0,00	
De Sniep	2015	0,92		0,44				
Vecht bij Utrecht	2015	0,82		0,54				
Vecht bij Oud-Zuilen	2015	1,14		0,62				
Vecht bij Loenen	2015	5,44		2,04				
Vecht bij Nederhorst	2015	0,95		0,58				
Vecht bij Nigtevecht	2015	1,35		0,65				
Vecht bij Uitermeer	2015	1,06		0,30				
Amstelveen '15	2015	0,74	0,29		0,15	1,23	0,00	
Ronde Venen '15	2015	1,24	0,20		0,47	1,28	0,00	
Femmeer	2015	0,68	0,46		0,43	0,45	0,28	
<b>LR-EBT exceedance</b>		<b>24,1%</b>	<b>5,0%</b>	<b>20,0%</b>	<b>0,0%</b>	<b>66,7%</b>	<b>0,0%</b>	
<b>heavily polluted</b>								
rwzi Amstelveen A3-1	2010	0,02		0,11			0,00	
rwzi Amstelveen A3-2	2011	0,00	0,07	0,14			0,00	
Zuider Legmeerpolder	2012	0,34	0,26	0,28			6,34	
Noorder Legmeerpolder	2012	0,27	0,14	0,41			0,00	
Zuider Legmeerpolder 1	2013-1	0,53	0,00				0,00	
Zuider Legmeerpolder 2	2013-1	0,30	0,79				1,06	
Zuider Legmeerpolder 3	2013-1	0,19	0,34				0,00	
Zuider Legmeerpolder 4	2013-1	0,54	1,17				2,86	
Zuider Legmeerpolder 5	2013-1	0,17	0,16				0,00	
Noorder Legmeerpolder 1	2013-1	0,31	0,42				0,00	
Noorder Legmeerpolder 2	2013-1	0,42	0,00				0,00	
Zuider Legmeerpolder 1	2013-2	0,42	0,33				0,00	
Zuider Legmeerpolder 2	2013-2	0,46	0,85				0,00	
Zuider Legmeerpolder 3	2013-2	1,09	1,76				0,00	
Zuider Legmeerpolder 4	2013-2	0,39	1,88				0,00	
Zuider Legmeerpolder 5	2013-2	0,44	1,52				0,00	
Noorder Legmeerpolder 1	2013-2	0,41	0,56				0,00	
Noorder Legmeerpolder 2	2013-2	0,84	0,00				0,00	
rwzi Hilversum	2014	0,54	0,00		0,21		0,24	
Hilversum '15	2015	0,83	0,34		0,43	1,47	0,44	
Blaricum	2015	0,70	0,54		0,39	0,80	0,00	
<b>LR-EBT exceedance</b>		<b>4,8%</b>	<b>20,0%</b>	<b>0,0%</b>	<b>0,0%</b>	<b>50,0%</b>	<b>14,3%</b>	<b>0,0%</b>

**TABLE 5**

Heatmap of all benchmark results of relative bioassay responses (response/EBT) and SIMONI 1.2 scores for overall ecological risks; orange = response above EBT, dark green = response below EBT; light green = no response; white = not measured, red: SIMONI score > 1, indication of potential environmental risks due to micropollutants.

Sites	year	General toxicity					Specific toxicity										Antibiotics					TOTAL
		field	bact	algae	daphnid	cytotox	ER	anti-AR	GR	DR	PPARG	PAH	Nrf2	PXR	p53-	p53+	amino	macro	sulfon	tetra	quino	
Effect-based trigger value		0,05	0,05	0,05	0,05	0,5	25	100	50	10	150	10	3	0,005	0,005	500	50	100	250	100	SIMONI 1.2	
<b>Clean</b>																						
Waterleidingplas '12	2012																					0,2
Naardermeer	2015																					0,3
Waterleidingplas '15	2015																					0,3
Botshol	2015																					0,4
Reeuwijk	2015																					0,2
Peelkanaal	2015																					0,6
Geestmerambacht	2015																					0,4
Kennemerland	2015																					0,1
<b>Moderately polluted</b>																						
KRW spagaat Zodden	2011																					0,2
KRW spagaat Strook	2011																					0,3
KRW spagaat Vecht	2011																					0,3
KRW spagaat WL kanaal	2011																					0,3
Amstel voor Uithoorn	2012																					0,8
Amstel na Uithoorn	2012																					0,6
Vecht Utrecht	2012																					0,4
Vecht Loenen	2012																					0,3
Weesp nabij Solvay	2012																					0,2
Zevenhoven	2013																					1,0
Zevenhoven	2013																					0,9
Horstermeer	2014																					0,6
Uithoorn	2014																					0,4
Ronde Venen '14	2014																					0,4
Amstelveen '14	2014																					0,6
De Sniep	2015																					-
Vecht bij Utrecht	2015																					-
Vecht bij Oud-Zuilen	2015																					-
Vecht bij Loenen	2015																					-
Vecht bij Nederhorst	2015																					-
Vecht bij Nigtevecht	2015																					-
Vecht bij Uitermeer	2015																					-
Amstelveen '15	2015																					0,5
Ronde Venen '15	2015																					0,4
Eemmeer	2015																					0,8
<b>Heavily polluted</b>																						
Zuider Legmeerpolder	2012																					3,1
Noorder Legmeerpolder	2012																					1,5
Zuider Legmeerpolder	2013-1																					1,4
Zuider Legmeerpolder	2013-1																					1,4
Zuider Legmeerpolder	2013-1																					2,0
Zuider Legmeerpolder	2013-1																					0,3
Noorder Legmeerpolder	2013-1																					1,0
Noorder Legmeerpolder	2013-1																					0,5
Zuider Legmeerpolder	2013-2																					0,8
Zuider Legmeerpolder	2013-2																					1,1
Zuider Legmeerpolder	2013-2																					1,2
Zuider Legmeerpolder	2013-2																					2,1
Zuider Legmeerpolder	2013-2																					0,8
Noorder Legmeerpolder	2013-2																					1,2
Noorder Legmeerpolder	2013-2																					0,5
Hilversum '14	2014																					1,6
Hilversum '15	2015																					1,6
Blaricum	2015																					1,5
<b>EBT exceedances</b>		19	0	0	4	0	13	11	3	7	9	5	0	3	3	1	2	4	4	0	0	

TABLE 6

Summary of the most relevant information for the EBT derivation for *in vitro* bioassays.

Endpoint*	REP2 > 0.001			REP1 > 0.1			EBT TEQ
	Safe TEQ	HC5 TEQ (range)	Safe TEQ	HC5 TEQ (range)	Clean TEQ		
<b>Estrogenic activity</b>	0.0066	0.52	0.016	0.47	0.06	<b>0.5</b>	
ER CALUX [ng EEQ/L]	LOEC/estrone	(0.019-5.4)	LOEC/17 $\alpha$ -ethinyl estradiol	(0.009-6.2)			
<b>Anti-androgenic</b>	0.00005	0.13	0.00005	0.29	4.6	<b>25</b>	
antiAR CALUX [ $\mu$ g F1EQ/L]	LC50/endsulfan	(0.05-0.27)	LC50/endsulfan	(0.1-0.6)			
<b>Dioxin and dioxin-like</b>	0.4	137	0.4	36	13.2	<b>50</b>	
DR CALUX [pg TEQ/L]	LOEC/2,3,7,8-TCDD	(15-736)	LOEC/TCDD	(1.7-308)			
<b>Glucocorticoid</b>	20	2145	20	3236	<LOD	<b>100</b>	
GR CALUX [ng DEQ/L]	LOEC/dexamethasone	(116-14311)	LOEC/dexamethasone	(80-29965)			
<b>PPAR<math>\gamma</math> receptor</b>	0.00014	0.3	1.34	45	4.4	<b>10</b>	
PPAR $\gamma$ CALUX[ng REQ/L]	PNEC/dibenzo[a,h]anthracene	(0.002-6.9)	EC50/retinoic acid	(0.8-371)			
<b>Toxic PAHs</b>	0.04	41	0.17	47	63	<b>150</b>	
PAH CALUX [ng BEQ/L]	LOEC/2,3,7,8-TCDD	(2.5-254)	PNEC/benzo[a]pyrene	(2-368)			
<b>Oxidative stress</b>	0.000006	0.034	0.0007	0.7	4.3	<b>10</b>	
Nrf2 CALUX [ $\mu$ g CEQ/L]	NOEC/estradiol	(0.008-0.11)	EC50/retinoic acid	(0.2-2.2)			
<b>Pregnane X receptor</b>	0.000004	0.008	0.000004	0.007	1.5	<b>3</b>	
PRX CALUX [ $\mu$ g N1EQ/L]	LOEC/chlorpyrifos-ethyl	(0.002-0.024)	LOEC/chlorpyrifos-ethyl	(0.001-0.030)			
<b>Antibiotics activities (RIKILT WaterSCAN):</b>							
<b>- Aminoglycosides</b>	300	3322	300	3322	<LOD	<b>500</b>	
[ng N2EQ/L]	PNEC/neomycin	(1546-219614)	PNEC/neomycin	(1546-219614)			
<b>- Macrolides &amp; <math>\beta</math>-lactams</b>	1.8	98	2.22	2615	<LOD	<b>50</b>	
[ng PEQ/L]	EC50/flamulin	(13-470)	PNEC/amoxicillin	(96-23135)			
<b>- Sulphonamides</b>	10	67037	10	67037	4.6	<b>100</b>	
[ng SEQ/L]	LOEC/sulfadiazine	(24675-148222)	LOEC/sulfadiazine	(24675-148222)			
<b>- Tetracyclines</b>	170	27275	170	32931	<LOD	<b>250</b>	
[ng OEQ/L]	PNEC/oxytetracycline	(8292-68544)	PNEC/oxytetracycline	(9837-83368)			
<b>- Quinolones</b>	5.3	8759	5.3	8759	<LOD	<b>100</b>	
[ng F2EQ/L]	EC50/friclosan	(2197-26050)	EC50/friclosan	(2197-26050)			
	Grey:	Grey:	Grey:	Grey:			
	EBT > HC5 TEQ	EBT > HC5 TEQ	EBT > HC5 TEQ	EBT > HC5 TEQ			

\*: expressed as equivalents of the reference compounds: EEQ = estradiol; F1EQ = flutamide; TEQ = 2378-TCDD; DEQ = dexamethasone; REQ = rosiglitazone; BEQ = benzo[a]pyrene; CEQ = curcumin; N1EQ = nicardipine; N2EQ = neomycin; PEQ = penicillin; SEQ = sulfamethoxazole; OEQ = oxytetracycline; F2EQ = flumequine.  
<LOD = all below limit of detection

## 4 DISCUSSION

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Protection and restoration of chemical water quality is a primary goal to achieve, considering the fundamental ecological processes that take part in this environmental compartment. In order to improve the water quality by the most relevant measures, a more holistic and reliable monitoring of the water quality is needed. Many authors stressed out the necessity to improve the current monitoring programs using chemical analyses of selected target compounds, and highlight that bioanalytical tools can be applied to achieve this (Sanchez and Porcher, 2009; Allan et al., 2006; Cairns and van der Schalie, 1980; Connon et al., 2012; Poulsen et al., 2011; Greenwood et al., 2007). Several studies demonstrated the added value of bioassays to investigate the chemical quality of wastewater, surface water and drinking water (Kienle et al., 2011, Wolska et al., 2007, Macova et al., 2011, Zegura et al., 2009). Despite many advantages, bioanalytical tools are hardly used in regular monitoring programs, because there are no generally accepted methods for the interpretation of the data that are generated by effect measurements (biomarkers and bioassays). Hazard identification with bioanalytical tools could be applied as an initial screening to assess the microchemical water quality, provided that the effects could be classified. Effect-based trigger values that are able to discriminate between low risk and potential microchemical risks are needed for such a classification. The present study aims to be a first step towards the integration of bioassays in regulatory environmental processes. The primary objective of this study is to establish EBTs for a selection of inexpensive bioassays, targeting specific and reactive chemical activities in the water. The EBTs will be used to discriminate sites where chemical pollution poses a minor risk to aquatic community from sites where ecological risks may occur. Only the latter sites need to be further analyzed by more advanced chemical and toxicological analyses.

Routine monitoring with bioassays is still hampered by the lack of reliable interpretation guidelines. A challenge for scientists now is to provide effect-based trigger values (EBT) that allow regulators to link the test results to possible adverse effects on environmental or human health (Escher and Leusch, 2012). The fact that in many studies only a small percentage of the effect observed in a bioassay can be explained by known (chemically determined) substances makes it imperative to derive such EBT (Tang et al., 2013; Escher et al., 2013). A very limited amount of effect-based trigger values for water quality assessment can be found in literature.

Brand et al., (2013) derived some human health EBT for hormonal activity in drinking water. These EBT were derived to interpret *in vitro* CALUX bioassay results on estrogenic, androgenic, progestagenic and glucocorticoid activities. As a protocol to derive these EBT, the researchers used the reported ADI (acceptable daily intake), toxicokinetics information on bioavailability (protein binding), relative endocrine potencies and drinking water allocation factors of selected hormonal active compounds of interest.

Another approach for EBT derivation was followed by Tang et al. (2013) for an apical endpoint, the cytotoxicity measured by the bioluminescence inhibition in *Vibrio fischeri*. They proposed an algorithm for the derivation of effect-based water quality trigger values that was based on combined effects of mixtures of regulated chemicals, according to the concentration addition model. They used a QSAR approach to estimate the 50% effect concentrations (EC50) of the nonspecific mode of action of baseline toxicity. The effect-based water quality trigger value, EBT-EC50, was calculated with predicted mixture effect of all chemicals in a given guideline, divided by an extrapolation factor. The derived 'human health' EBT for drinking water was expressed as a relative enrichment factor (REF) of 3 to measure an EC50 (which is equal to 0.33 TU), which is less conservative than the 'environmental' EBT at REF 20 and 0.05 TU, proposed by Durand et al. (2009). The authors stated that the cytotoxicity based trigger value should not be used in isolation, but must be applied in conjunction with EBT targeting critical specific modes of action. Escher et al. (2013) used a similar strategy to derive an EBT for the oxidative stress response pathway with the AREC32 cell line. They derived an EBT that corresponded to a REF of 6 to measure an induction ratio of 1.5 (ECIR1.5), which means a 50% higher response than the blank control.

Hamers et al. (2010) obtained toxicity pathway profiles as toxicological 'fingerprints' for environmental samples, using

a bioassay battery with different modes of action. They used bioassays for genotoxicity, (anti)-estrogenic activity, thyroid activity, dioxin-like activity and nonspecific cell toxicity for hazard profiling and quality assessment of surface water sediments. Three potential approaches were described. In the first approach, toxicity profiles were translated into hazard profiles, indicating the relative distance to the desired or acceptable sediment quality status for each toxic mode of action. In the second approach, toxicity profiles were translated into ecological risk profiles indicating the ratio between the measured bioassay responses and the responses considered safe for environmental health. In the third approach, toxicity and hazard profiles were used to select samples with unusually high bioassay responses for further in-depth effect-directed analysis (EDA). A combination of the second and third of these approaches is most similar to the strategy that is proposed in the present paper (Figure 1). The main difference is that for our strategy a suite of effect-based trigger values are derived for 'low environmental risks' (confirmed at sites with good ecological status), instead of a limited number of known responses that indicate 'save for environmental health'.

A similar bioanalytical strategy as proposed in the present study for water quality monitoring, is already in use for determination of food quality in Europe. Bioassays are being used for high-throughput screening of large amounts of samples, and chemical analyses are only performed in samples where the bioassay responses indicate a potential risk. An EU working group derived a decision limit for bioanalyses of dioxins, on the basis of a GC/MS confirmation method and the condition that the chance of false negatives should be less than 5%. This bioanalytical procedure is now laid down in EU legislation (European Union, 2012). For other groups of substances (hormones, antibiotics), there are also established methods to regularly apply bioassays in food quality control (e.g., Bovee et al., 2006; Gizzi et al., 2005).

The *in vitro* bioassay panel that is proposed for hazard identification (Table 1), together with bioassays for non-specific toxicity, should be able to distinguish microchemical quality of sites for low or potential risks for adverse ecological health effects. The authors realize that it is 'scientifically impossible' to derive solid and realistic trigger values for bioassays that distinguish between 'good' and 'bad' chemical status, if the identity of compounds that cause the bioassay responses is unknown. Since the identity of the compounds that cause a bioassay effect is unknown, both over- and underestimations of the overall toxic impact of the mixture can be made with the TEQ concept. Moreover, it is hard to predict adverse *in vivo* effects with *in vitro* responses (Escher and Leusch, 2012). Nevertheless, an approach is suggested to make an initial screening for potential environmental risks with *in vivo* as well as *in vitro* bioassay responses. It is again emphasized that we do not claim to make a sharp division between good and bad, but we suggest a toxicity screening to distinguish between low and potential microchemical risks for the environment. Potential risks have to be verified in a second phase risk assessment with a combination of advanced chemical and toxicological tools (Figure 1). The discriminative power of the first phase should be able to indicate potential hazards, but not all of the investigated sites should be classified as being a potential risk. This means that the precautionary principle and the use of high safety factors are not consequently adapted. Instead, this approach tries applied low safety factors to derive safe TEQ. In addition, the design of low risk EBT is based on more realistic environmental estimations, comparison with a species sensitivity distribution and a benchmark with known chemical and biological data (e.g., good ecological status of sites). The trigger values that were developed with this approach are listed in Table 6. The EBTs proposed in this report are most probably not the definite values, since they could be subject to refinement when additional information becomes available.

Jarosova et al. (2014) derived safe environmental concentrations of estrogenic equivalents (EEQ-SSE) in municipal wwtp effluents, based on simplified assumption that only steroid estrogens are responsible for *in vitro* estrogenicity. EEQ-SSE were derived using the estrogenic REP in bioassays, the *in vivo* PNECs of the compounds, and their relative contributions to the overall estrogenicity of wwtp effluents. The EEQ-SSEs for ER-CALUX varied from 0.2 to 0.4 ng EEQ/L for long-term exposure and from 0.6 to 2.0 for short-term exposure. Kunz et al. (2015) proposed to use the annual average environmental quality standard (AA-EQS) of 17-estradiol (E2) of 0.4 ng/L as a trigger value for overall estrogenic activity, among others because using the 0.035 ng/L AA-EQS of ethinyl-estradiol, would overestimate the risks in most cases. The EBT of 0.5 ng EEQ/L we



proposed for ERa CALUX was close to the trigger values proposed by Jarosova and Kunz. Johnson et al. (2007) suggested a PNEC for combined estrogenic activity of 1 ng EEQ/L, but this was based upon an outdated PNEC value for estradiol.

Of course there are several limitations and assumptions connected to the present study that need to be considered and validated in future research. Four assumptions had to be made in the EBT design, regarding i) ratio of 10 used to convert acute to chronic toxicity, ii) estimated duration of chronic toxicity studies, iii) assessment factors to convert concentrations of toxicity endpoints (LOEC, EC50 and LC50) to safe values, and iv) the selection of compounds was restricted to their relative effect potencies (REP) in the bioassay. There are also limitations regarding toxicological data from which the EBTs were derived. Only chemicals with a known REP in the bioassay could be considered due to the toxic equivalents (TEQ) approach. Reliable REP values are essential for a good conversion of substance concentrations to TEQ, but it was not always possible to assess REPs with EC50 data due to cytotoxicity in the bioassay at higher concentrations. The CALUX REPs that were estimated with EC10 values may not be 100% accurate if the slopes of the dose-response curves differ from that of the reference compound.

For several compounds that met the REP requirements it was hard or impossible to find aquatic toxicity data. This resulted in a limited dataset for some of the bioassays considered in the present study (e.g. DR CALUX and GR CALUX). Toxicity studies with antibiotics are mainly focused on invertebrate species, although some data are available for fish. No data, however, could be found for reptiles or mammals and only one study reported acute effects of antibiotics on an amphibian species. Since the EBT values will be used to assess the water quality, all toxicity data had to be water concentrations that indicate a certain effect (PNEC, NOEC, LOEC, EC50 or LC50) to aquatic organisms. Laboratory exposures investigate the toxic effects of single compounds to single species under controlled conditions. Toxic effects of substances in the natural environment can be enhanced or decreased by different physico-chemical conditions, such as pH, temperature and light exposure. This can lead to an under- or over-estimation of the risk connected to the concentrations of chemicals in the environment. In addition, the majority of the studies investigating the toxicity of highly hydrophobic compounds only reported nominal concentrations, as a result of which the toxicity can be underestimated. Therefore, future research should focus the actual concentrations of these compounds causing the reported negative effects.

Despite its limitations and uncertainties, the approach proposed in this study constitutes a better alternative to the current EU WFD monitoring that chemically analyses a limited amount of priority compounds in grab samples. Grab samples are only snapshots of the varying water contamination that does not consider the bioavailability of the compounds, while passive sampling assesses a time-weight average concentration of bioavailable compounds. The highest uncertainty, however, is to analyze only 45 priority compounds in order to assess the overall risk of micropollutants in the water, while the potential impact of more than 100,000 unknown compounds remains unknown. Therefore, reliable alternatives are needed to provide a more realistic hazard identification and risk assessment of the chemical pollution of surface waters. The strategy proposed in this paper represents one of the first attempts to connect bioassays responses with potential negative effects for aquatic population. This hazard assessment approach, combined with chemical analysis, is more reliable and realistic than the current monitoring conducted with chemical analyses only.

Nowadays, it is fundamental to propose better monitoring strategies in order to preserve the quality of freshwater environment, since availability of water supplies and his capacity to sustain human life and economy as we know it is becoming an issue due to the constant increase of human population. As pointed out by Stikker (1998) “the availability of clean water is one of the basic conditions for achieving sustainable development in the 21st century. Sustainable development implies that future generations should have the same opportunities of enjoying a decent quality of life as does the present generation” since “where there is no water, there is no food, no consumer and no business activity”. The introduction of bioassays can lead to a more sustainable water quality monitoring, as they meet the sustainability criteria better than the classical strategies (Gagnon et al., 2007).

Further refinement of EBT derivation should be based upon expert judgment and future validation studies. Together, the two types of triggers (safe TEQ and low risk EBT) should make a distinction between sites with negligible microchemical risks, low risks or potential risks for the ecosystem health. EAWAG Switzerland uses a similar approach with 3 categories: i) good (<safe TEQ), ii) in range of quality criterion (<low risk EBT) or iii) poor (>EBT) (Cornelia Kienle [EAWAG], personal communication). The outcome of the SIMONI model indicates which sites are hot-spots, relevant for additional chemical-toxicological research. Moreover, if specific or reactive activities are above EBT, the model will indicate which class of chemicals may cause the main problem for aquatic organisms. If the information from the SIMONI hazard assessment is combined with the available knowledge of various aspects of the water system (such as influences of other ecological key factors), a tailor-made plan can be designed for further ecological risk assessment. This assessment should be able to identify the main causes of an impaired ecological status, so that the most cost-effective and efficient measures can be taken to improve the water quality. The next steps to make the proposed concept attractive for risk assessors would be to gain experience upon the applicability to case studies and to evaluate its robustness for practical use. Field validation studies with this strategy will be described in the second paper of the “SIMONI as a novel bioanalytical strategy for water quality assessment” series (Van der Oost et al., in progress). Due to its low costs and high relevance, this SIMONI model has the potential to become the first bioanalytical strategy to be applied in regular monitoring of surface water quality. Most Dutch water authorities will start feasibility studies with the SIMONI strategy in 2016.



## 5 CONCLUSIONS

This study aimed to derive EBTs for a selection of bioassays (see Table 7). These trigger values were derived by an extensive review of the available literature on aquatic toxicity of compounds that had a significant response in the bioassays, an SSD analysis on TEQ values and benchmarking with environmental field investigations conducted by Waternet. In only a small percentage of the field samples activities were found above the proposed low-risk effect-based trigger values. However, since these draft EBTs have been derived with some assumptions and simplifications, they may be adjusted when new information becomes available. Nevertheless, if the primary aim of a monitoring program is to assess the chemical water quality, bioassays will be powerful holistic tools to detect situations at risk where additional risk assessment is needed. This project may provide a new standard for the interpretation of bioassay data, and implementation in regular monitoring programs. The alternative monitoring strategy presented in this study has the potential to reduce monitoring costs if advanced chemical analyses only have to be applied at sites where EBTs are exceeded. In addition, costs on expensive measures to improve the water quality can be reduced since the chemical risks can be better assessed with the SIMONI strategy.

**TABLE 7**

*Summary of all safe and low-risk EBTs for the initial selection of in vitro bioassays.*

Activity	Units	Safe EBT	Low risk EBT
Estrogenic	ng EEQ/L	0.0066	0.5
Anti-androgenic	µg F1EQ/L	0.00005	25
Dioxin and dioxin-like	pg TEQ/L	0.4	50
Glucocorticoid	ng DEQ/L	20	100
PPAR $\gamma$ receptor	ng REQ/L	0.00014	10
Toxic PAHs	ng BEQ/L	0.04	150
Oxidative stress	µg CEQ/L	0.000006	10
Pregnane X receptor	µg N1EQ/L	0.000004	3
Aminoglycosides	ng N2EQ/L	300	500
Macrolides & $\beta$ -Lactam	ng PEQ/L	1.8	50
Antibiotics: Sulphonamides	ng SEQ/L	10	100
Tetracyclines	ng OEQ/L	170	250
Quinolones	ng F2EQ/L	5.3	100

## 6 REFERENCES

- Adams W.J., DeGraeve G.M., Sabourin T.D., Cooney J.D., Mosher G.M. (1986). Toxicity and bioconcentration of 2,3,7,8-TCDD to fathead minnows (*Pimephales promelas*). *Chemosphere* 15(9-12):1503-1511.
- Agra A.R. and Soares A.M. (2009). Effects of two insecticides on survival, growth and emergence of *Chironomus riparius meigen*. *Bull. Environ. Contam. Toxicol.* 82(4):501-504.
- Alam M.N., Lakshmi V., Mishra S.K. (2010). Toxic effects of some pesticides on fingerlings of a carp, *Labeo rohita*. *Nat. Environ. Pollut. Technol.* 9(3):633-637.
- Albertsson E., Kling P., Gunnarsson L., Larsson D.G.J., Forlin L. (2007). Proteomic analyses indicate induction of hepatic carbonyl reductase/20beta-hydroxysteroid dehydrogenase b in rainbow trout exposed to sewage effluent. *Ecotoxicol. Environ. Saf.* 68(1):33-39.
- Ali D., Nagpure N.S., Kumar S., Kumar R., Kushwaha B., Lakra W.S. (2009). Assessment of genotoxic and mutagenic effects of chlorpyrifos in freshwater fish *Channa punctatus* (bloch) using micronucleus assay and alkaline single-cell gel electrophoresis. *Food Chem. Toxicol.* 47(3):650-656.
- Aliotta G., Pinto G., Pollio A. (1983). Observations on tolerance to heavy metals of four green algae in relation to pH. *G. Bot. Ital.* 117(5/6):247-251.
- Allan I.J., Vrana B., Greenwood R., Mills G.A., Roig B., Gonzalez C. (2006) A "toolbox" for biological and chemical monitoring requirements for the European Union's Water Framework Directive. *Talanta* 69:302-322.
- Allen Y.T., Katsiadaki I., Pottinger T.G., Jolly C., Matthiessen P., Mayer I., Smith A., Scott A.P., Eccles P., San M.B. (2008). Intercalibration exercise using a stickleback endocrine disrupter screening assay. *Environ. Toxicol. Chem.* 27(2):404-412.
- Alonso A., De Lange H.J., Peeters E.T.H.M. (2009). Development of a feeding behavioural bioassay using the freshwater amphipod *Gammarus pulex* and the multispecies freshwater biomonitor. *Chemosphere* 75:341-346.
- Altenburger R., Ait-Aissa S., Antczak P., Backhaus T., Barceló D., Seiler T.B., Brion F., Busch W., Chipman K., de Alda M.L., de Aragão Umbuzeiro G., Escher B.I., Falciani F., Faust M., Focks A., Hilscherova K., Hollender J., Hollert H., Jäger F., Jahnke A., Kortenkamp A., Krauss M., Lemkine G.F., Munthe J., Neumann S., Schymanski E.L., Scrimshaw M., Segner H., Slobodnik J., Smedes F., Kughathas S., Teodorovic I., Tindall A.J., Tollefsen K.E., Walz K.H., Williams T.D., Van den Brink P.J., van Gils J., Vrana B., Zhang X., Brack W. (2015). Future water quality monitoring - Adapting tools to deal with mixtures of pollutants in water resource management. *Science of The Total Environment* 512-513: 540-551.
- Altenburger R., Walter H., Grote M. (2004). What Contributes to the Combined Effect of a Complex Mixture?. *Environ. Sci. Technol.* 38(23):6353-6362.
- Amanullah B., Stalin A., Prabu P., Dhanapal S. (2010). Analysis of AchE and LDH in mollusc, *Lamellidens marginalis* after exposure to chlorpyrifos. *J. Environ. Biol.* 31(4):417-419.
- Andersen H.R., Wollenberger L., Halling-Sorensen B., Kusk K.O. (2001). Development of copepod Nauplii to copepodites - A parameter for chronic toxicity including endocrine disruption. *Environ. Toxicol. Chem.* 20(12):2821-2829.
- Andersen L., Bjerregaard P., Korsgaard B. (2003). Vitellogenin induction and brain aromatase activity in adult male and female zebrafish exposed to endocrine disrupters. *Fish Physiol. Biochem.* 28(1-4):319-321.
- Anderson P.D., Johnson A.C., Pfeiffer D., Caldwell D.J., Hannah R., Mastrocco F., Sumpter J.P., William R.J. (2012). Endocrine disruption due to estrogens derived from human predicted to be low in the majority of U.S. surface waters. *Environmental Toxicology and Chemistry* 31:1407-1415.
- Andrieu M., Rico A., Phu T.M., Huong D.T.T., Phuong N.T., Van den Brink P.J., (2015). Ecological risk assessment of the antibiotic enrofloxacin applied to *Pangasius catfish* farms in the Mekong Delta, Vietnam. *Chemosphere* 119C:407-414.
- Ankley G.T., Jensen K.M., Kahl M.D., Durhan E.J., Makynen E.A., Cavallin J.E., Martinovic D., Wehmas L.C., Mueller N.D., Villeneuve D.L. (2010). Use of chemical mixtures to differentiate mechanisms of endocrine action in a small fish model. *Aquat. Toxicol.* 99(3): 389-396.
- Annabi A., Messaoudi I., Kerkeni A., Said K. (2009). Comparative study of the sensitivity to cadmium of two populations of *Gambusia affinis* from two different sites. *Environ. Monit. Assess.* 155(1-4):459-465.
- Anton F.A. (1993). Acute toxicity of technical captan to algae and fish. *Bull. Environ. Contam. Toxicol.* 50(3):392-399.

- Antunes S.C., Pereira J.L., Cachada A., Duarte A.C., Goncalves F., Sousa J.P., Pereira R. (2010). Structural effects of the bioavailable fraction of pesticides in soil: suitability of elutriate testing. *J. Hazard. Mater.* 184(1-3):215-225.
- Ashauer R., Boxall A.B.A., Brown C.D. (2007). New ecotoxicological model to simulate survival of aquatic invertebrates after exposure to fluctuating and sequential pulses of pesticides. *Environ. Sci. Technol.* 41(4):1480-1486.
- Atienzar F.A., Conradi M., Evenden A.J., Jha A.N., Depledge M.H. (1999). Qualitative assessment of genotoxicity using random amplified polymorphic DNA: comparison of genomic template stability with key fitness parameters in *Daphnia magna* exposed to benzo(a)pyrene. *Environ. Toxicol. Chem.* 18(10):2275-2282.
- Bachmann J. (2002). Entwicklung und erprobung eines teratogenitäts-screening tests mit embryonen des zebrabärblings *Danio rerio*, 249 S. PhD thesis, University of Dresden, Germany.
- Backhaus T., Grimme L.H. (1999). The toxicity of antibiotic agents to the luminescent bacterium *Vibrio fischeri*. *Chemosphere* 38:3291-3301.
- Baer K.N. and Owens K.D. (1999). Evaluation of selected endocrine disrupting compounds on sex determination in *Daphnia magna* using reduced photoperiod and different feeding rates. *Bull. Environ. Contam. Toxicol.* 62(2):214-221.
- Baldisserotto B., Chowdhury M.J., Wood C.M. (2005). Effects of dietary calcium and cadmium on cadmium accumulation, calcium and cadmium uptake from the water, and their interactions in juvenile rainbow trout. *Aquat. Toxicol.* 72(1/2):99-117.
- Baldwin I.G., Harman M.M.I., Neville D.A. (1994). Performance characteristics of a fish monitor for detection of toxic substances-i. laboratory trials. *Water Res.* 28(10):2191-2199.
- Baldwin W.S., Milam D.L., LeBlanc G.A. (1995). Physiological and biochemical perturbations in *Daphnia magna* following exposure to the model environmental estrogen diethylstilbestrol. *Environ. Toxicol. Chem.* 14(6):945-952.
- Bao M.L., Dai S.G., Pantani F. (1997). Effect of dissolved humic material on the toxicity of tributyltin chloride and triphenyltin chloride to *Daphnia magna*. *Bull. Environ. Contam. Toxicol.* 59(4):671-676.
- Baran W., Sochacka J., Wardas W. (2006). Toxicity and biodegradability of sulfonamides and products of their photocatalytic degradation in aqueous solutions. *Chemosphere* 65:1295-9.
- Becker M.C. (1991). Toxicity of polychlorinated diphenyl ethers in *Hydra attenuata* and in rat whole embryo culture. Ph.D. Thesis, Texas A&M University, College Station TX:87p.
- Beehler J.W., Quick T.C., DeFoliart G.R. (1991). Residual toxicity of four insecticides to *Aedes triseriatus* in scrap tires. *J. Am. Mosq. Control Assoc.* 7(1):121-122.
- Belden J.B. and Lydy M.J. (2006). Joint toxicity of chlorpyrifos and esfenvalerate to fathead minnows and midge larvae. *Environ. Toxicol. Chem.* 25(2):623-629.
- Belgers J.D.M., Aalderink G.H., Van den Brink P.J. (2009). Effects of four fungicides on nine non-target submersed macrophytes. *Ecotoxicol. Environ. Saf.* 72(2):579-584.
- Bertilsson G, Heidrich J, Svensson K, Asman M, Jendeberg L, Sydow-Bäckman M, Ohlsson R, Postlind H, Blomquist P, Berkenstam A (October 1998). Identification of a human nuclear receptor defines a new signaling pathway for CYP3A induction. *Proc. Natl. Acad. Sci. U.S.A.* 95 (21): 12208-13.
- Bhilave M.P., Muley D.V., Deshpande V.Y. (2008). Biochemical changes in the fish *Cirrhinus mrigala* after acute and chronic exposure of heavy metals. *Nat. Environ. Pollut. Technol.* 7(1):65-71.
- Biales A.D., Bencic D.C., Lazorchak J.L., Lattier D.L. (2007). A quantitative real-time polymerase chain reaction method for the analysis of vitellogenin transcripts in model and nonmodel fish species. *Environ. Toxicol. Chem.* 26(12):2679-2686.
- Bjerregaard P., Hansen P.R., Larsen K.J., Erratico C., Korsgaard B., Holbech H. (2008). Vitellogenin as a biomarker for estrogenic effects in brown trout, *Salmo trutta*: laboratory and field investigations. *Environ. Toxicol. Chem.* 27(11):2387-2396.
- Bjorkblom C., Hogfors E., Salste L., Bergelin E., Olsson P.E., Katsiadaki I., Wiklund T. (2009). Estrogenic and androgenic effects of municipal wastewater effluent on reproductive endpoint biomarkers in three-spined stickleback (*Gasterosteus aculeatus*). *Environ. Toxicol. Chem.* 28(5):1063-1071.
- Borgmann U., Chau Y.K., Wong P.T.S., Brown M., Yaromich J. (1996). The relationship between tributyltin (TBT) accumulation and toxicity to *Hyalella azteca* for use in identifying TBT toxicity in the field. *J. Aquat. Ecosyst. Health* 5(3):199-206.
- Braathen M., Mdegela R.H., Correia D., Rundberget T., Myburgh J., Botha C., Skaare J.U., Sandvik M. (2009). Vitellogenin in african sharptooth catfish (*Clarias gariepinus*): purification, characterization, and elisa development. *J. Toxicol. Environ. Health Part*

A 72(3):173-183.

- Brain R., Johnson D., Richards S., Sanderson H., Sibley P., Solomon K. (2004). Effects of 25 pharmaceutical compounds to *lemna gibba* using a seven-day static-renewal test. *Environ. Toxicol. Chem.* 23:371-382.
- Brand W., de Jongh C.M., van der Linden S.C., Mennes W., Puijker L.M., van Leeuwen C.J., van Wezel A.P., Schriks M., Heringa M.B. (2013). Trigger values for investigation of hormonal activity in drinking water and its sources using CALUX bioassays. *Environment International* 55:109-118
- Brande-Lavridsen N., Christensen-Dalsgaard J., Korsgaard B. (2010). Effects of ethinylestradiol and the fungicide prochloraz on metamorphosis and thyroid gland morphology in *Rana temporaria*. *Open Zool. J.* 3:7-16.
- Breitholtz M. and Bengtsson B.E. (2001). Oestrogens have no hormonal effect on the development and reproduction of the harpacticoid copepod *Nitocra spinipes*. *Mar. Pollut. Bull.* 42(10):879-886.
- Brennan S.J., Brougham C.A., Roche J.J., Fogarty A.M. (2006). Multi-generational effects of four selected environmental oestrogens on *Daphnia magna*. *Chemosphere* 64(1):49-55.
- Brian J.V., Harris C.A., Scholze M., Backhaus T., Booy P., Lamoree M., Pojana G., Jonkers N., Runnalls T., Bonfa A., Marcomini A., Sumpter J.P. (2005). Accurate prediction of the response of freshwater fish to a mixture of estrogenic chemicals. *Environ. Health Perspect.* 113(6):721-728.
- Bringolf R.B., Belden J.B., Summerfelt R.C. (2004). Effects of atrazine on fathead minnow in a short-term reproduction assay. *Environ. Toxicol. Chem.* 23(4):1019-1025.
- Brodeur J.C., Sassone A., Hermida G.N., Codugnello N. (2013). Environmentally-relevant concentrations of atrazine induce non-monotonic acceleration of developmental rate and increased size at metamorphosis in *Rhinella arenarum* tadpoles. *Ecotoxicol. Environ. Saf.* 92:10-17
- Brodeur J.C., Woodburn K.B., Klecka G.M. (2005). Potentiation of the vitellogenic response to 17 $\alpha$ -ethinylestradiol by cortisol in the fathead minnow *Pimephales promelas*. *Environmental Toxicology and Chemistry* 24(5):1125-1132.
- Brooke L.T. (1993). Acute and chronic toxicity of fluoranthene, with and without additional ultraviolet light, to twelve species of freshwater organisms. U.S.EPA Contract No.68-C1-0034, to R.Spehar, U.S.EPA, Duluth, MN:51 p.
- Brooke L.T. (1995). Acute toxicity of chlorpyrifos to the cladocerans *Ceriodaphnia dubia* and *Daphnia magna*. U. S. EPA Contract 68-C1-0034, Lake Superior Research Institute, Superior, WI:9 p.
- Brown K.H., Schultz I.R., Cloud J.G., Nagler J.J. (2008). Aneuploid sperm formation in rainbow trout exposed to the environmental estrogen 17 $\alpha$ -ethinylestradiol. *Proc. Natl. Acad. Sci. U.S.A.* 105(50):19786-19791.
- Brown K.H., Schultz I.R., Nagler J.J. (2007). Reduced embryonic survival in rainbow trout resulting from paternal exposure to the environmental estrogen 17 $\alpha$ -ethinylestradiol during late sexual maturation. *Reproduction (Camb.)* 134(5):659-666.
- Bruggemann R., Schwaiger J., Negele R.D. (1995). Applying hasse diagram technique for the evaluation of toxicological fish tests. *Chemosphere* 30(9):1767-1780.
- Cacciatore L.C., Guerrero N.V., Cochon A.C. (2013). Cholinesterase and carboxylesterase inhibition in *Planorbarius corneus* exposed to binary mixtures of azinphos-methyl and chlorpyrifos. *Aquat. Toxicol.* 128/129:124-134.
- Cairns jr J. and van der Schalie W.H. (1980). Biological monitoring Part I -- Early warning systems. *Water Research* 14(9):1179-1196.
- Caldwell D.J., Mastrocco F., Anderson P.D., Lange R., Sumpter J.P. (2012). Predicted-no-effect concentrations for the steroid estrogens estrone, 17 $\beta$ -estradiol, estriol, and 17 $\alpha$ -ethinyl estradiol. *Environ Toxicol Chem* 31:1396-1406.
- Call D.J., Brooke L.T., Hammermeister D.H., Northcott C.E., Hoffman A.D. (1983). Variation of acute toxicity with water source. Center for Lake Superior Environmental Studies, Report No. LSRI0 273:58 p.
- Canton J.H. (1976). The toxicity of benomyl, thiophanate-methyl, and BCM to four freshwater organisms. *Bull. Environ. Contam. Toxicol.* 16(2):214-218.
- Capleton, A.C., Courage, C., Rumsby, P., Holmes, P., Stutt, E., Boxall, A.B. a, Levy, L.S., 2006. Prioritising veterinary medicines according to their potential indirect human exposure and toxicity profile. *Toxicol. Lett.* 163:213-23.
- Carder J.P. and Hoagland K.D. (1998). Combined effects of alachlor and atrazine on benthic algal communities in artificial streams. *Environ. Toxicol. Chem.* 17(7):1415-1420.
- Carlson A.R. and Kosian P.A. (1987). Toxicity of chlorinated benzenes to fathead minnows (*Pimephales promelas*). *Arch. Environ. Contam. Toxicol.* 16(2):129-135.

- Carlsson, G., Örn, S., Joakim Larsson, D., 2009. Pharmaceuticals and Personal Care Products in the Environment – effluent from bulk drug production is toxic to aquatic vertebrates. *Environ. Toxicol. Chem.* 28, 2656–2662.
- Carr J.A., Gentles A., Smith E.E., Goleman W.L., Urquidi L.J., Thuett K., Kendall R.J., Giesy J.P., Gross T.S., Solomon K.R., van Der Kraak G. (2003). Response of larval *Xenopus laevis* to atrazine: assessment of growth, metamorphosis, and gonadal and laryngeal morphology. *Environ. Toxicol. Chem.* 22(2):396-405.
- Carraschi S.P., Shiogiri N.S., Venturini F.P., Cruz C., Gírio A.C.F., Machado Neto J.G. (2011). acute toxicity and environmental risk of oxytetracycline and florfenicol antibiotics to pacu (*Piaractus mesopotamicus*). *Bol. Inst. Pesca* 37:115–122.
- Carvalho G.G.A., De Franca J.G., Dias D.C., Lombardi J.V., De Paiva M.J.R., Carvalho S., Sarries G.A., Ferreira J.R. (2009). Selenite and selenate effects on mercury (Hg<sup>2+</sup>) uptake and distribution in tilapia, *Oreochromis niloticus* L., assessed by chronic bioassay. *Bull. Environ. Contam. Toxicol.* 82(3):300-304.
- Chadwick Ecological Consultants Inc. (2003). Acute and Chronic toxicity of cadmium to freshwater crustaceans at different water hardness concentrations. Rep.Prepared for Thompson Creek Mining Company, Challis, ID, by Chadwick Ecological Consultants, Inc., Littleton, CO:51p.
- Chandre F., Darriet F., Doannio J.M.C., Riviere F., Pasteur N., Guillet P. (1997). Distribution of organophosphate and carbamate resistance in *Culex pipiens quinquefasciatus* (Diptera: Culicidae) in west Africa. *J. Med. Entomol.* 34(6):664-671.
- Chang K.S., Jung J.S., Park C., Lee D.K., Shin E.H. (2009). Insecticide susceptibility and resistance of larvae of the *Anopheles sinensis* group (Diptera: Culicidae) from Paju, Republic of Korea. *Entomol. Res.* 39(3):196-200.
- Chang L.W., Toth G.P., Gordon D.A., Graham D.W., Meier J.R., Knapp C.W., DeNoyelles Jr.F.J., Campbell S., Latt D.L. (2005). Responses of molecular indicators of exposure in mesocosms: common carp (*Cyprinus carpio*) exposed to the herbicides alachlor and atrazine. *Environ. Toxicol. Chem.* 24(1):190-197.
- Chapman H.F., Leusch F.D.L., Prochazka E., Cumming J., Ross V., Humpage A., Froscio S., Laingam S., Khan S.J., Trinh T., McDonald J.A. (2011). A national approach to health risk assessment, risk communication and management of chemical hazards from recycled water. *Waterlines report* 48.
- Chen C.M. and Coopoe K.R. (1999). Developmental toxicity and EROD induction in the japanese medaka (*Oryzias latipes*) treated with dioxin congeners. *Bull. Environ. Contam. Toxicol.* 63(4):423-429.
- Cheng S.S., Chua M.T., Chang E.H., Huang C.G., Chen W.J., Chang S.T. (2009b). Variations in insecticidal activity and chemical compositions of leaf essential oils from *Cryptomeria japonica* at different ages. *Bioresour. Technol.* 100:465-470.
- Cheng S.S., Huang C.G., Chen Y.J., Yu J.J., Chen W.J., Chang S.T. (2009a). Chemical compositions and larvicidal activities of leaf essential oils from two eucalyptus species. *Bioresour. Technol.* 100:452-456.
- Cho E.A.. (2005). Bioturbation as a novel method to characterize the toxicity of aquatic sediment. Ph.D. Thesis, North Carolina State University, NC:153p.
- Choi J. and Ha M.H. (2009). Effect of cadmium exposure on the globin protein expression in 4th instar larvae of *Chironomus riparius* mg. (Diptera: Chironomidae): an ecotoxicoproteomics approach. *Proteomics (Weinh.)* 9(1):31-39.
- Chourpagar A.R. and Kulkarni G.K. (2011). Heavy Metal toxicity to a freshwater crab, *Barytelphusa cunicularis* (Westwood) from aurangabad region. *Recent Res. Sci. Technol.* 3(3):1-5.
- Clark B.W. (2010). Molecular mechanisms underlying adaptation to PAHs in *Fundulus heteroclitus*. Ph.D. Thesis, Duke University, Raleigh, NC:230p.
- Clark B.W. and Di Giulio R.T. (2012). *Fundulus heteroclitus* adapted to PAHs are cross-resistant to multiple insecticides. *Ecotoxicology* 21(2):465-474.
- Clark B.W., Matson C.W., Jung D., Di Giulio R.T. (2010). AHR2 mediates cardiac teratogenesis of polycyclic aromatic hydrocarbons and PCB-126 in atlantic killifish (*Fundulus heteroclitus*). *Aquat. Toxicol.* 99(2):232-240.
- Clayton M.E., Steinmann R., Fent K. (2000). Different expression patterns of heat shock proteins hsp 60 and hsp 70 in zebra mussels (*Dreissena polymorpha*) exposed to copper and tributyltin. *Aquat. Toxicol.* 47(3-4):213-226.
- Cline P.V., Denslow N., Meyer P., Goudey S., Lewellen A., Johnson I. (2003). Evaluating endocrine disruption in receiving waters: screening for biomarkers. Final Report. Water Environment Research Foundation, Alexandria, VA:284p.
- Clubbs R.L. and Brooks B.W. (2007). *Daphnia magna* responses to a vertebrate estrogen receptor agonist and an antagonist: a multi-generational study. *Ecotoxicol. Environ. Saf.* 67(3):385-398.

- Coady K.K., Murphy M.B., Villeneuve D.L., Hecker M., Jones P.D., Carr J.A., Solomon K.R., Smith E.E., Van der Kraak G. (2004). Effects of atrazine on metamorphosis, growth, and gonadal development in the green frog (*Rana clamitans*). *J. Toxicol. Environ. Health Part A* 67(12):941-957.
- Coe T.S., Hamilton P.B., Hodgson D., Paull G.C., Tyler C.R. (2009). Parentage outcomes in response to estrogen exposure are modified by social grouping in zebrafish. *Environ. Sci. Technol.* 43(21):8400-8405.
- Collier A., Orr L., Morris J., Blank J. (2008). The effects of 2,3,7,8 tetrachlorodibenzo-p-dioxin (TCDD) on the mortality and growth of two amphibian species (*Xenopus laevis* and *Pseudacris triseriata*). *Int. J. Environ. Res. Public Health* 5(5):368-377.
- Colman J.R., Baldwin D., Johnson L.L., Scholz N.L. (2009). Effects of the synthetic estrogen, 17-ethinylestradiol, on aggression and courtship behavior in male zebrafish (*Danio rerio*). *Aquat. Toxicol.* 91:346-354.
- Cong L., Qin Z.F., Jing X.N., Yang L., Zhou J.M., Xu X.B. (2006). *Xenopus laevis* is a potential alternative model animal species to study reproductive toxicity of phytoestrogens. *Aquat. Toxicol.* 77(3):250-256.
- Connon R.E., Geist J., Werner I. (2012). Effect-based tools for monitoring and predicting the ecotoxicological effects of chemicals in the aquatic environment. *Sensors* 12:12741-12771.
- Cope W.G., Bartsch M.R., Marking L.L. (1997). Efficacy of candidate chemicals for preventing attachment of zebra mussels (*Dreissena polymorpha*). *Environ. Toxicol. Chem.* 16(9):1930-1934
- Crump D., Lean D., Trudeau V.L. (2002). Octylphenol and UV-B radiation alter larval development and hypothalamic gene expression in the leopard frog (*Rana pipiens*). *Environ. Health Perspect.* (3):277-284.
- Daam M.A. and Van den Brink P.J. (2007). Effects of chlorpyrifos, carbendazim, and linuron on the ecology of a small indoor aquatic microcosm. *Arch. Environ. Contam. Toxicol.* 53(1):22-35.
- De Coen W.M., Vangheluwe M.L., Janssen C.R. (1998). The use of biomarkers in *Daphnia magna* toxicity testing. III. Rapid toxicity testing of pure chemicals and sediment pore waters using ingestion and digestive enzyme activity. *Chemosphere* 37(13):2677-2694.
- De la Broise, D., and S. Stachowski-Haberhorn. 2012. Evaluation of the Partial Renewal of In Situ Phytoplankton Microcosms and Application to the Impact Assessment of Bentazon and Dimethenamid. *Mar. Pollut. Bull.* 64(11): 2480-2488
- De Liguoro, M., Fioretto, B., Poltronieri, C., Gallina, G., 2009. The toxicity of sulfamethazine to *Daphnia magna* and its additivity to other veterinary sulfonamides and trimethoprim. *Chemosphere* 75:1519–1524.
- De Vries H., Penninks A.H., Snoeij N.J., Seinen W. (1991). Comparative toxicity of organotin compounds to rainbow trout (*Oncorhynchus mykiss*) Yolk Sac Fry. *Sci. Total Environ.* 103(2-3):229-243.
- De Zwart D. (1995). Monitoring water quality in the future. Volume 3: biomonitoring. RIVM. Bilthoven, The Netherlands
- Deanovic L.A., Markiewicz D., Stillway M., Fong S., Werner I. (2013). Comparing the effectiveness of chronic water column tests with the crustaceans *Hyalella azteca* (Order: Amphipoda) and *Ceriodaphnia dubia* (Order: Cladocera) in detecting toxicity of current-use insecticides. *Environ. Toxicol. Chem.* 32:707-712.
- Deleebeeck N.M.E., De Schampelaere K.A.C., Janssen C.R. (2009). Effects of Mg<sup>2+</sup> and H<sup>+</sup> on the toxicity of Ni<sup>2+</sup> to the unicellular green alga *Pseudokirchneriella subcapitata*: model development and validation with surface waters. *Sci. Total Environ.* 407(6):1901-1914.
- DellaGreca M., Fiorentino A., Isidori M., Lavorgna M., Previtiera L., Rubino M., Temussi F. (2004). Toxicity of prednisolone, dexamethasone and their photochemical derivatives on aquatic organisms. *Chemosphere* 54(5):629–637.
- Delorenzo M.E., Keller J.M., Arthur C.D., Finnegan M.C., Harper H.E., Winder V.L., Zdankiewicz D.L. (2008). Toxicity of the antimicrobial compound triclosan and formation of the metabolite methyl-triclosan in estuarine systems. *Environ Toxicol.* 23(2):224-32.
- Demetrio P.M., Rossini G.D.B., Bonetto C.A., Ronco A.E. (2012). Effects of pesticide formulations and active ingredients on the coelenterate *Hydra attenuata* (Pallas, 1766). *Bull. Environ. Contam. Toxicol.* 88(1):15-19.
- Desouky M.M.A. (2012). Metallothionein is up-regulated in molluscan responses to cadmium, but not aluminum, exposure. *J. Basic Appl. Zool.* 65:139-143.
- Devlin E.W. (2006). Acute toxicity, uptake and histopathology of aqueous methyl mercury to Fathead Minnow embryos. *Ecotoxicology* 15:97-110.
- Dietrich S., Ploessl F., Bracher F., Laforsch C. (2010). Single and combined toxicity of pharmaceuticals at environmentally relevant concentrations in *Daphnia magna* - A multigenerational study. *Chemosphere* 79(1):60-66.



- Dijksterhuis J., Van Doorn T., Samson R., Postma J. (2011). Effects of seven fungicides on non-target aquatic fungi. *Water Air Soil Pollut.* 222(1-4):421-425.
- Dimitrie D.A.. (2010). The Effects of two insecticides on california anurans (*Rana sierrae* and *Pseudacris sierra*) and the implications for declining amphibian populations. M.S.Thesis, Southern Illinois University, Carbondale, IL:101 p.
- Ding Y., Landrum P.F., You J., Harwood A.D., Lydy M.J. (2012). Use of solid phase microextraction to estimate toxicity: relating fiber concentrations to toxicity - Part I. *Environ. Toxicol. Chem.* 31(9):2159-2167.
- Djomo J.E., Dauta A., Ferrier V., Narbonne J.F., Monkiedje A., Njine T., Garrigues P. (2004). Toxic effects of some major polyaromatic hydrocarbons found in crude oil and aquatic sediments on *Scenedesmus subspicatus*. *Water Res.* 38(7):1817-1821.
- Domingues I., Guilhermino L., Soares A.M.V.M., Nogueira A.J.A., Monaghan K.A. (2009). Influence of exposure scenario on pesticide toxicity in the midge *Kiefferulus calligaster* (Kieffer). *Ecotoxicol. Environ. Saf.* 72(2):450-457.
- Douglas M.T., Chanter D.O., Pell I.B., Burney G.M. (1986). A proposal for the reduction of animal numbers required for the acute toxicity to fish test (LC50 determination). *Aquat. Toxicol.* 8(4):243-249.
- Doyle C.J. and Lim R.P. (2005). Sexual behavior and impregnation success of adult male mosquitofish following exposure to 17beta-estradiol. *Ecotoxicol. Environ. Saf.* 61(3):392-397.
- Durand A.M., Rotteveel S., Collombon M.T., Van der Grinten E., Maas J.L., Verweij W. (2009). Toxicity measurements in concentrated water samples; evaluation and validation. RIVM (National Institute for Public Health and the Environment), report 607013010/2009.
- Dussault E.B., Balakrishnan V.K., Borgmann U., Solomon K.R., Sibley P.K. (2009). Bioaccumulation of the synthetic hormone 17alpha-ethinylestradiol in the benthic invertebrates *Chironomus tentans* and *Hyalella azteca*. *Ecotoxicol. Environ. Saf.* 72(6):1635-1641.
- Dussault E.B., Balakrishnan V.K., Solomon K.R., Sibley P.K. (2008b). Chronic toxicity of the synthetic hormone 17alpha-ethinylestradiol to *Chironomus tentans* and *Hyalella azteca*. *Environ. Toxicol. Chem.* 27(12):2521-2529.
- Dussault E.B., Balakrishnan V.K., Sverko E., Solomon K.R., Sibley P.K. (2008a). Toxicity of human pharmaceuticals and personal care products to benthic invertebrates. *Environ. Toxicol. Chem.* 27(2):425-432.
- Dworak T., Gonzalez C., Laaser C., Interwies E.(2005). The need for new monitoring tools to implement the WFD. *Environmental Science & Policy* 8:301-306.
- Dziewieczynski T.L. (2011). Short-term exposure to an endocrine disruptor affects behavioural consistency in male Threespine Stickleback. *Aquat. Toxicol.* 105(3/4):681-687.
- Ebert, I., Bachmann, J., Kühnen, U., Küster, A., Kussatz, C., Maletzki, D., Schlüter, C., 2011. Toxicity of the fluoroquinolone antibiotics enrofloxacin and ciprofloxacin to photoautotrophic aquatic organisms. *Environ. Toxicol. Chem.* 30:2786-2792.
- Ebrahimi M. (2007). Vitellogenin assay by enzyme-linked immunosorbant assay as a biomarker of endocrine disruptor chemicals pollution. *Pak. J. Biol. Sci.* 10(18):3109-3114.
- Eguchi, K., Nagase, H., Ozawa, M., Endoh, Y.S., Goto, K., Hirata, K., Miyamoto, K., Yoshimura, H., 2004. Evaluation of antimicrobial agents for veterinary use in the ecotoxicity test using microalgae. *Chemosphere* 57:1733-1738.
- Ekman D.R., Teng Q., Villeneuve D.L., Kahl M.D., Jensen K.M., Durhan E.J., Ankley G.T., Collette T.W. (2008). Investigating compensation and recovery of fathead minnow (*Pimephales promelas*) exposed to 17alpha-ethinylestradiol with metabolite profiling. *Environ. Sci. Technol.* 42(11):4188-4194.
- Elo B., Villano C.M., Govorko D., White L.A. (2007). Larval zebrafish as a model for glucose metabolism: expression of phosphoenolpyruvate carboxykinase as a marker for exposure to anti-diabetic compounds. *J. Mol. Endocrinol.* 38(4):433-440.
- Emtithal A.E.S. and Thanaa A.E.B. (2012). Efficacy of some insecticides on field populations of *Culex pipiens* (Linnaeus) from Egypt. *J. Basic Appl. Zool.* 65(1):62-73.
- Erickson B.E. (2002). Analyzing the ignored environmental contaminants. The U.S. geological survey reports some of the first monitoring data on pharmaceuticals and other emerging organic wastewater contaminants in U.S. streams. *Environ. Sci. Technol.* 36(7):140-145.
- Escher B.I. and Leusch F.D.L. (2012). Bioanalytical tools in water quality assessment. IWA publishing, London (UK).
- Escher B.I., Baumgartner B., Koller M., Treyer K., Lienent J., McAndell C.S. (2011). Environmental toxicology and risk assessment of pharmaceuticals from hospital wastewaters. *Water Research* 45(1):75-92.

- European Communities (2000). Directive 2000/60/EC of the European Parliament and of the Council of 23 October 2000 establishing a framework for community action in the field of water policy. Official journal of the European Communities L327(43), 1–72.
- European Parliament and the Council of the European Union (2008). Directive 2008/105/EC Environmental Quality Standards in the Field of Water Policy, Amending and Subsequently Repealing Council Directives 82/176/EEC, 83/513/EEC, 84/156/EEC, 84/491/EEC, 86/280/EEC and Amending Directive 2000/60/EC.
- European Union Risk Assessment Report (2008). Coal-tar pitch, high temperature: Rapporteur for the risk assessment of coal-tar pitch, high temperature is the Netherlands. CAS No: 65996-93-2; EINECS No: 266-028-2.
- European Union, 2012. Commission Regulation (EU) No 252/2012. Laying down methods of sampling and analysis for the official control of levels of dioxins, dioxin-like PCBs and non-dioxin-like PCBs in certain foodstuffs and repealing Regulation (EC) No 1883/2006
- Ewell W.S., Gorsuch J.W., Kringle R.O., Robillard K.A., Spiegel R.C. (1986). Simultaneous evaluation of the acute effects of chemicals on seven aquatic species. *Environ. Toxicol. Chem.* 5(9):831-840.
- Faria M., Carrasco L., Diez S., Riva M.C., Bayona J.M., Barata C. (2009). Multi-biomarker responses in the freshwater mussel *Dreissena polymorpha* exposed to polychlorobiphenyls and metals. *Comp. Biochem. Physiol. C Comp. Pharmacol. Toxicol.* 149(3):281-288.
- Faria M., Lopez M.A., Fernandez-Sanjuan M., Lacorte S., Barata C. (2010). Comparative toxicity of single and combined mixtures of selected pollutants among larval stages of the native freshwater mussels (*Unio elongatulus*) and the invasive zebra mussel (*Dreissena polymorpha*). *Sci. Total Environ.* 408(12):2452-2458.
- Farré M., Asperger D., Kantiani L., González S., Petrovic M., Barceló D. (2008). Assessment of the acute toxicity of triclosan and methyl triclosan in wastewater based on the bioluminescence inhibition of *Vibrio fischeri*. *Anal Bioanal Chem.* 390(8):1999-2007.
- Fent K. and Meier W. (1992). Tributyltin-induced effects on early life stages of minnows *Phoxinus phoxinus*. *Arch. Environ. Contam. Toxicol.* 22(4):428-438.
- Ferrari, B., Mons, R., Vollat, B., Fraysse, B., Paxeus, N., Lo Giuduce, R., Pollio, A., Garric, J., 2004. ENVIRONMENTAL RISK ASSESSMENT OF SIX HUMAN PHARMACEUTICALS: ARE THE CURRENT ENVIRONMENTAL RISK ASSESSMENT PROCEDURES SUFFICIENT FOR THE PROTECTION OF THE AQUATIC ENVIRONMENT? *Environ. Toxicol. Chem.* 23, 1344–1354.
- Ferreira A.L.G., Loureiro S., Soares A.M.V.M. (2008). Toxicity prediction of binary combinations of cadmium, carbendazim and low dissolved oxygen on *Daphnia magna*. *Aquat. Toxicol.* 89(1):28-39.
- Ferreira C.S.G., Nunes B.A., Henriques-Almeida J.M.D.M., Guilhermino L. (2007). Acute toxicity of oxytetracycline and florfenicol to the microalgae *Tetraselmis chuii* and to the crustacean *Artemia parthenogenetica*. *Ecotoxicol. Environ. Saf.* 67:452–8.
- Filby A.L. and Tyler C.R. (2007). Appropriate 'housekeeping' genes for use in expression profiling the effects of environmental estrogens in fish. *BMC Mol. Biol.* 8(10):13p.
- Filby A.L., Paull G.C., Searle F., Ortiz-Zarragoitia M., Tyler C.R. (2012). Environmental estrogen-induced alterations of male aggression and dominance hierarchies in fish: a mechanistic analysis. *Environ. Sci. Technol.* 46(6):3472-3479.
- Filby A.L., Thorpe K.L., Maack G., Tyler C.R. (2007). Gene expression profiles revealing the mechanisms of anti-androgen and estrogen-induced feminization in fish. *Aquat. Toxicol.* 81(2):219-231.
- Flaherty C.M. and Dodson S.I. (2005). Effects of pharmaceuticals on daphnia survival, growth, and reproduction. *Chemosphere* 61(2):200-207.
- Flores-Valverde A.M., Horwood J., Hill E.M. (2010). Disruption of the steroid metabolome in fish caused by exposure to the environmental estrogen 17alpha-ethinylestradiol. *Environ. Sci. Technol.* 44(9):3552-3558.
- Flynn K., Wedin M.B., Bonventre J.A., Dillon-White M., Hines J., Weeks B.S., Andre C., Schreiber M.P., Gagne. F. (2013). Burrowing in the freshwater mussel *Elliptio complanata* is sexually dimorphic and feminized by low levels of atrazine. *J. Toxicol. Environ. Health Part A* 76(20):1168-1181.
- Foran C.M., Bennett E.R., Benson W.H. (2000). Developmental evaluation of a potential non-steroidal estrogen: triclosan. *Mar Environ Res.* 50(1-5):153-156.
- Forget-Leray J., Landriau I., Minier C., Leboulenger F. (2005). Impact of endocrine toxicants on survival, development, and reproduction of the estuarine copepod *Eurytemora affinis* (Poppe). *Ecotoxicol. Environ. Saf.* 60(3):288-294.
- Fraker S.L. and Smith G.R. (2004). Direct and interactive effects of ecologically relevant concentrations of organic wastewater con-



- taminants on *Rana pipiens* tadpoles. *Environ. Toxicol.* 19(3):250-256.
- Frick L.P. and DeJimenez W.Q. (1963). Egg clutches as against individual eggs of *Australorbis glabratus* as test units in molluscicide evaluation. *Bull. W. H. O.* 29(2):286-287.
- Frick L.P. and DeJimenez W.Q. (1964). Molluscicidal qualities of three organo-tin compounds revealed by 6-hour and 24-hour exposures against representative stages and sizes of *Australorbis glabratus*. *Bull. W. H. O.* 31(3):429-431.
- Gagnon B., Marcoux G., Leduc R., Pouet M.-F., Thomas O. (2007). Emerging tools and sustainability of water-quality monitoring. *Trends in Analytical Chemistry* 26(4).
- Gala W.R. and Giesy J.P. (1992). Photo-Induced Toxicity of Anthracene to the Green Alga, *Selenastrum capricornutum*. *Arch. Environ. Contam. Toxicol.* 23(3):316-323.
- Garcia-Reyero N., Villeneuve D.L., Kroll K.J., Liu L., Orlando E.F., Watanabe K.H., Sepulveda M.S., Ankley G.T., Denslow N.D (2009). Expression signatures for a model androgen and antiandrogen in the fathead minnow (*Pimephales promelas*) ovary. *Environ. Sci. Technol.* 43(7):2614-261.
- Garten C.T.Jr. and Frank M.L. (1984). Comparison of toxicity to terrestrial plants with algal growth inhibition by herbicides. Inter-agency Agreement No.40-1067-80, Prepared for the U.S.EPA, Washington, D.C., by the Oak Ridge Natl.Lab., Oak Ridge, TN:32p.
- Gawish A.M., Issa A.M., Ali M.A., Ismail G.A. (2011). Histopathological, histochemical and biochemical studies on the effects of lorsban on the liver of Nile tilapia and the possible declaring effect of antioxidants. *Aust. J. Basic. Appl. Sci.* 5(12):75-94.
- Ghedira J., Jebali J., Bouraoui Z., Banni M., Chouba L., Boussetta H. (2009). Acute effects of chlorpyrifos-ethyl and secondary treated effluents on acetylcholinesterase and butyrylcholinesterase activities in *Carcinus maenas*. *J. Environ. Sci. (China)* 21(10):1467-1472.
- Ghekiere A., Verslycke T., Janssen C. (2006). effects of methoprene, nonylphenol, and estrone on the vitellogenesis of the mysid *Neomysis integer*. *Gen. Comp. Endocrinol.* 147(2):190-195.
- Gimeno S., Komen H., Jobling S., Sumpter J., Bowmer T. (1998). Demasculinisation of sexually mature male common carp, *Cyprinus carpio*, exposed to 4-tert-pentylphenol during spermatogenesis. *Aquat. Toxicol.* 43(2-3):93-109.
- Giron-Perez M.I., Barcelos-Garcia R., Vidal-Chavez Z.G., Romero-Banuelos C.A., Robledo-Marengo M.L. (2006). Effect of chlorpyrifos on the hematology and phagocytic activity of Nile tilapia cells (*Oreochromis niloticus*). *Toxicol. Mech. Methods* 16(9):495-499.
- Giusti A., Ducrot V., Joaquim-Justo C., Lagadic L. (2013). testosterone levels and fecundity in the hermaphroditic aquatic snail *Lymnaea stagnalis* exposed to testosterone and endocrine disruptors. *Environ. Toxicol. Chem.* 32(8):1740-1745.
- Goto T. and Hiromi J. (2003). Toxicity of 17alpha-ethynylestradiol and norethindrone, constituents of an oral contraceptive pill to the swimming and reproduction of cladoceran *Daphnia magna*, with special reference to their synergetic effect. *Mar. Pollut. Bull.* 47(1-6):139-142.
- Graff L., Isnard P., Cellier P., Bastide J., Cambon J.P., Narbonne J.F., Budzinski H., Vasseur P. (2003). Toxicity of chemicals to microalgae in river and in standard waters. *Environ. Toxicol. Chem.* 22(6):1368-1379.
- Greco L., Capri E., Rustad T. (2007). Biochemical responses in *Salmo salar* muscle following exposure to ethynylestradiol and tributyltin. *Chemosphere* 68:564-571.
- Greenwood R., Mills G.A., Roig B. (2007). Introduction to emerging tools and their use in water monitoring. *Trends in Analytical Chemistry* 26(4):263-267.
- Gross-Sorokin M.Y., Roast S.D. and Brighty G.C. (2006). Assessment of Feminization of Male Fish in English Rivers by the Environment Agency of England and Wales. *Environ Health Perspect* 114:147-151
- Grung M., Källqvist T., Thomas K. (2006). Initial assessment of eleven pharmaceuticals using the EMEA guideline in Norway. *Statens forurensningstilsyn (SFT)*, Oslo.
- Gustafson A.L., Stedman D.B., Ball J., Hillegass J.M., Flood A., Zhang C.X., Panzica-Kelly J., Cao J., Coburn A., Enright B.P., Tornesi M.B., Hetheridge M., Augustine-Rauch K.A. (2012). Inter-laboratory assessment of a harmonized zebrafish developmental toxicology assay – Progress report on phase I. *Reprod. Toxicol.* 33(2):155-164.
- Gutierrez M.F., Gagneten A.M., Paggi J.C. (2013). Acute and behavioral sensitivity of *Mesocyclops longisetus* to atrazine and endosulfan formulations under predation pressure. *Water Air Soil Pollut.* 224(1):9 p.
- Gyllenhammar I., Holm L., Eklund R., Berg C. (2009). Reproductive toxicity in *Xenopus tropicalis* after developmental exposure to environmental concentrations of ethynylestradiol. *Aquat. Toxicol.* 91:171-178.

- Ha M.H. and Choi J. (2008a). Effects of environmental contaminants on hemoglobin of larvae of aquatic midge, *Chironomus riparius* (Diptera: Chironomidae): a potential biomarker for ecotoxicity monitoring. *Chemosphere* 71(10):1928-1936.
- Ha M.H. and Choi J. (2008b). Chemical-induced alteration of hemoglobin expression in the 4th instar larvae of *Chironomus tentans* Mg. (Diptera: Chironomidae). *Environ. Toxicol. Pharmacol.* 25(3):393-398.
- Ha M.H. and Choi J. (2009). Effects of environmental contaminants on hemoglobin gene expression in *Daphnia magna*: a potential biomarker for freshwater quality monitoring. *Arch. Environ. Contam. Toxicol.* 57(2):330-337.
- Haap T. and Kohler H.R. (2009). Cadmium tolerance in seven *Daphnia magna* clones is associated with reduced hsp70 baseline levels and induction. *Aquat. Toxicol.* 94:131-137.
- Haeba M.H., Hilscherova K., Mazurova E., Blaha L. (2008). Selected endocrine disrupting compounds (vinclozolin, flutamide, ketoconazole and dicofol): effects on survival, occurrence of males, growth, molting and reproduction of *Daphnia magna*. *Environ. Sci. Pollut. Res.* 15(3):222-227.
- Hahlbeck E. (2004). The juvenile three-spined stickleback: model organism for the study of estrogenic and androgenic endocrine disruption in laboratory and field. Ph.D.Thesis, Stockholm University, Stockholm, Sweden:40p.
- Hahlbeck E., Griffiths R., Bengtsson B.E. (2004a). The juvenile three-spined stickleback (*Gasterosteus aculeatus* L.) as a model organism for endocrine disruption: I. sexual differentiation. *Aquat. Toxicol.* 70(4):287-310.
- Hahlbeck E., Katsiadaki I., Mayer I., Adolfsson-Erici M., James J., Bengtsson B.E. (2004b). The juvenile three-spined stickleback (*Gasterosteus aculeatus* L.) as a model organism for endocrine disruption ii - kidney hypertrophy, vitellogenin and spiggin induction. *Aquat. Toxicol.* 70(4):311-326.
- Halappa R. and David M. (2009a). Behavioural responses of the freshwater fish, *Cyprinus carpio* (Linnaeus) following sublethal exposure to chlorpyrifos. *Turk. J. Fish. Aquat. Sci.* 9(2):233-238.
- Halappa R. and David M. (2009b). In vivo inhibition of acetylcholinesterase activity in functionally different tissues of the freshwater fish, *Cyprinus carpio*, under chlorpyrifos exposure. *Drug Metab. Drug Interact.* 24(2-4):123-136.
- Hallgren P., Sorita Z., Berglund O., Persson A. (2012). Effects of 17alpha-ethinylestradiol on individual life-history parameters and estimated population growth rates of the freshwater Gastropods *Radix balthica* and *Bithynia tentaculata*. *Ecotoxicology* 21(3):803-810.
- Hallgren S. and Olsen K.H. (2010). Effects on guppy brain aromatase activity following short-term steroid and 4-nonylphenol exposures. *Environ. Toxicol.* 25(3):261-271.
- Halling-Sorensen, B., 2000. Algal toxicity of antibacterial agents used in intensive farming. *Chemosphere* 40, 731-739.
- Halling-Sorensen, B., Holten Lützhøft, H.-C., Andersen, H.R., Ingerslev, F., 2000. Environmental risk assessment of antibiotics: comparison of mecillinam, trimethoprim and ciprofloxacin. *J. Antimicrob. Chemother.* 46, 53-58.
- Hannas B.R., Wang Y.H., Thomson S., Kwon G., Li H., LeBlanc G.A. (2011). Regulation and dysregulation of vitellogenin mRNA accumulation in daphnids (*Daphnia magna*). *Aquat. Toxicol.* 101(2):351-357.
- Hano T., Oshima Y., Kinoshita M., Tanaka M., Mishima N., Ohya T., Yanagawa T., Wakamatsu Y., Ozato K., Honjo T. (2007). Quantitative bioimaging analysis of gonads in olvas-gfp/st-ii yi medaka (transgenic *Oryzias latipes*) exposed to ethinylestradiol. *Environ. Sci. Technol.* 41(4):1473-1479.
- Hashimoto S., Watanabe E., Ikeda M., Terao Y., Strussmann C.A., Inoue M., Hara A. (2009). Effects of ethinylestradiol on medaka (*Oryzias latipes*) as measured by sperm motility and fertilization success. *Arch. Environ. Contam. Toxicol.* 56(2):253-259.
- Hatch A.C.Jr. (1999). Photo-induced toxicity of PAHs to *Hyalella azteca* and *Chironomus tentans*: effects of mixtures and behavior. *Environ. Pollut.* 106(2):157-167.
- Helder T. (1980). Effects of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) on early life stages of the pike (*Esox lucius* L.). *Sci. Total Environ.* 14(3):255-264.
- Hemmer M.J., Salinas K.A., Harris P.S. (2011). Application of protein expression profiling to screen chemicals for androgenic activity. *Aquat. Toxicol.* 103(1/2):71-78.
- Hernando M.D., Ejerhoon M., Fernandez-Alba A.R., Chisti Y. (2003). Combined toxicity effects of MTBE and pesticides measured with *Vibrio fischeri* and *Daphnia magna* bioassays. *Water Res.* 37(17):4091-4098.
- Herwig H.J. and Holwerda D.A (1986). Cytochemical localization of tin in freshwater mussels exposed to di-n-butyltin dichloride. *Aquat. Toxicol.* 9(2-3):117-128.

- Hillegass J.M., Villano C.M., Cooper K.R., White L.A. (2007). Matrix metalloproteinase-13 is required for zebra fish (*Danio rerio*) development and is a target for glucocorticoids. *Toxicol. Sci.* 100(1):168-179.
- Hillegass J.M., Villano C.M., Cooper K.R., White L.A. (2008). Glucocorticoids alter craniofacial development and increase expression and activity of matrix metalloproteinases in developing zebrafish (*Danio rerio*). *Toxicol. Sci.* 102(2):413-424.
- Hirano M., Ishibashi H., Matsumura N., Nagao Y., Watanabe N., Watanabe A., Onikura N., Kishi K., Arizono K. (2004). Acute toxicity responses of two crustaceans, *Americamysis bahia* and *Daphnia magna*, to endocrine disrupters. *J. Health Sci.* 50(1):97-100.
- Hodson P.V., Dixon D.G., Kaiser K.L.E. (1984). measurement of median lethal dose as a rapid indication of contaminant toxicity to fish. *Environ. Toxicol. Chem.* 3(2):243-254.
- Hogan N.S., Currie S., LeBlanc S., Hewitt L.M., MacLatchy D.L. (2010). Modulation of steroidogenesis and estrogen signalling in the estuarine killifish (*Fundulus heteroclitus*) exposed to ethinylestradiol. *Aquat. Toxicol.* 98(2):148-156.
- Hogan N.S., Lean D.R.S., Trudeau V.L. (2006). Exposures to estradiol, ethinylestradiol and octylphenol affect survival and growth of *Rana pipiens* and *Rana sylvatica* tadpoles. *J. Toxicol. Environ. Health Part A* 69(15-16):1555-1569.
- Hogan N.S., Wartman C.A., Finley M.A., Van der Lee J.G., Van den Heuvel M.R. (2008). Simultaneous determination of androgenic and estrogenic endpoints in the threespine stickleback (*Gasterosteus aculeatus*) using quantitative RT-PCR. *Aquat. Toxicol.* 90(4):269-276.
- Holbech H., Kinnberg K., Petersen G.I., Jackson P., Hylland K., Norrgren L., Bjerregaard P. (2006). Detection of endocrine disrupters: evaluation of a fish sexual development test (FSDT). *Comp. Biochem. Physiol. C Comp. Pharmacol. Toxicol.* 144(1):57-66.
- Holbech H., Kinnberg K., Petersen G.I., Jackson P., Hylland K., Norrgren L., Bjerregaard P. (2006). Detection of endocrine disrupters: evaluation of a fish sexual development test (FSDT). *Comp. Biochem. Physiol. C Comp. Pharmacol. Toxicol.* 144(1):57-66.
- Holten Lützhof, H.-C., Halling-Sorensen, B., Jørgensen, S.E., 1999. Algal Toxicity of Antibacterial Agents Applied in Danish Fish Farming. *Arch* 36, 1-6.
- Hook S.E., Skillman A.D., Small J.A., Schultz I.R. (2006). Gene expression patterns in rainbow trout, *Oncorhynchus mykiss*, exposed to a suite of model toxicants. *Aquat. Toxicol.* 77(4):372-385.
- Huang D.J., Chen H.C., Wu J.P., Wang S.Y. (2006). Reproduction obstacles for the female green neon shrimp (*Neocaridina denticulata*) after exposure to chlordane and lindane. *Chemosphere* 64(1):11-16.
- Huang D.-J., Hou J.-H., Kuo T.-F., Lai H.-T. (2014). Toxicity of the veterinary sulfonamide antibiotic sulfamonomethoxine to five aquatic organisms. *Environ. Toxicol. Pharmacol.* 38:874-880.
- Huang G., Dai S., Sun H. (1996). Toxic effects of organotin species on algae. *Appl. Organomet. Chem.* 10(5):377-387.
- Huang G.Y., Ying G.G., Liang Y.Q., Liu Y.S., Liu S.S. (2013). Effects of steroid hormones on reproduction- and detoxification-related gene expression in adult male mosquitofish, *Gambusia affinis*. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* 158(1):36-43.
- Huang G.Y., Ying G.G., Liu S., Fang Y.X. (2012b). Regulation of reproduction- and biomarker-related gene expression by sex steroids in the livers and ovaries of adult female western mosquitofish (*Gambusia affinis*). *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* 162(1):36-43.
- Huang L., Xi Y., Zha C., Zhao L., Wen X. (2012a). Effects of dieldrin and 17beta-estradiol on life history characteristics of freshwater rotifer *Brachionus calyciflorus* Pallas. *J. Freshw. Ecol.* 27(3):381-392.
- Hulsen K., Minne V., Lootens P., Vandecasteele P., Hofte M. (2002). A chlorophyll a fluorescence-based *Lemna* minor bioassay to monitor microbial degradation of nanomolar to micromolar concentrations of linuron. *Environ. Microbiol.* 4(6):327-337.
- Huynh H.P.V. and Nugegoda D. (2012). Effects of chlorpyrifos exposure on growth and food utilization in australian catfish, *Tandanus tandanus*. *Bull. Environ. Contam. Toxicol.* 88(1):25-29.
- Irving E.C., Baird D.J., Culp J.M. (2009). cadmium toxicity and uptake by mats of the freshwater diatom: *Navicula pelliculosa* (Breb) hilse. *Arch. Environ. Contam. Toxicol.* 57:524-530.
- Ishibashi H., Matsumura N., Hirano M., Matsuoka M., Shiratsuchi H., Ishibashi Y., Takao Y., Arizono K. (2004). Effects of triclosan on the early life stages and reproduction of medaka *Oryzias latipes* and induction of hepatic vitellogenin. *Aquat. Toxicol.* 67(2):167-179.
- Isidori, M., Lavorgna, M., Nardelli, A., Pascarella, L., Parrella, A., 2005. Toxic and genotoxic evaluation of six antibiotics on non-target organisms. *Sci. Total Environ.* 346, 87-98. doi:10.1016/j.scitotenv.2004.11.017
- Jagtap J.T. and Shejule K.B. (2010). Study of acute toxicity of organotin tributyltin chloride on the freshwater bivalve mollusc, *Lamel-*

- lidens marginalis from godavari river at Maharashtra State, India. *Recent Res. Sci. Technol.* 2(5):8-11.
- James Laurenson P., Bloom R.A., Page S., Sadrieh N. (2014). Ethinyl Estradiol and Other Human Pharmaceutical Estrogens in the Aquatic Environment: A Review of Recent Risk Assessment Data. *The AAPS Journal*, Vol. 16, No. 2:299-310.
- Janer G., Lyssimachou A., Bachmann J., Oehlmann J., Schulte-Oehlmann U., Porte C. (2006). Sexual dimorphism in esterified steroid levels in the gastropod *Marisa cornuarietis*: the effect of xenoandrogenic compounds. *Steroids* 71(6):435-444.
- Janssens L. and Stoks R. (2012). How does a pesticide pulse increase vulnerability to predation? Combined effects on behavioral antipredator traits and escape swimming. *Aquat. Toxicol.* 110/111:91-98.
- Jarošová B., Bláha L., Giesy J.P., Hilscherová, K., 2014. What level of estrogenic activity determined by in vitro assays in municipal waste waters can be considered as safe? *Environment International* 64: 98–109.
- Jarvinen A.W., Tanner D.K., Kline E.R. (1988). Toxicity of chlorpyrifos, endrin, or fenvalerate to fathead minnows following episodic or continuous exposure. *Ecotoxicol. Environ. Saf.* 15:78–95.
- Jaylet A., Deparis P., Ferrier V., Grinfeld S., Siboulet R. (1986). A new micronucleus test using peripheral blood erythrocytes of the newt *Pleurodeles waltl* to detect mutagens in fresh-water pollution. *Mutat. Res.* 164 (4):245-257.
- Jensen K.M. and Ankley G.T. (2006). Evaluation of a commercial kit for measuring vitellogenin in the fathead minnow (*Pimephales promelas*). *Ecotoxicol. Environ. Saf.* 64(2):101-105.
- Jin Y., Chen R., Liu W., Fu Z. (2010). Effect of endocrine disrupting chemicals on the transcription of genes related to the innate immune system in the early developmental stage of zebrafish (*Danio rerio*). *Fish Shellfish Immunol.* 28(5-6):854-861.
- Jin Y., Chen R., Sun L., Liu W., Fu Z. (2009). Photoperiod and temperature influence endocrine disruptive chemical-mediated effects in male adult zebrafish. *Aquat. Toxicol.* 92(1):38-43.
- Johnson I., Hetheridge M., Tyler C.R. (2007). Assessment of (anti-) oestrogenic and (anti-) androgenic activities of final effluents from sewage treatment works. *Science Report – SC020118/SR*. Environment Agency, Bristol
- Johnson W.W. and Finley M.T. (1980). *Handbook of acute toxicity of chemicals to fish and aquatic invertebrates*. U.S.F.W.S., Resource. Pub.
- Jolly C., Katsiadaki I., Morris S., Le Belle N., Dufour S., Mayer I., Pottinger T.G., Scott A.P. (2009). Detection of the anti-androgenic effect of endocrine disrupting environmental contaminants using in vivo and in vitro assays in the three-spined stickleback. *Aquat. Toxicol.* 92(4):228-239.
- Jones J.R., Barrick C., Kim K.A., Lindner J., Blondeau B., Fujimoto Y., Shiota M., Kesterson R.A., Kahn B.B. and Magnuson M.A., 2005. Deletion of PPAR in adipose tissues of mice protects against high fat diet-induced obesity and insulin resistance. *PNAS Proc. Natl. Acad. Sci. U.S.A.* 102 (17): 6207–6212.
- Jones O., Voulvoulis N., Lester J. (2002). Aquatic environmental assessment of the top 25 English prescription pharmaceuticals. *Water Res.* 36:5013–5022.
- Jonsson M.E., Brunstrom B., Brandt I. (2009). The zebrafish gill model: induction of CYP1A, EROD and PAH adduct formation. *Aquat. Toxicol.* 91(1):62-70.
- Jukosky J.A., Watzin M.C., Leiter J.C. (2008a). Elevated concentrations of ethinylestradiol, 17beta-estradiol, and medroxyprogesterone have little effect on reproduction and survival of *Ceriodaphnia dubia*. *Bull. Environ. Contam. Toxicol.* 81(3):230-235.
- Jukosky J.A., Watzin M.C., Leiter J.C. (2008b). The effects of environmentally relevant mixtures of estrogens on japanese medaka (*Oryzias latipes*) reproduction. *Aquat. Toxicol.* 86(2):323-331.
- Julius M.L., Stepanek J., Tedrow O., Gamble C., Schoenfuss H.L. (2007). Estrogen-receptor independent effects of two ubiquitous environmental estrogens on *Melosira varians* Agardh, a common component of the aquatic primary production community. *Aquat. Toxicol.* 85(1):19-27.
- Jung J., Kim Y., Kim J., Jeong D.-H., Choi K. (2008). Environmental levels of ultraviolet light potentiate the toxicity of sulfonamide antibiotics in *Daphnia magna*. *Ecotoxicology* 17:37–45.
- Junghans M., Backhaus T., Faust M., Scholze M., Grimme L.H. (2006). Application and validation of approaches for the predictive hazard assessment of realistic pesticide mixtures. *Toxicol.* 76(2):93-110
- Kagan J., Kagan E.D., Kagan I.A., Kagan P.A. (1987). Do polycyclic aromatic hydrocarbons, acting as photosensitizers, participate in the toxic effects of acid rain?. *ACS Symp. Ser.* 327:191-204.
- Kagan J., Kagan P.A., Jr Buhse H.E. (1984). Light-dependent toxicity of alpha-terthienyl and anthracene toward late embryonic stages

- of *Rana pipiens*. *J. Chem. Ecol.* 10(7):1115-1122.
- Kallqvist T. (2009). Effect of water hardness on the toxicity of cadmium to the green alga *Pseudokirchneriella subcapitata* in an artificial growth medium and nutrient-spiked natural lake waters. *J. Toxicol. Environ. Health Part A* 72(3):277-283.
- Kamata R., Itoh K., Nakajima D., Kageyama S., Sawabe A., Terasaki M., Shiraishi F. (2011). The feasibility of using mosquitofish (*Gambusia affinis*) for detecting endocrine-disrupting chemicals in the freshwater environment. *Environ. Toxicol. Chem.* 30(12):2778-2785.
- Kammann U., Vobach M., Wosniok W., Schaffer A., Telscher A. (2009). Acute toxicity of 353-nonylphenol and its metabolites for zebrafish embryos. *Environ. Sci. Pollut. Res.* 16(2):227-231.
- Kang H.J., Choi K., Kim M.Y., Kim P.G. (2006b). Endocrine disruption induced by some sulfa drugs and tetracyclines on *Oryzias latipes*. *Hanguk Hwangyeong Bogeon Haghoeji* 32(3):227-234.
- Kang H.J., Kim H.S., Choi K.H., Kim K.T., Kim P.G. (2005). Several human pharmaceutical residues in aquatic environment may result in endocrine disruption in Japanese medaka (*Oryzias latipes*). *Hanguk Hwangyeong Bogeon Haghoeji* 31(3):227-233.
- Kang I.J., Hano T., Oshima Y., Yokota H., Tsuruda Y., Shimasaki Y., Honjo T. (2006a). Anti-androgen flutamide affects gonadal development and reproduction in medaka (*Oryzias latipes*). *Mar. Environ. Res.* 62 (suppl. 1):S253 - S257.
- Kang I.J., Yokota H., Oshima Y., Tsuruda Y., Yamaguchi T., Maeda M., Imada N., Tadokoro H., Honjo T. (2002). Effect of 17 $\beta$ -estradiol on the reproduction of Japanese medaka (*Oryzias latipes*). *Chemosphere* 47(1):71-80.
- Kaoud H.A., Zaki M.M., Ismail M.M. (2011). Effect of exposure to mercury on health in tropical *Macrobrachium rosenbergii*. *Life Sci. J.* 8(1):154-163.
- Kar S. and Aditya A.K. (2010). Impact of heavy metal and pesticide on total protein content in intact and regenerating hydra. *Environ. Ecol.* 28(3B):2003-2007.
- Kasherwani D., Lodhi H.S., Tiwari K.J., Shukla S., Sharma U.D. (2009). Cadmium toxicity to freshwater catfish, *Heteropneustes fossilis* (Bloch). *Asian J. Exp. Sci* 23(1):149-156.
- Kashian D.R. and Dodson S.I. (2004). Effects of vertebrate hormones on development and sex determination in *Daphnia magna*. *Environ. Toxicol. Chem.* 23(5):1282-1288.
- Kashiwada S., Ishikawa H., Miyamoto N., Ohnishi Y., Magara Y. (2002). Fish test for endocrine-disruption and estimation of water quality of Japanese rivers. *Water Res.* 36(8):2161-2166.
- Katsiadaki I., Williams T.D., Ball J.S., Bean T.P., Sanders M.B., Wu H., Santos E.M., Brown M.M., Baker P., Ortega F., Francesco F., Craft J.A., Tyler C.R., Viant M.R., Chipman J.K. (2010). Hepatic transcriptomic and metabolomic responses in the stickleback (*Gasterosteus aculeatus*) exposed to ethinyl-estradiol. *Aquat. Toxicol.* 97(3):174-187.
- Katsu Y., Lange A., Urushitani H., Ichikawa R., Paull G.C., Cahill L.L., Jobling S., Tyler C.R., Iguchi T. (2007). Functional associations between two estrogen receptors, environmental estrogens, and sexual disruption in the roach (*Rutilus rutilus*). *Environ. Sci. Technol.* 41(9):3368-3374.
- Kazeto Y., Place A.R., Trant J.M. (2004). Effects of endocrine disrupting chemicals on the expression of CYP19 genes in zebrafish (*Danio rerio*). Juveniles. *Aquat. Toxicol.* 69(1):25-34.
- Ke L.X., Xi Y.L., Zha C.W., Dong L.L. (2009). Effects of three organophosphorus pesticides on population growth and sexual reproduction of rotifer *Brachionus calyciflorus* Pallas. *Shengtai Xuebao* 29(3):182-185.
- Keithly J., Brooker J.A., DeForest D.K., Wu B.K., Brix K.V. (2004). Acute and chronic toxicity of nickel to a cladoceran (*Ceriodaphnia dubia*) and an amphipod (*Hyalella azteca*). *Environ. Toxicol. Chem.* 23(3):691-696.
- Khalil M., Furness D.N., Zholobenko V., Hoole D. (2014). Effect of tapeworm parasitisation on cadmium toxicity in the bioindicator copepod, *Cyclops strenuus*. *Ecol. Indic.* 37:21-26.
- Khengarot B.S. and Das S. (2009). Acute toxicity of metals and reference toxicants to a freshwater ostracod, *Cypris subglobosa* Sowerby, 1840 and correlation to EC50 values of other test models. *J. Hazard. Mater.* 172(2/3):641-649.
- Khayrandish A. and Wood R.J. (1993). Organophosphorus insecticide resistance in a new strain of *Culex quinquefasciatus* (Diptera: Culicidae) from Tanga, Tanzania. *Bull. Entomol. Res.* 83(1):67-74.
- Kienle C., Kase R., Werner I. (2011). Evaluation of bioassays and wastewater quality: In vitro and in vivo bioassays for the performance review in the Project "Strategy MicroPoll". Swiss Centre for Applied Ecotoxicology, Eawag-EPFL, Dübendorf.
- Kikuchi M. (1993). Toxicity evaluation of selected pesticides used in golf links by algal growth inhibition test. *Mizu Kankyo Gakkaiishi*



16(10):704-710.

- Kim J.W., Ishibashi H., Yamauchi R., Ichikawa N., Takao Y., Hirano M., Koga M., Arizono K. (2009). Acute toxicity of pharmaceutical and personal care products on freshwater crustacean (*Thamnocephalus platyurus*) and fish (*Oryzias latipes*). *J. Toxicol. Sci.* 34(2): 227-232
- Kim S., Ji K., Lee S., Lee J., Kim J., Kim S., Kho Y., Choi K. (2011). Perfluorooctane sulfonic acid exposure increases cadmium toxicity in early life stage of zebrafish, *Danio rerio*. *Environ. Toxicol. Chem.* 30(4):870-877.
- Kim Y. and Cooper K.R. (1998). Interactions of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) and 3,3',4,4',5-pentachlorobiphenyl (PCB 126) for producing lethal and sublethal effects in the japanese medaka embryos and larvae. *Chemosphere* 36(2):409-418.
- Kim Y. and Cooper K.R. (1999). Toxicity of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) and polychlorinated biphenyls (PCBs) in the embryos and newly hatched larvae of the japanese medaka (*Oryzias latipes*). *Chemosphere* 39(3):527-538.
- Kim Y., Choi K., Jung J., Park S., Kim P.-G., Park J. (2007). Aquatic toxicity of acetaminophen, carbamazepine, cimetidine, diltiazem and six major sulfonamides, and their potential ecological risks in Korea. *Environ. Int.* 33:370-5.
- Kiparissis Y., Metcalfe T.L., Balch G.C., Metcalfe C.D. (2003). Effects of the antiandrogens, vinclozolin and cyproterone acetate on gonadal development in the japanese medaka (*Oryzias latipes*). *Aquat. Toxicol.* 63(4):391-403.
- Kliwer S, Goodwin B, Willson T (2002). "The nuclear pregnane X receptor: a key regulator of xenobiotic metabolism". *Endocr. Rev.* 23 (5): 687-702.
- Knorr S. and Braunbeck T. (2002). Decline in reproductive success, sex reversal, and developmental alterations in japanese medaka (*Oryzias latipes*) after continuous exposure to octylphenol. *Ecotoxicol. Environ. Saf.* 51(3):187-196.
- Kołodziejka M., Maszkowska J., Białk-Bieli ska A., Steudte S., Kumirska J., Stepnowski P., Stolte S. (2013). Aquatic toxicity of four veterinary drugs commonly applied in fish farming and animal husbandry. *Chemosphere* 92, 1253-9.
- Kopf W. (1997). The action of endocrine substances in biological tests with aquatic organisms (wirkung endokriner stoffe in biotests mit wasserorganismen). *Munch.Beitr.Abwasser-, Fisch.- Flussbiol.*:13p.
- Korner O., Kohno S., Schonenberger R., Suter M.J.F., Knauer K., Jr. Guillelte L.J., Burkhardt-Holm P. (2008). Water temperature and concomitant waterborne ethinylestradiol exposure affects the vitellogenin expression in juvenile brown trout (*Salmo trutta*). *Aquat. Toxicol.* 90:188-196.
- Kramer V.J., Miles-Richardson S., Pierens S.L., Giesy J.P. (1998). Reproductive impairment and induction of alkaline-labile phosphate, a biomarker of estrogen exposure, in fathead minnows (*Pimephales promelas*) exposed to waterborne 17beta-estradiol. *Aquat. Toxicol.* 40(4):335-360.
- Kratky B.A. and Warren G.F. (1971). A rapid bioassay for photosynthetic and respiratory inhibitors. *Weed Sci.* 19(6):658-661.
- Kugathas S. (2011). Synthetic glucocorticoids in the aquatic environment: their potential impacts on fish. A thesis submitted to the degree of Doctor of Philosophy. Institute for the Environment Brunel University Uxbridge, Middlesex, UB8 3PH United Kingdom.
- Kumar S., Thomas A., Sahgal A., Verma A., Samuel T., Pillai M.K.K. (2004). Variations in the insecticide-resistance spectrum of *Anopheles stephensi* after selection with deltamethrin or a deltamethrin-piperonyl-butoxide combination. *Ann. Trop. Med. Parasitol.* 98(8):861-871.
- Kümmerer K. (2009). Antibiotics in the aquatic environment--a review--part I. *Chemosphere* 75:417-34.
- Kungolos A., Hadjispyrou S., Samaras P., Petala M., Tsiroidis V., Aravossis K., Sakellaropoulos G.P. (2001). Assessment of toxicity and bioaccumulation of organotin compounds. In: *Proceedings of the 7th International Conference on Environmental Science and Technology*, Syros, Greece:499-505.
- Kuykendall J.R., Miller K.L., Mellinger K.M., Cain A.J., Perry M.W., Bradley M., Jarvi E.J., Paustenbach D.J. (2009). DNA-protein cross-links in erythrocytes of freshwater fish exposed to hexavalent chromium or divalent nickel. *Arch. Environ. Contam. Toxicol.* 56(2):260-267.
- Laetz C.A., Baldwin D.H., Collier T.K., Hebert V., Stark J.D., Scholz N.L. (2009). The synergistic toxicity of pesticide mixtures: implications for risk assessment and the conservation of endangered pacific salmon. *Environ. Health Perspect.* 117(3):348-353.
- Lahnsteiner F. (2008). The sensitivity and reproducibility of the zebrafish (*Danio rerio*) embryo test for the screening of waste water quality and for testing the toxicity of chemicals. *ATLA* 36:299-311.
- Lahnsteiner F., Berger B., Kletzl M., Weismann T. (2006). Effect of 17beta-estradiol on gamete quality and maturation in two salmonid species. *Aquat. Toxicol.* 79(2):124-131.

- LaLone C.A., Villeneuve D.L., Olmstead A.W., Medlock E.K., Kahl M.D., Jensen K.M., Durhan E.J., Makynen E.A., Blank C.A. (2012). Effects of a glucocorticoid receptor agonist, dexamethasone, on fathead minnow reproduction, growth, and development. *Environ. Toxicol. Chem.* 31(3):611-622.
- Lampi M.A., Gurska J., McDonald K.I.C., Xie F., Huang X.D., Dixon D.G., Greenberg B.M. (2005). Photoinduced toxicity of polycyclic aromatic hydrocarbons to *Daphnia magna*: ultraviolet-mediated effects and the toxicity of polycyclic aromatic hydrocarbon photoproducts. *Environ. Toxicol. Chem.* 25(4):1079-1087.
- Lange A., Katsu Y., Ichikawa R., Paull G.C., Chidgey L.L., Coe T.S., Iguchi T., Tyler C.R. (2008). Altered sexual development in roach (*Rutilus rutilus*) exposed to environmental concentrations of the pharmaceutical 17alpha-ethinylestradiol and associated expression dynamics of aromatases and estrogen receptors. *Toxicol. Sci.* 106(1):113-123.
- Lange A., Katsu Y., Miyagawa S., Ogino Y., Urushitani H., Kobayashi T., Hirai T., Shears J.A., Nagae M., Yamamoto J., Ohnishi Y., Oka T., Tatarazako N., Ohta Y., Tyler C.R., Iguchi T. (2012). Comparative responsiveness to natural and synthetic estrogens of fish species commonly used in the laboratory and field monitoring. *Aquat. Toxicol.* 109:250-258.
- Lange A., Paull G.C., Coe T.S., Katsu Y., Urushitani H., Iguchi T., Tyler C.R. (2009). Sexual reprogramming and estrogenic sensitization in wild fish exposed to ethinylestradiol. *Environ. Sci. Technol.* 43(4):1219-1225.
- Langer-Jaesrich M., Kienle C., Kohler H.R., Gerhardt A. (2010). Impairment of trophic interactions between zebrafish (*Danio rerio*) and midge larvae (*Chironomus riparius*) by chlorpyrifos. *Ecotoxicology* 19(7):1294-1301.
- Lanzky, P., Halling-Sorensen, B., 1997. The toxic effect of the antibiotic metronidazole on aquatic organisms. *Chemosphere* 35, 2553-2561.
- Larsen M.G., Bilberg K., Baatrup E. (2009). Reversibility of estrogenic sex changes in zebrafish (*Danio rerio*). *Environ. Toxicol. Chem.* 28(8):1783-1785.
- Lawrence A.J. and Poulter C. (1998). Development of a sub-lethal pollution bioassay using the estuarine amphipod *Gammarus duebeni*. *Water Res.* 32(3):569-578.
- Le Mer C., Roy R.L., Pellerin J., Couillard C.M., Maltais D. (2013). Effects of chronic exposures to the herbicides atrazine and glyphosate to larvae of the threespine stickleback (*Gasterosteus aculeatus*). *Ecotoxicol. Environ. Saf.* 89(0):174-181.
- LeBlanc G.A. and McLachlan J.B. (2000). Changes in the metabolic elimination profile of testosterone following exposure of the crustacean *Daphnia magna* to tributyltin. *Ecotoxicol. Environ. Saf.* 45(3):296-303.
- LeBlanc G.A. and Surprenant D.C. (1980). The chronic toxicity of 8 of the 65 priority pollutants to the water flea (*Daphnia magna*). Draft Manuscr., EG&G Bionomics, Aquat.Toxicol.Lab., Wareham, MA:36p.
- LeBlanc H.M.K., Culp J.M., Baird D.J., Alexander A.C., Cessna A.J. (2012). Single versus combined lethal effects of three agricultural insecticides on larvae of the freshwater insect *Chironomus dilutus*. *Arch. Environ. Contam. Toxicol.* 63(3):378-390.
- Lee S.M., Lee S.B., Park C.H., Choi J. (2006). Expression of heat shock protein and hemoglobin genes in *Chironomus tentans* (Diptera, Chironomidae) larvae exposed to various environmental pollutants: a potential biomarker of freshwater monitoring. *Chemosphere* 65(6):1074-1081.
- Lee W., Kang C.W., Su C.K., Okubo K., Nagahama Y. (2012). Screening estrogenic activity of environmental contaminants and water samples using a transgenic medaka embryo bioassay. *Chemosphere* 88(8):945-952.
- Lei B., Kan J., Yu Y., Zha J., Li W., Wang Z., Wang Y, Wen Y. (2014). Long-term exposure investigating the estrogenic potency of estriol in Japanese medaka (*Oryzias latipes*). *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* 160:86-98
- León A., Teh S.J., Hall L.C., Teh F.C. (2007). Androgen disruption of early development in Qurt strain medaka (*Oryzias latipes*). *Aquat. Toxicol.* 82(3):195-203.
- Li M.H. (2013). Acute toxicity of industrial endocrine-disrupting chemicals, natural and synthetic sex hormones to the freshwater planarian, *Dugesia japonica*. *Toxicol. Environ. Chem.* 95(6):984-991.
- Li M.H. (2013). Acute toxicity of industrial endocrine-disrupting chemicals, natural and synthetic sex hormones to the freshwater planarian, *Dugesia japonica*. *Toxicol. Environ. Chem.* 95(6):984-991
- Li M.H. and Wang Z.R. (2005). Effect of nonylphenol on plasma vitellogenin of male adult guppies (*Poecilia reticulata*). *Environ. Toxicol.* 20(1):53-59.
- Li S. and Tan Y. (2011). Hormetic response of cholinesterase from *Daphnia magna* in chronic exposure to triazophos and chlorpyrifos. *J. Environ. Sci.* 23(5):852-858.

- Lister A., Regan C., Van Zwol J., Van der Kraak G. (2009). Inhibition of egg production in zebrafish by fluoxetine and municipal effluents: a mechanistic evaluation. *Aquat. Toxicol.* 95(4):320-329.
- Liu H., Cupp E.W., Micher K.M., Guo A., Liu N. (2004). Insecticide resistance and cross-resistance in Alabama and Florida strains of *Culex quinquefasciatus*. *J. Med. Entomol.* 41(3):408-413.
- Liu H., Yuan B., Li S. (2012). Altered quantities and in vivo activities of cholinesterase from *Daphnia magna* in sub-lethal exposure to organophosphorus insecticides. *Ecotoxicol. Environ. Saf.* 80:118-125.
- Liu N.A., Huang H., Yang Z., Herzog W., Hammerschmidt M., Lin S., Melmed S. (2003). Pituitary corticotroph ontogeny and regulation in transgenic zebrafish. *Molecular Endocrinology* 17(5):959-966.
- Liu X.C., Dong H.W., Zhou L., Du S.S., Liu Z.L. (2013). Essential oil composition and larvicidal activity of *Toddalia asiatica* roots against the mosquito *Aedes albopictus* (Diptera: Culicidae). *Parasitol. Res.* 112:1197-1203.
- Liwag A.G., Ocampo P.P., Ubaldo N.N. (2009). Effects of acetylcholinesterase-inhibiting chemicals on Nile tilapia (*Oreochromis niloticus* Linn) at pre- and post-hatching stages of development. *Philipp. Entomol.* 23(2):155-173.
- Lorenz C., Opitz R., Lutz I., Kloas W. (2009). Corticosteroids disrupt amphibian metamorphosis by complex modes of action including increased prolactin expression. *Comparative Biochemistry and Physiology, Part C: Toxicology & Pharmacology* 150(2):314-321.
- Loureiro S., Svendsen C., Ferreira A.L.G., Pinheiro C., Ribeiro F., Soares A.M. (2010). Toxicity of three binary mixtures to *Daphnia magna*: comparing chemical modes of action and deviations from conceptual models. *Environ. Toxicol. Chem.* 29(8):1716-1726.
- Lundqvist A., Bertilsson S., Goedkoop W. (2012). Interactions with DOM and biofilms affect the fate and bioavailability of insecticides to invertebrate grazers. *Ecotoxicology* 21(8):2398-2408.
- Lussier S.M. and Cardin J.A. (1985). Results of acute toxicity tests conducted with nickel at ERL, Narragansett. Memo from S.M.Lussier and J.A.Cardin to D.Hansen, U.S.EPA, Narragansett, RI:4p.
- Lyssimachou A., Janssen B.M., Arukwe A. (2006). Brain cytochrome P450 aromatase gene isoforms and activity levels in Atlantic salmon after waterborne exposure to nominal environmental concentrations of the pharmaceutical ethinylestradiol and anti-foulant tributyltin. *Toxicol. Sci.* 91(1):82-92.
- Ma J., Wang S., Wang P., Ma L., Chen X., Xu R. (2006). Toxicity assessment of 40 herbicides to the green alga *Raphidocelis subcapitata*. *Ecotoxicol. Environ. Saf.* 63(3):456-462.
- Ma J., Xu L., Wang S. (2002b). A quick, simple, and accurate method of screening herbicide activity using green algae cell suspension cultures. *Weed Sci.* 50(5):555-559.
- Ma J., Zheng R., Xu L., Wang S. (2002a). Differential sensitivity of two green algae, *Scenedesmus obliquus* and *Chlorella pyrenoidosa*, to 12 pesticides. *Ecotoxicol. Environ. Saf.* 52(1):57-61.
- Ma L., Li D., Wang J., He J., Yin Z. (2009). Effects of adrenergic agonists on the extrahepatic expression of vitellogenin Ao1 in heart and brain of the Chinese rare minnow (*Gobiocypris rarus*). *Aquat. Toxicol.* 91:19-25.
- Ma T., Wang Z., Gong S. (2007). Comparative sensitivity in Chinese rare minnow (*Gobiocypris rarus*) and Japanese medaka (*Oryzias latipes*) exposed to ethinylestradiol. *J. Environ. Sci. Health. Part A, Environ. Sci. Eng. Toxic Hazard. Substance Control* 42(7):889-894.
- Maas J. and van den Heuvel-Greve M.J. (2004). Opportunities for bio-analysis in WFD chemical monitoring using bioassays. RWS-RIZA working document.
- Mackenzie C.A., Berrill M., Metcalfe C., Pauli B.D. (2003). Gonadal differentiation in frogs exposed to estrogenic and antiestrogenic compounds. *Environ. Toxicol. Chem.* 22(10):2466-2475.
- Macova M., Escher B.I., Reungoat J., Carswell S., Lee Chue K., Keller J., Mueller J.F. (2010). Monitoring the biological activity of micropollutants during advanced wastewater treatment with ozonation and activated carbon filtration. *Water research* 44:477-492
- Macova M., Toze S., Hodggers L., Mueller J.F., Bartkow M., Escher B.I. (2011). Bioanalytical tools for the evaluation of organic micropollutants during sewage treatment, water recycling and drinking water generation. *Water research* 45:4238-4247.
- Makynen E.A., Kahl M.D., Jensen K.M., Tietge J.E., Wells K.L., Van der Kraak G., Ankley G.T. (2000). Effects of the mammalian antiandrogen vinclozolin on development and reproduction of the fathead minnow (*Pimephales promelas*). *Aquat. Toxicol.* 48(4):461-475.
- Malhi G.S. (2012). The Chronic toxicity of titanium dioxide nanoparticles to the freshwater amphipod *Hyalomma azteca*. M.S. Thesis,



Wilfrid Laurier University, Waterloo, Ontario, Canada:88p.

- Malla F.A., Sharma G., Singh S. (2009a). Chlorpyrifos pesticide toxicity on erythrocyte sedimentation rate in fish, *Channa punctatus* (Bloch). *Biol. Med.* 1(2):54-55.
- Malla F.A., Singh S., Singh A.P. (2009b). Changes in blood clotting time in the fish, *Channa punctatus*, on exposure to chlorpyrifos. *Bionotes* 11(2):67.
- Mallakin A., McConkey B.J., Miao G., McKibben B., Snieckus V., Dixon D.G., Greenberg B.M. (1999). Impact of structural photo-modification on the toxicity of environmental contaminants: anthracene photooxidation products. *Ecotoxicol. Environ. Saf.* 43(2):204-212.
- Maltby L. and Hills L. (2008). Spray drift of pesticides and stream macroinvertebrates: experimental evidence of impacts and effectiveness of mitigation measures. *Environ. Pollut.* 156(3):1112-1120.
- Mani V.G.T. and Konar S.K. (1986). Acute toxicity of some pesticides to fish, plankton and worm. *Environ. Ecol.* 4(1):121-123.
- Marquis O., Miaud C., Ficetola G.F., Bocher A., Mouchet F., Guittonneau S., Devaux A. (2009). Variation in genotoxic stress tolerance among frog populations exposed to UV and pollutant gradients. *Aquat. Toxicol.* 95(2):152-161.
- Martinovic D., Blake L.S., Durhan E.J., Greene K.J., Kahl M.D., Jensen K.M., Makynen E.A., Villeneuve D.L., Ank G.T. (2008). Reproductive toxicity of vinclozolin in the fathead minnow: confirming an anti-androgenic mode of action. *Toxicol. Chem.* 27(2):478-488.
- Martins J.C., Saker M.L., Oliva Teles L.F., Vasconcelos V.M. (2007). Oxygen consumption by *Daphnia magna* Straus as a marker of chemical stress in the aquatic environment. *Environ. Toxicol. Chem.* 26(9):1987-1991.
- Martyniuk C.J., Alvarez S., McClung S., Villeneuve D.L., Ankley G.T., Denslow N.D. (2009). Quantitative proteomic profiles of androgen receptor signaling in the liver of fathead minnows (*Pimephales promelas*). *J. Proteome Res.* 8(5):2186-2200.
- Martyniuk C.J., Gerrie E.R., Popescu J.T., Ekker M., Trudeau V.L. (2007). Microarray analysis in the zebrafish (*Danio rerio*) liver and telencephalon after exposure to low concentration of 17alpha-ethinylestradiol. *Aquat. Toxicol.* 84(1):38-49.
- Martyniuk C.J., Kroll K.J., Doperalski N.J., Barber D.S., Denslow N.D. (2010). Environmentally relevant exposure to 17alpha-ethinylestradiol affects the telencephalic proteome of male fathead minnows. *Aquat. Toxicol.* 98(4):344-353.
- Mastrangelo M., Afonso M.D.S., Ferrari L. (2011). Cadmium toxicity in tadpoles of *Rhinella arenarum* in relation to calcium and humic acids. *Ecotoxicology* 20(6):1225-1232.
- Matsumoto K.I., Hosokawa M., Kuroda K., Endo G. (2009). Toxicity of agricultural chemicals in *Daphnia magna*. *Osaka City Med. J.* 55(2):89-97.
- Matsumura N., Ishibashi H., Hirano M., Nagao Y., Watanabe N., Shiratsuchi H., Kai T., Nishimura T., Kashiwagi A., Arizono K. (2005). Effects of nonylphenol and triclosan on production of plasma vitellogenin and testosterone in male south african clawed frogs (*Xenopus laevis*). *Biol. Pharm. Bull.* 28(9):1748-1751.
- Maunder R.J., Matthiessen P., Sumpter J.P., Pottinger T.G. (2007). Impaired reproduction in three-spined sticklebacks exposed to ethinyl estradiol as juveniles. *Biol. Reprod.* 77(6): 999-1006
- Mayer F.L. Jr. (1987). Acute toxicity handbook of chemicals to estuarine organisms. EPA 600/8-87-017, U.S.EPA, Gulf Breeze, FL:274p.
- McCloskey J.T. and Oris J.T. (1991) Effect of water temperature and dissolved oxygen concentration on the photo-induced toxicity of anthracene to juvenile bluegill sunfish (*Lepomis macrochirus*). *Aquat. Toxicol.* 21(3-4):145-156.
- McGee M.R., Julius M.L., Vajda A.M., Norris D.O., Barber L.B., Schoenfuss H.L. (2009). Predator avoidance performance of larval fathead minnows (*Pimephales promelas*) following short-term exposure to estrogen mixtures. *Aquat. Toxicol.* 91:355-361.
- Meador J.P. (1986). An analysis of photobehavior of *Daphnia magna* exposed to tributyltin. In: *Oceans 86*, IEEE Publishing Services, New York, NY:1213-1218.
- Mebane C.A., Hennessy D.P., Dillon F.S. (2008). Developing acute-to-chronic toxicity ratios for lead, cadmium, and zinc using rainbow trout, a mayfly, and a midge. *Water Air Soil Pollut.* 188(1-4):41-66.
- Metcalfe C.D., Metcalfe T.L., Cormier J.A., Huestis S.Y., Niimi A.J. (1997). Early life-stage mortalities of japanese medaka (*Oryzias latipes*) exposed to polychlorinated diphenyl ethers. *Environ. Toxicol. Chem.* 16(8):1749-1754.
- Metcalfe C.D., Metcalfe T.L., Kiparissis Y., Koenig B.G., Khan C., Hughes R.J., Croley T.R., March R.E., Potter T. (2001). Estrogenic potency of chemicals detected in sewage treatment plant effluents as determined by in vivo assays with japanese medaka (*Oryzias latipes*). *Environ. Toxicol. Chem.* 20(2):297-308.

- Miana P., Scotto S., Perin G., Argese E. (1993). Sensitivity of *Selenastrum capricornutum*, *Daphnia magna* and submitochondrial particles to tributyltin. *Environ. Technol.* 14(2):175-181.
- Micael J., Reis-Henriques M.A., Carvalho A.P., Santos M.M. (2007). Genotoxic effects of binary mixtures of xenoandrogens (tributyltin, triphenyltin) and a xenoestrogen (ethinylestradiol) in a partial life-cycle test with zebrafish (*Danio rerio*). *Environ. Int.* 33(8):1035-1039.
- Migliore L., Brambilla G., Casoria P., Civitareale C., Cozzolino S., Gaudio L. (1996). Effect of sulphadimethoxine contamination on barley (*Hordeum distichum* L., Poaceae, Liliopsida). *Agric. Ecosyst. Environ.* 60, 121-128.
- Miller R.A., Norris L.A., Hawkes C.L. (1973). Toxicity of 2,3,1,8-tetrachlorodibenzo-p-dioxin (TCDD) in aquatic organisms. *Environ. Health Persp.* 5:177-186.
- Miyoshi N., Kawano T., Tanaka M., Kadono T., Kosaka T., Kunimoto M., Takahashi T., Hosoya H. (2003). Use of paramecium species in bioassays for environmental risk management: determination of IC50 values for water pollutants. *J. Health Sci.* 49(6):429-435.
- Mouchet F., Gauthier L., Mailhes C., Ferrier V., Devaux A. (2006). Comparative evaluation of genotoxicity of captan in amphibian larvae (*Xenopus laevis* and *Pleurodeles waltl*) using the comet assay and the micronucleus test. *Environ. Toxicol.* 21(3):264-277.
- Mugni H., Demetrio P., Marino D., Ronco A., Bonetto C. (2010). Toxicity persistence following an experimental cypermethrin and chlorpyrifos application in pampasic surface waters (Buenos Aires, Argentina). *Bull. Environ. Contam. Toxicol.* 84(5):524-528.
- Mugni H., Paracampo A., Marrochi N., Bonetto C. (2012). Cypermethrin, chlorpyrifos and endosulfan toxicity to two non-target freshwater organisms. *Fresenius Environ. Bull.* 21(8):2085-2089.
- Munoz M.J. and Tarazona J.V.(1993). Synergistic effect of two- and four-component combinations of the polycyclic aromatic hydrocarbons: phenanthrene, anthracene, naphthalene and acenaphthene on *Daphnia magna*. *Bull. Environ. Contam. Toxicol.* 50(3):363-368.
- Murk, A.J., J. Legler, M.S. Denison, J.P. Giesy, C. van de Guchte, and A. Brouwer, 1996). Chemical-Activated Luciferase Gene Expression (CALUX): A Novel in-Vitro Bioassay for Ah Receptor Active Compounds in Sediments and Pore Water. *Fun. Appl. Toxicol.* 33 (2): 149-160.
- Mwangi J.N., Wang N., Ingersoll C.G., Hardesty D.K., Brunson E.L., Li H., Deng B. (2012). Toxicity of carbon nanotubes to freshwater aquatic invertebrates. *Environ. Toxicol. Chem.* 31(8):1823-1830.
- Nallani G.C., Paulos P.M., Venables B.J., Edziyie R.E., Constantine L.A., Huggett D.B. (2012). Tissue-specific uptake and bioconcentration of the oral contraceptive norethindrone in two freshwater fishes. *Arch. Environ. Contam. Toxicol.* 62(1):306-313.
- Nassef M., Matsumoto S., Seki M., Khalil F., Kang I.J., Shimasaki Y., Oshima Y., Honjo T. (2010). Acute effects of triclosan, diclofenac and carbamazepine on feeding performance of japanese medaka fish (*Oryzias latipes*). *Chemosphere* 80(9):1095-1100.
- Nendza M. and Wenzel A. (2006). Discriminating toxicant classes by Mode of Action 1. (Eco)toxicity profiles. *Environ. Sci. Pollut.* 13(3):192-203.
- Newsted J.L. and Giesy J.P. (1987). Predictive models for photoinduced acute toxicity of polycyclic aromatic hydrocarbons to *Daphnia magna*, strauss (Cladocera, Crustacea). *Environ. Toxicol. Chem.* 6(6):445-461.
- Nguyen, T., Nioi, P., and Pickett, C. B. (2009) The Nrf2-antioxidant response element signaling pathway and its activation by oxidative stress. *J. Biol. Chem.* 284 (20): 13291-13295.
- Nie X., Gu J., Lu J., Pan W., Yang Y. (2009). Effects of norfloxacin and butylated hydroxyanisole on the freshwater microalga *Scenedesmus obliquus*. *Ecotoxicology* 18:677-84.
- Nielsen L. and Baatrup E. (2006). Quantitative studies on the effects of environmental estrogens on the testis of the guppy, *Poecilia reticulata*. *Aquat. Toxicol.* 80(2):140-148.
- Nishimura N., Fukazawa Y., Uchiyama H., Iguchi T. (1997). Effects of estrogenic hormones on early development of *Xenopus laevis*. *J. Exp. Zool.* 278(4):221-233.
- Nunes, B., Carvalho, F., Guilhermino, L., 2005. Acute toxicity of widely used pharmaceuticals in aquatic species: *Gambusia holbrooki*, *Artemia parthenogenetica* and *Tetraselmis chunii*. *Ecotoxicol. Environ. Saf.* 61, 413-419.
- Oberdorster E., Rittschof D., LeBlanc G.A. (1998). Alteration of (14C)-testosterone metabolism after chronic exposure of *Daphnia magna* to tributyltin. *Arch. Environ. Contam. Toxicol.* 34(1):21-25.
- OECD (2012). Draft report of the test method validation zebrafish embryo toxicity test (ZFET)-Phase 2 testing of 13 chemicals. 35p. Oekotoxzentrum, Centre Ecotox, [http://www.oekotoxzentrum.ch/expertenservice/qualitaetskriterien/vorschlaege/index\\_EN](http://www.oekotoxzentrum.ch/expertenservice/qualitaetskriterien/vorschlaege/index_EN)

- Ogueji O.E. (2008). Comparative acute toxicity of chlorpyrifos-ethyl (organophosphate) and lambda-cyhalothrin (pyrethroid) to the african catfish (*C. gariepinus*) using some biochemical parameters. *Global J. Pure Appl. Sci.* 14(3):263-269.
- Olivieri C.E. and Cooper K.R. (1997). Toxicity of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) in embryos and larvae of the fathead minnow (*Pimephales promelas*). *Chemosphere* 34(5-7):1139-1150.
- Oris J.T., Giesy J.P., Allred P.M., Grant D.F., Landrum P.F.(1984). Photoinduced toxicity of anthracene in aquatic organisms: an environmental perspective. T.N.Veziroglu (Ed.), *The Biosphere: Problems and Solutions*, Elsevier Science Publ., Amsterdam, Netherlands 639-658.
- Oruc E. (2012). Oxidative stress responses and recovery patterns in the liver of *Oreochromis niloticus* exposed to chlorpyrifos-ethyl. *Bull. Environ. Contam. Toxicol.* 88(5):678-684.
- Oruc E.O. (2010). Oxidative stress, steroid hormone concentrations and acetylcholinesterase activity in *Oreochromis niloticus* exposed to chlorpyrifos. *Pestic. Biochem. Physiol.* 96(3):160-166.
- Orvos D.R., Versteeg D.J., Inauen J., Capdevielle M., Rothenstein A., Cunningham V. (2002). Aquatic toxicity of triclosan. *Environ. Toxicol. Chem.* 21(7):1338-1349.
- OSPAR Agreement (2014-05). Establishment of a list of Predicted No Effect Concentrations (PNECs) for naturally occurring substances in produced water.
- Ostrander G.K., Landolt M.L., Kocan R.M. (1988). The ontogeny of coho salmon (*Oncorhynchus kisutch*) behavior following embryonic exposure to benzo(a)pyrene. *Aquat. Toxicol.* 13(4):325-346.
- Overturf M.D., Overturf C.L., Baxter D., Hala D.N., Constantine L., Venables B., Huggett D.B. (2012). Early life-stage toxicity of eight pharmaceuticals to the fathead minnow, *Pimephales promelas*. *Arch. Environ. Contam. Toxicol.* 62(3):455-464.
- Ozkan F., Gunduz S.G., Berkoz M., Hunt A.O., Yalin S. (2011). Effects of dietary supplementation of vitamin C on level of protein carbonyl in liver and nuclear abnormalities in peripheral erythrocytes of *Oreochromis niloticus*. *Fresenius Environ. Bull.* 20(6A):1520-1524.
- Ozkan F., Gunduz S.G., Berkoz M., Hunt A.O., Yalin S. (2012). The protective role of ascorbic acid (vitamin C) against chlorpyrifos-induced oxidative stress in *Oreochromis niloticus*. *Fish Physiol. Biochem.* 38(3):635-643.
- Pablos M.V., Fernandez C., del Mar Babin M., Navas J.M., Carbonell G., Martini F., Garcia-Hortiguera P., Tarazona J.V.(2009). Use of a novel battery of bioassays for the biological characterisation of hazardous wastes. *Ecotoxicology and Environmental Safety* 72(2009)1594-1600
- Padilla S., Corum D., Padnos B., Hunter D.L., Beam A., Houck K.A., Sipes N., Kleinstreuer N., Knudsen T., Dix D.J., Reif D.M. (2012). Zebrafish developmental screening of the ToxCast™ Phase I chemical library. *Reprod. Toxicol.* 33(2):174-187.
- Palace V.P., Evans R.E., Wautier K.G., Mills K.H., Blanchfield P.J., Park B.J., Baron C.L., Kidd K.A. (2009). Interspecies differences in biochemical, histopathological, and population responses in four wild fish species exposed to ethynylestradiol added to a whole lake. *Can. J. Fish. Aquat. Sci.* 66(11):1920-1935.
- Palma P., Palma V.L., Fernandes R.M., Bohn A., Soares A.M., Barbosa I.R. (2009). Embryo-toxic effects of environmental concentrations of chlorpyrifos on the crustacean *Daphnia magna*. *Ecotoxicol. Environ. Saf.* 72(6):1714-1718.
- Panter G.H., Hutchinson T.H., Hurd K.S., Sherren A., Stanley R.D., Tyler C.R. (2004). Successful detection of (anti-)androgenic and aromatase inhibitors in pre-spawning adult fathead minnows (*Pimephales promelas*) using easily measured endpoints of sexual development. *Aquat. Toxicol.* 70(1):11-21.
- Panter G.H., Hutchinson T.H., Hurd K.S., Sherren A., Stanley R.D., Tyler C.R. (2004). Successful detection of (anti-)androgenic and aromatase inhibitors in pre-spawning adult fathead minnows (*Pimephales promelas*) using easily measured endpoints of sexual development. *Aquat. Toxicol.* 70(1):11-21.
- Park B.J. (2003). Effects of the environmental estrogen 17alpha-ethynylestradiol on early development of green frogs (*Rana clamitans*) and mink frogs (*R. septentrionalis*) at the experimental lakes area (Ontario, Canada). M.S.Thesis, University of Manitoba, Canada:146p.
- Park B.J. and K. Kidd. (2005). Effects of the synthetic estrogen ethynylestradiol on early life stages of mink frogs and green frogs in the wild and in situ. *Environ. Toxicol. Chem.* 24(8):2027-2036.
- Park J.W., Tompsett A.R., Zhang X., Newsted J.L., Jones P.D., Au D.W.T., Kong R., Wu R.S.S., Giesy J.P., Hecker M. (2009). Advanced fluorescence in situ hybridization to localize and quantify gene expression in japanese medaka (*Oryzias latipes*) exposed to

- endocrine-disrupting compounds. *Environ. Toxicol. Chem.* 28(9):1951-1962.
- Park S. and Choi K. (2008). Hazard assessment of commonly used agricultural antibiotics on aquatic ecosystems. *Ecotoxicology* 17:526-38.
- Park S.Y., Nair P.M.G., Choi J. (2012). Characterization and expression of superoxide dismutase genes in *Chironomus riparius* (Diptera, Chironomidae) larvae as a potential biomarker of ecotoxicity. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* 156(3/4):187-194.
- Partridge C., Boettcher A., Jones A.G. (2010). Short-term exposure to a synthetic estrogen disrupts mating dynamics in a pipefish. *Horm. Behav.* 58(5):800-807.
- Patra R.W., Chapman J.C., Lim R.P., Gehrke P.C., Sunderam R.M. (2009). Effects of temperature on ventilatory behavior of fish exposed to sublethal concentrations of endosulfan and chlorpyrifos. *Environ. Toxicol. Chem.* 28(10):2182-2190.
- Pennington P.L. (2002). The replicated modular estuarine mesocosm: assessing direct and indirect effects of pesticide exposure. Ph.D.Thesis, University of South Carolina, Columbia, SC:249p.
- Perez D.J., Lukaszewicz G., Menone M.L., Camadro E.L. (2011a). Sensitivity of *Bidens laevis* L. to mutagenic compounds. Use of chromosomal aberrations as biomarkers of genotoxicity. *Environ. Pollut.* 159(1):281-286.
- Perez J., Domingues I., Monteiro M., Soares A.M., Loureiro S. (2013a). Synergistic effects caused by atrazine and terbuthylazine on chlorpyrifos toxicity to early-life stages of the zebrafish *Danio rerio*. *Environ. Sci. Pollut. Res. Int.* 20(7):4671-4680.
- Perez J., Domingues I., Soares A.M., Loureiro S. (2011b). Growth rate of *pseudokirchneriella subcapitata* exposed to herbicides found in surface waters in the Alqueva reservoir (Portugal): a bottom-up approach using binary mixtures. *Ecotoxicology* 20(6):1167-1175.
- Perez J., Monteiro M.S., Quintaneiro C., Soares A.M., Loureiro S. (2013b). Characterization of cholinesterases in *Chironomus riparius* and the effects of three herbicides on chlorpyrifos toxicity. *Aquat. Toxicol.* 144/145:296-302.
- Peterson J.K., Kashian D.R., Dodson S.I. (2001). Methoprene and 20-OH-Ecdysone affect male production in *Daphnia pulex*. *Environ. Toxicol. Chem.* 20(3):582-588.
- Peute J., Huiskamp R., Van Oordt P.G.W.J. (1985). Quantitative analysis of estradiol-17beta-induced changes in the ultrastructure of the liver of the male zebrafish, *Brachydanio rerio*. *Cell Tissue Res.* 242:377-382.
- Pickford D.B. and Morris I.D. (1999). Effects of endocrine-disrupting contaminants on amphibian oogenesis: methoxychlor inhibits progesterone-induced maturation of *Xenopus laevis* oocytes in vitro. *Environ. Health Perspect.* 107(4):285-292.
- Piferrer F. and Donaldson E.M. (1992). The comparative effectiveness of the natural and a synthetic estrogen for the direct feminization of chinook salmon (*Oncorhynchus tshawytscha*). *Aquaculture* 106(2):183-193.
- Pikkemaat M.G., Dijk S.O., Schouten J., Rapallini M., van Egmond H.J. (2008). A new microbial screening method for the detection of antimicrobial residues in slaughter animals: The Nouws antibiotic test (NAT-screening). *Food Control* 19:781-789.
- Plhalova L., Haluzova I., Macova S., Dolezelova P., Praskova E., Marsalek P., Skoric M., Svobodova Z., Pistekova V., Bedanova I. (2011). Effects of subchronic exposure to simazine on zebrafish (*Danio rerio*). *Neuroendocrinol. Lett.* 32(Suppl. 1):89-94.
- Pomati F., Netting A.G., Calamari D., Neilan B. (2004). Effects of erythromycin, tetracycline and ibuprofen on the growth of *Synechocystis* sp. and *Lemna minor*. *Aquat. Toxicol.* 67:387-96.
- Posthuma, L., G.W. Suter II, and TP Traas. 2002. *Species Sensitivity Distributions in Ecotoxicology*. Lewis Publishers, Boca Raton, FL. 587 pp.
- Poulsen A., Chapman H., Leusch F., Escher B. (2011). Application of Bioanalytical Tools for Water Quality Assessment. Urban Water Security Research Alliance Technical Report No. 41
- Pourkhabbaz A., Khazaei T., Behraves S., Ebrahimpour M., Pourkhabbaz H. (2011). Effect of water hardness on the toxicity of cobalt and nickel to a freshwater fish, *Capoeta fusca*. *Biomed. Environ. Sci.* 24(6):656-660.
- Prasertsup P. and Ariyakanon N. (2011). Removal of chlorpyrifos by water lettuce (*Pistia stratiotes* L.) and duckweed (*Lemna minor* L.). *Int. J. Phytoremediat.* 13:383-395.
- Prescott L.M., Kubovec M.K., Tryggstad D. (1977). The effects of pesticides, polychlorinated biphenyls and metals on the growth and reproduction of *Acanthamoeba castellanii*. *Bull. Environ. Contam. Toxicol.* 18(1): 29-34.
- Preston B.L., Snell T.W., Robertson T.L., Dingmann B.J. (2000). Use of freshwater rotifer *Brachionus calyciflorus* in screening assay for potential endocrine disruptors. *Environ. Toxicol. Chem.* 19(12):2923-2928.

- Pro J., Ortiz J., Boleas S., Fernández C., Carbonell G., Tarazona J.V. (2003). Effect assessment of antimicrobial pharmaceuticals on the aquatic plant *Lemna minor*. *Bull. Environ. Contam. Toxicol.* 70:290–5.
- Propst T.L., Fort D.J., Stover E.L. (1997). Evaluation of the developmental toxicity of benzo(a)pyrene and 2-acetylaminofluorene using *Xenopus*: modes of biotransformation. *Drug Chem. Toxicol. (N.Y.)* 20(1-2):45-61.
- Purdom C.E., Hardiman P.A., Bye V.J., Eno N.C., Tyler C.R., Sumpter J.P. (1994). Estrogenic effects of effluents from sewage treatment works. *Chem. Ecol.* 8:275-285.
- Puttaswamy N. and Liber K. (2012). Influence of inorganic anions on metals release from oil sands coke and on toxicity of nickel and vanadium to *Ceriodaphnia dubia*. *Chemosphere* 86(5):521-529.
- Qian H., Li J., Pan X., Sun Z., Ye C., Jin G., Fu Z. (2010). Effects of Streptomycin on Growth of Algae *Chlorella vulgaris* and *Microcystis aeruginosa*. *Environ. Toxicol.* 229–237.
- Qian H., Pan X., Chen J., Zhou D., Chen Z., Zhang L., Fu Z. (2012). Analyses of gene expression and physiological changes in *Microcystis aeruginosa* reveal the phytotoxicities of three environmental pollutants. *Ecotoxicology* 21(3):847-859.
- Quinn B., Gagné F., Blaise C. (2008). An investigation into the acute and chronic toxicity of eleven pharmaceuticals (and their solvents) found in wastewater effluent on the cnidarian, *Hydra attenuata*. *Sci. Total Environ.* 389:306–14.
- Ra J.S., Oh S.Y., Lee B.C., Kim S.D. (2008). The effect of suspended particles coated by humic acid on the toxicity of pharmaceuticals, estrogens, and phenolic compounds. *Environ. Int.* 34(2):184-192.
- Ramos E.U., Vaes W.H.J., Mayer P., Hermens J.L.M. (1999). Algal growth inhibition of *Chlorella*
- Ramos E.U., Vermeer C., Vaes W.H.J., Hermens J.L.M. (1998). Acute toxicity of polar narcotics to three aquatic species (*Daphnia magna*, *Poecilia reticulata* and *Lymnaea stagnalis*) and its relation to hydrophobicity. *Chemosphere* 37(4):633-650.
- Ranatunge R.A., Wijesinghe M.R., Ratnasooriya W.D., Dharmarathne H.A., Wijesekera R.D. (2012). Cadmium-induced toxicity on larvae of the common asian toad *Duttaphrynus melanostictus* (Schneider 1799): evidence from empirical trials. *Bull. Environ. Contam. Toxicol.* 89(1):143-146.
- Rawson C.A., Lim R.P., Warne M.S.J., Doyle C.J. (2006). The effect of 17 beta-estradiol on the development of modified hemal spines in early-life stage *Gambusia holbrooki*. *Arch. Environ. Contam. Toxicol.* 51(2):253-262.
- Reddy P.S., Reddy P.R., Sainath S.B. (2011). Cadmium and mercury-induced hyperglycemia in the fresh water crab, *Oziotelphusa senex senex*: involvement of neuroendocrine system. *Ecotoxicol. Environ. Saf.* 74(3):279-283.
- Reyhalian N., Volkova K., Hallgren S., Bollner T., Olsson P.E., Olsen H., Hallstrom I.P. (2011). 17alpha-ethinyl estradiol affects anxiety and shoaling behavior in adult male zebra fish (*Danio rerio*). *Aquat. Toxicol.* 105(1/2):41-48.
- Reynaud S., Worms I.A.M., Veyrenc S., Portier J., Maitre A., Miaud C., Raveton M. (2012). Toxicokinetic of benzo(a)pyrene and fipronil in female green frogs (*Pelophylax kl. esculentus*). *Environ. Pollut.* 161:206-214.
- Rhen T. and Cidlowski J.A., 2005. Antiinflammatory action of glucocorticoids--new mechanisms for old drugs. *N. Engl. J. Med.* 353 (16): 1711–1723.
- Richards, S.M., Cole, S.E., 2006. A toxicity and hazard assessment of fourteen pharmaceuticals to *Xenopus laevis* larvae. *Ecotoxicology* 15, 647–56. doi:10.1007/s10646-006-0102-4
- Rico A., Dimitrov M.R., Van Wijngaarden R.P., Satapornvanit K., Smidt H., Van den Brink P.J. (2014a). Effects of the antibiotic enrofloxacin on the ecology of tropical eutrophic freshwater microcosms. *Aquat. Toxicol.* 147:92–104.
- Rico A., Oliveira R., McDonough S., Matser A., Khatikarn J., Satapornvanit K., Nogueira A.J., Soares A.M.V.M. Domingues I., Van den Brink P.J., (2014b). Use, fate and ecological risks of antibiotics applied in tilapia cage farming in Thailand. *Environ. Pollut.* 191:8–16.
- Rico A., Waichman A.V., Geber-Correa R., Van den Brink P.J. (2011). Effects of malathion and carbendazim on amazonian freshwater organisms: comparison of tropical and temperate species sensitivity distributions. *Ecotoxicology* 20(4):625-634.
- Robinson A.A., Belden J.B., Lydy M.J. (2005). Toxicity of fluoroquinolone antibiotics to aquatic organisms. *Environ. Toxicol. Chem.* 24:423–430.
- Robinson C.D., Craft J.A., Moffat C.F., Davies I.M., Brown E.S., Megginson C. (2004). Oestrogenic markers and reduced population fertile egg production in a sand goby partial life-cycle test. *Mar. Environ. Res.* 58(2-5):147-150.
- Rose J., Holbech H., Lindholst C., Norum U., Povlsen A., Korsgaard B., Bjerregaard P. (2002). Vitellogenin induction by 17beta-estradiol and 17alpha-ethinylestradiol in male zebrafish (*Danio rerio*). *Comp. Biochem. Physiol. C Comp. Pharmacol. Toxicol.*



131(4):531-539.

- Rubach M.N., Crum S.J.H., Van den Brink P.J. (2011). Variability in the dynamics of mortality and immobility responses of freshwater arthropods exposed to chlorpyrifos. *Arch. Environ. Contam. Toxicol.* 60(4):708-721.
- Saglio P., Olsen K.H., Bretaud S. (2001). Behavioral and olfactory responses to prochloraz, bentazone, and nicosulfuron-contaminated flows in goldfish. *Arch. Environ. Contam. Toxicol.* 41(2):192-200.
- Salierno J.D. and Kane A.S. (2009). 17alpha-ethinylestradiol alters reproductive behaviors, circulating hormones, and sexual morphology in male fathead minnows (*Pimephales promelas*). *Environ. Toxicol. Chem.* 28(5):953-961.
- Sanchez W. and Porcher J.M. (2009). Fish biomarkers for environmental monitoring within the water framework directive of the European union. *Trends in Analytical Chemistry* 28(2):150-158.
- Sanchez-Arguello P., Aparicio N., Fernandez C. (2012). Linking embryo toxicity with genotoxic responses in the freshwater snail *Physa acuta*: single exposure to benzo(a)pyrene, fluoxetine, bisphenol A, vinclozolin and exposure to binary mixtures with benzo(a)pyrene. *Ecotoxicol. Environ. Saf.* 80:152-160.
- Sandhu N., McGeer J.C., Vijayan M.M. (2014). Exposure to environmental levels of waterborne cadmium impacts corticosteroidogenic and metabolic capacities, and compromises secondary stressor performance in rainbow trout. *Aquat. Toxicol.* 146:20-27.
- Saradhamani N. and Kumari S.B. (2011). Impact of insecticide (chlorpyrifos) on the biochemical components of the fish, *Oreochromis mossambicus* (Peters, 1852). *Curr. Biotica* 5(1):79-84.
- Sarma K., Pal A.K., Sahu N.P., Ayyappan S., Baruah K. (2009). Dietary high protein and vitamin C mitigates endosulfan toxicity in the spotted murrel, *Channa punctatus* (Bloch, 1793). *Sci. Total Environ.* 407(12):3668-3673.
- Satapornvanit K., Baird D.J., Little D.C. (2009). Laboratory toxicity test and post-exposure feeding inhibition using the giant freshwater prawn *Macrobrachium rosenbergii*. *Chemosphere* 74(9):1209-1215.
- Schirmer K and Knoebel M. (2012). CELLsens - unpublished data, provided for this review, EAWAG, Switzerland.
- Schoeny R., Cody T., Warshawsky D., Radike M. (1988). Metabolism of mutagenic polycyclic aromatic hydrocarbons by photosynthetic algal species. *Mutat. Res.* 197(2):289-302.
- Scholz S. (2012). Unpublished data provided for this review, UFZ-Helmholtz Center, Leipzig Germany.
- Schultz M.M., Bartell S.E., Schoenfuss H.L. (2012). Effects of triclosan and triclocarban, two ubiquitous environmental contaminants, on anatomy, physiology, and behavior of the fathead minnow (*Pimephales promelas*). *Arch. Environ. Contam. Toxicol.* 63(1):114-124.
- Schulz R.W., Bogerd J., Male R., Ball J., Fenske M., Olsen L.C., Tyler C.R. (2007). Estrogen-induced alterations in amh and dmrt1 expression signal for disruption in male sexual development in the zebrafish. *Environ. Sci. Technol.* 41(17):6305-6310.
- Seinen W., Helder T., Vernij H., Penninks A., Leeuwangh P. (1981). Short term toxicity of tri-n-butyltin chloride in rainbow trout (*Salmo gairdneri* Richardson) Yolk Sac Fry. *Sci. Total Environ.* 19(2):155-166.
- Selderslaghs I.W.T., Blust R., Witters H.E. (2012). Feasibility study of the zebrafish assay as an alternative method to screen for developmental toxicity and embryotoxicity using a training set of 27 compounds. *Reprod. Toxicol.* 33(2):142-154.
- Sellin M.K. and Kolok A.S. (2009). Cadmium exposures during early development: do they lead to reproductive impairment in fathead minnows?. *Environ. Toxicol. Chem.* 25(11):2957-2963.
- Seoane M., Rioboo C., Herrero C., Cid A. (2014). Toxicity induced by three antibiotics commonly used in aquaculture on the marine microalga *Tetraselmis suecica* (Kylin) Butch. *Mar. Environ. Res.* 101C:1-7.
- Shah S.L. and Altindag A. (2004). Hematological parameters of tench (*Tinca tinca* L.) after acute and chronic exposure to lethal and sublethal mercury treatments. *Bull. Environ. Contam. Toxicol.* 73(5):911-918.
- Shelley L.K., Ross P.S., Miller K.M., Kaukinen K.H., Kennedy C.J. (2012). Toxicity of atrazine and nonylphenol in juvenile rainbow trout (*Oncorhynchus mykiss*): effects on general health, disease susceptibility and gene expression. *Aquat. Toxicol.* 124/125:217-226.
- Shenoy K. (2012). Environmentally realistic exposure to the herbicide atrazine alters some sexually selected traits in male guppies. *PLoS One* 7(2):1-10.
- Sherrard R.M., Murray-Gulde C.L., Jr. Rodgers J.H., Shah Y.T. (2002). Comparative toxicity of chlorothalonil and chlorpyrifos: *Ceriodaphnia dubia* and *Pimephales promelas*. *Environ. Toxicol.* 17(6):503-512.
- Shugart L.R., D'Surney S.J., Gerrys-Hull G., Greeley M.S. (1991). Biological (molecular and cellular) markers of toxicity. ORNL/M-1426,

- Semi-Annual Technical Report No.5, U.S.Army Biomedical & Development Laboratories, Oct.1-March 30, 1991; Oak Ridge Natl. Lab., Oak Ridge, TN:65.
- Shuhaimi-Othman M., Nadzifah Y., Umirah N.S., Ahmad A.K. (2012). Toxicity of metals to an aquatic worm, *Nais elinguis* (Oligochaeta, Naididae). *Res. J. Environ. Toxicol.* 6(4):122-132.
- Shuhaimi-Othman M., Yakub N., Ramle N.A., Abas A. (2011a). Toxicity of metals to a freshwater ostracod: *Stenocypris major*. *J. Toxicol.* 2011:8p.
- Shuhaimi-Othman M., Yakub N., Umirah N.S., Abas A. (2011b). Toxicity of eight metals to Malaysian freshwater midge larvae *Chironomus javanus* (Diptera, Chironomidae). *Toxicol. Ind. Health* 27(10):879-886.
- Sieratowicz A., Stange D., Schulte-Oehlmann U., Oehlmann J. (2011). Reproductive toxicity of bisphenol A and cadmium in *Potamo-pyrgus antipodarum* and modulation of bisphenol A effects by different test temperature. *Environ. Pollut.* 159(10):2766-2774.
- Sies, H., 1985. Oxidative stress: introductory remarks. In *Oxidative Stress*, H. Sies, (Ed.) London: Academic Press Inc, (1985)
- Silva K.T.U. and Pathiratne A. (2008). In vitro and in vivo effects of cadmium on cholinesterases in Nile tilapia fingerlings: implications for biomonitoring aquatic pollution. *Ecotoxicology* 17(8):725-731.
- Sledge D., Yen J., Morton T., Dishaw L., Petro A., Donerly S., Linney E., Levin E.D. (2011). Critical duration of exposure for developmental chlorpyrifos-induced neurobehavioral toxicity. *Neurotoxicol. Teratol.* 33(6):742-751.
- Snel J.F.H., Vos J.H., Gylstra R., Brock T.C.M. (1998). Inhibition of photosystem ii (PSII) electron transport as a convenient endpoint to assess stress of the herbicide linuron on freshwater plants. *Aquat. Ecol.* 32(2): 113-123
- Snell T.W. (1991). New rotifer bioassays for aquatic toxicology. Final Report, U.S.Army Medical Research and Development Command, Fort Detrick, Frederick, MD:29 p.
- Soares S.S., Martins H., Gutierrez-Merino C., Aureliano M. (2008). Vanadium and cadmium in vivo effects in teleost cardiac muscle: metal accumulation and oxidative stress markers. *Comp. Biochem. Physiol. C Comp. Pharmacol. Toxicol.* 147(2): 168-178
- Sonneveld E., Jansen H.J., Riteco J.A.C., Brouwer A., van der Burg B. (2005). Development of androgen- and estrogen-responsive bioassays, members of a panel of human cell line-based highly selective steroid-responsive bioassays. *Toxicol Sci* 83:136-48.
- Sotomayor V., Lascano C., De D'Angelo A.M.P., Venturino A. (2012). developmental and polyamine metabolism alterations in *Rhinella arenarum* embryos exposed to the organophosphate chlorpyrifos. *Environ. Toxicol. Chem.* 31(9):2052-2058.
- Sparling D.W. and Fellers G.M. (2009). Toxicity of two insecticides to California, USA, anurans and its relevance to declining amphibian populations. *Environ. Toxicol. Chem.* 28(8):1696-1703.
- Spitsbergen J.M., Walker M.K., Olson J.R., Peterson R.E. (1991). Pathologic alterations in early life stages of lake trout, *Salvelinus namaycush*, exposed to 2,3,7,8-tetrachlorodibenzo-p-dioxin as fertilized eggs. *Aquat. Toxicol.* 19(1):41-72.
- Srivastav A.K., Srivastava S.K., Mishra D., Srivastav S.K. (2011). Histological alterations in the ultimobranchial gland of teleost *Heteropneustes fossilis* in response to chlorpyrifos treatment. *J. Basic Clin. Physiol. Pharmacol.* 22(1/2):23-28.
- Stara A., Machova J., Velisek J. (2012). Effect of chronic exposure to simazine on oxidative stress and antioxidant response in common carp (*Cyprinus carpio* L.). *Environ. Toxicol. Pharmacol.* 33(2):334-343.
- Stephenson R.R. and Kane D.F. (1984). Persistence and effects of chemicals in small enclosures in ponds. *Arch. Environ. Contam. Toxicol.* 13(3):313-326.
- Stikker A. (1998). Water today and tomorrow. *Futures* 30(1):43-62.
- STOWA, 2016a. Leo Posthuma, Dick de Zwart, Leonard Osté, Ron van der Oost en Jaap Postma. Ecologische Sleutelfactore Toxiciteit: deel 1 hoofdrapport: Methode voor het in beeld brengen van de effecten van giftige stoffen in het oppervlaktewater. 2016-15A. STOWA - Stichting Toegepast Onderzoek Waterbeheer, Amersfoort.
- STOWA, 2016b. Ron van der Oost, Mai Thao Nguyen. Ecologische sleutelfactor toxiciteit. Deel 4: SIMONI procedures voor effectgerichte monitoring. 2016 - 15D. STOWA - Stichting Toegepast Onderzoek Waterbeheer, Amersfoort.
- Stromqvist M., Tooke N., Brunstrom B. (2010). DNA methylation levels in the 5' flanking region of the vitellogenin i gene in liver and brain of adult zebrafish (*Danio rerio*) - sex and tissue differences and effects of 17alpha-ethinylestradiol exposure. *Aquat. Toxicol.* 98(3):275-281.
- Sun L., Xu W., He J., Yin Z. (2010a). In vivo alternative assessment of the chemicals that interfere with anterior pituitary POMC expression and interrenal steroidogenesis in POMC: EGFP transgenic zebrafish. *Toxicol. Appl. Pharmacol.* 248(3):217-225.
- Sun L., Xu W., He J., Yin Z. (2010b). In vivo alternative assessment of the chemicals that interfere with anterior pituitary pomc ex-

- pression and interrenal steroidogenesis in POMC: EGFP transgenic zebrafish. *Toxicol. Appl. Pharmacol.* 248(3):217-225.
- Sun L., Zha J., Wang Z. (2009). Interactions between estrogenic chemicals in binary mixtures investigated using vitellogenin induction and factorial analysis. *Chemosphere* 75:410-415.
- Swapna I. and Senthikumar B. (2009). Influence of ethynylestradiol and methyltestosterone on the hypothalamo-hypophyseal-gonadal axis of adult air-breathing catfish, *Clarias gariepinus*. *Aquat. Toxicol.* 95(3):222-229.
- Sztrum A.A., D'Eramo J.L., Herkovits J. (2011). Nickel toxicity in embryos and larvae of the south american toad: effects on cell differentiation, morphogenesis, and oxygen consumption. *Environ. Toxicol. Chem.* 30(5):1146-1152.
- Tabata A., Kashiwada S., Ohnishi Y., Ishikawa H., Miyamoto N., Itoh M., Magara Y. (2001). Estrogenic influences of estradiol-17beta, p-nonylphenol and bis-phenol-A on japanese medaka (*Oryzias latipes*) at detected environmental concentrations. *Water Sci. Technol.* 43(2):109-116.
- Tan Q.G. and Wang W.X. (2011). Acute toxicity of cadmium in *Daphnia magna* under different calcium and pH conditions: importance of influx rate. *Environ. Sci. Technol.* 45(5):1970-1976.
- Tatarazako N., Ishibashi H., Teshima K., Kishi K., Arizono K. (2004). Effects of triclosan on various aquatic organisms. *Environ. Sci.* 11(2):133-140.
- Taylor N.S., Weber R.J.M., White T.A., Viant M.R. (2010). Discriminating between different acute chemical toxicities via changes in the daphnid metabolome. *Toxicol. Sci.* 118(1):307-317.
- Teixido E., Pique E., Gomez-Catalan J., Llobet J.M. (2013). Assessment of developmental delay in the zebrafish embryo teratogenicity assay. *Toxicol. In Vitro* 27(1):469-478.
- Tetreau G., Chandor-Proust A., Faucon F., Stalinski R., Akhouayri I., Prud'homme S.M., Regent Kloeckner M., Raveton M., Reynaud S. (2014). UV light and urban pollution: bad cocktail for mosquitoes?. *Aquat. Toxicol.* 146:52-60.
- Thomas V.M. Jr., Buckley L.J., Sullivan Jr.J.D., Ikawa M. (1973). Effect of herbicides on the growth of chlorella and bacillus using the paper disc method. *Weed Sci.* 21(5):449-451.
- Thompson S., Tilton F., Schlenk D., Benson W.H. (2000). Comparative vitellogenic responses in three teleost species: extrapolation to in situ field studies. *Mar. Environ. Res.* 51(1-5):185-189.
- Thompson S.C. (2000). Physiological indicators of endocrine disruptor exposure in japanese medaka (*Oryzias latipes*): relationship to reproduction and development. M.S.Thesis, University of Mississippi, Oxford, MS:122p.
- Thorpe K.L., Benstead R., Hutchinson T.H., Tyler C.R. (2007). Associations between altered vitellogenin concentrations and adverse health effects in fathead minnow (*Pimephales promelas*). *Aquat. Toxicol.* 85(3):176-183.
- Thorpe K.L., Cummings R.I., Hutchinson T.H., Scholze M., Brighty G., Sumpter J.P., Tyler C.R. (2003). Relative potencies and combination effects of steroidal estrogens in fish. *Environ. Sci. Technol.* 37(6):1142-1149.
- Thorpe K.L., Hutchinson T.H., Hetheridge J.P., Sumpter J.P., Tyler C.R. (2000). Development of an in vivo screening assay for estrogenic chemicals using juvenile rainbow trout (*Oncorhynchus mykiss*). *Environ. Toxicol. Chem.* 19(11):2812-2820.
- Tierney K.B., Ross P.S., Jarrard H.E., Delaney K.R., Kennedy C.J. (2006). Changes in juvenile coho salmon electro-olfactogram during and after short-term exposure to current-use pesticides. *Environ. Toxicol. Chem.* 25(10):2809-2817.
- Tillmann M., Schulte-Oehlmann U., Duft M., Markert B., Oehlmann J. (2001a). Effects of endocrine disruptors on prosobranch snails (Mollusca: Gastropod) in the laboratory. Part III: cyproterone acetate and vinclozolin as antiandrogens. *Ecotoxicology* 10(6):373-388.
- Tilton F.A., Bammler T.K., Gallagher E.P. (2011b). Swimming impairment and acetylcholinesterase inhibition in zebrafish exposed to copper or chlorpyrifos separately, or as mixtures. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* 153(1):9-16.
- Tilton F.A., Tilton S.C., Bammler T.K., Beyer R.P., Stapleton P.L., Scholz N.L., Gallagher E.P. (2011a). Transcriptional impact of organophosphate and metal mixtures on olfaction: copper dominates the chlorpyrifos-induced response in adult zebrafish. *Aquat. Toxicol.* 102(3/4):205-215.
- Tilton S.C., Foran C.M., Benson W.H. (2004). Effects of cadmium on the reproductive axis of japanese medaka (*Oryzias latipes*). *Comp. Biochem. Physiol. C Comp. Pharmacol. Toxicol.* 136(3):265-276.
- Tilton S.C., Foran C.M., Benson W.H. (2005). Relationship between ethynylestradiol-mediated changes in endocrine function and reproductive impairment in japanese medaka (*Oryzias latipes*). *Environ. Toxicol. Chem.* 24(2):352-359.
- To T.T., Hahner S., Nica G., Rohr K.B., Hammerschmidt M., Winkler C., Bruno Allolio B. (2007). Pituitary-interrenal interaction in



- zebrafish interrenal organ development. *Molecular Endocrinology* 21(2):472-485.
- Toomey B.H., Bello S., Hahn M.E., Cantrell S., Wright P., Tillitt D.E., Di Giulio R.T. (2001). 2,3,7,8-tetrachlorodibenzo-p-dioxin induces apoptotic cell death and cytochrome P4501A expression in developing fundulus heteroclitus embryos. *Aquat. Toxicol.* 53(2):127-138.
- Tremblay L. and Van der Kraak G. (1999). Comparison between the effects of the phytosterol beta-sitosterol and pulp and paper mill effluents on sexually immature rainbow trout. *Environ. Toxicol. Chem.* 18(2):329-336.
- Tripathi G. and Shasmal J. (2010). Reparation of chlorpyrifos-induced impairment by thyroxine and vitamin C in fish. *Ecotoxicol. Environ. Saf.* 73(6):1397-1401.
- Tripathi G. and Shasmal J. (2011). Concentration related responses of chlorpyrifos in antioxidant, anaerobic and protein synthesizing machinery of the freshwater fish, *Heteropneustes fossilis*. *Pestic. Biochem. Physiol.* 99(3):215-220.
- Trucco R.G., Engelhardt F.R., Stacey B. (1983). Toxicity, accumulation and clearance of aromatic hydrocarbons in *Daphnia pulex*. *Environ. Pollut. A.* 31(3):191-202.
- Tsuji S., Tonogai Y., Ito Y., Kanoh S. (1986). The influence of rearing temperatures on the toxicity of various environmental pollutants for killifish (*Oryzias latipes*). *Eisei Kagaku (Jpn. J. Toxicol. Environ. Health)* 32(1):46-53.
- U.S. Environmental Protection Agency, and Office of Pesticide Programs (2013). Pesticide Ecotoxicity Database (Formerly: Environmental Effects Database (EEDB)). Environmental Fate and Effects Division, U.S.EPA, Washington, D.C.
- U.S. EPA, Office of Solid Waste and Emergency Response, (1996) Ecotox Thresholds. Publication 9345.0-12FSI, EPA 540/F-95/038. PB95-963324.
- Valenti T.W., Cherry D.S., Neves R.J., Locke B.A., Schmerfeld J.J. (2006). Case study: sensitivity of mussel glochidia and regulatory test organisms to mercury and a reference toxicant. In: J.L.Farris and J.H.Van Hassel (Eds.), *Freshwater Bivalve Ecotoxicology, SETAC, Pensacola, FL*:351-367.
- Van den Belt K., Berckmans P., Vangenechten C., Verheyen R., Witters H. (2004). Comparative Study on the In Vitro/In Vivo Estrogenic Potencies of 17beta-Estradiol, Estrone, 17alpha-Ethinylestradiol and Nonylphenol. *Aquat. Toxicol.* 66(2):183-195.
- Van den Brink P.J., Hattink J., Bransen F., Van Donk E., Brock T.C.M. (2000). Impact of the fungicide carbendazim in freshwater microcosms. II. zooplankton, primary producers and final conclusions. *Aquat. Toxicol.* 48(2-3):251-264.
- Van der Grinten E., Pikkemaat M.G., van den Brandhof E.J., Stroomberg G.J., Kraak M.H.S. (2010). Comparing the sensitivity of algal, cyanobacterial and bacterial bioassays to different groups of antibiotics. *Chemosphere* 80:1-6.
- Van der Oost, R., G. Sileno, M. Suarez Muños, M. T. Nguyen, H. Besselink and A. Brouwer. in preparation a. SIMONI as a novel bio-analytical monitoring strategy for surface water quality assessment; Part I: model design with effect-based trigger values
- Van der Oost, R., G. Sileno, M. Suarez Muños, M. T. Nguyen, H. Besselink and A. Brouwer. in preparation b. SIMONI as a novel bio-analytical monitoring strategy for surface water quality assessment; Part II: feasibility of the model in field surveys
- Van Wijngaarden R.P.A., Van den Brink P.J., Oude Voshaar J.H., Leeuwangh P. (1995). Ordination techniques for analysing response of biological communities to toxic stress in experimental ecosystems. *Ecotoxicology* 4(1):61-77.
- Vandenbrouck T., Soetaert A., Van der Ven K., Blust R., De Coen W. (2009). Nickel and binary metal mixture responses in *Daphnia magna*: molecular fingerprints and (sub)organismal effects. *Aquat. Toxicol.* 92(1):18-29.
- Vasquez M.I., Violaris M., Hadjivassilis A., Wirth M.C. (2009). Susceptibility of *Culex pipiens* (Diptera: Culicidae) field populations in cyprus to conventional organic insecticides, *Bacillus thuringiensis* subsp. *israelensis*, and methoprene. *J. Med. Entomol.* 46(4):881-887.
- Velisek J., Stara A., Machova J., Dvorak P., Zuskova E., Svobodova Z. (2012b). Effects of low-concentrations of simazine on early life stages of common carp (*Cyprinus carpio* L.). *Neuroendocrinol. Lett.* 33(3):90-95.
- Velisek J., Stara A., Machova J., Svobodova Z. (2012a). Effects of long-term exposure to simazine in real concentrations on common carp (*Cyprinus carpio* L.). *Ecotoxicol. Environ. Saf.* 76(1):79-86.
- Vellinger C., Gismondì E., Felten V., Rousselle P., Mehennaoui K., Parant M., Usseglio-Polatera P. (2013). Single and combined effects of cadmium and arsenate in *Gammarus pulex* (Crustacea, Amphipoda): understanding the links between physiological and behavioural responses. *Aquat. Toxicol.* 140/141:106-116.
- Verma S.R., Bansal S.K., Gupta A.K., Pal N., Tyagi A.K., Bhatnagar M.C., Kumar V., Dalela R.C. (1982). Bioassay trials with twenty-three pesticides to a fresh water teleost, *Saccobranhus fossilis*. *Water Res.* 16(5):525-529.

- Verma S.R., Bhatnagar M.C., Dalela R.C. (1978b). Biocides in relation to water pollution. Part 2: Bioassay studies of few biocides to a fresh water fish, *Channa gachua*. *Acta Hydrochim. Hydrobiol.* 6(2):137-144.
- Verslycke T., Poelmans S., De Wasch K., De Brabander H.F., Janssen C.R. (2004). Testosterone and energy metabolism in the estuarine mysid *Neomysis integer* (Crustacea: Mysidacea) following exposure to endocrine disruptors. *Environ. Toxicol. Chem.* 23(5):1289-1296.
- Vervliet-Scheebaum M., Straus A., Tremp H., Hamer M., Maund S.J., Wagner E., Schulz R. (1993). Field corn, wireworm larval control, 1992. *Environ. Pollut.* 18:208-209.
- Vighi M. and Calamari D. (1985). QSARs for organotin compounds on *Daphnia magna*. *Chemosphere* 14(11-12):1925-1932.
- Villeneuve D.L., Blake L.S., Brodin J.D., Greene K.J., Knoebel I., Miracle A.L., Martinovic D., Ankley G.T. (2007). Transcription of key genes regulating gonadal steroidogenesis in control and ketoconazole- or vinclozolin exposed fathead minnows. *Toxicol. Sci.* 98(2):395-407.
- Vlaardingen P.L.A., Traas T.P., Wintersen A.M., Aldenberg T. (2004). ETX 2.0 – A Program to Calculate Hazardous Concentrations and Fraction Affected, Based on Normally Distributed Toxicity Data. RIVM (National Institute for Public Health and the Environment), report 601501028/2004
- Walker M.K., Spitsbergen J.M., Olson J.R., Peterson R.E. (1991). 2,3,7,8-Tetrachlorodibenzo-p-dioxin (TCDD) toxicity during early life stage development of lake trout (*Salvelinus namaycush*) *Can. J. Fish. Aquat. Sci.* 48(5):875-883.
- Wang C., Lu G., Cui J. (2012). Responses of AChE and GST activities to insecticide coexposure in *Carassius auratus*. *Environ. Toxicol.* 27(1):50-57.
- Wang H.Y., Olmstead A.W., Li H., LeBlanc G.A. (2005). The screening of chemicals for juvenoid-related endocrine activity using the water flea *Daphnia magna*. *Aquat. Toxicol.* 74(3):193-204.
- Ward A.J.W., Duff A.J., Currie S. (2006). The effects of the endocrine disrupter 4-nonylphenol on the behaviour of juvenile rainbow trout (*Oncorhynchus mykiss*). *Can. J. Fish. Aquat. Sci.* 63(2):377-382.
- Warshawsky D., Cody T., Radike M., Reilman R., Schumann B., LaDow K., Schneider J. (1995). Biotransformation of benzo(a)pyrene and other polycyclic aromatic hydrocarbons and heterocyclic analogs by several green algae and other algal species. *Chem.-Biol. Interact.* 97(2):131-148.
- Weigt S., Huebler N., Strecker R., Braunbeck T., Broschard T.H. (2011). Zebrafish (*Danio rerio*) embryos as a model for testing proteratogens. *Toxicology* 281(1-3):25-36.
- Weinstein J.E. and Garner T.R. (2008). Piperonyl butoxide enhances the bioconcentration and photoinduced toxicity of fluoranthene and benzo(a)pyrene to larvae of the grass shrimp (*Palaemonetes pugio*). *Aquat. Toxicol.* 87(1):28-36.
- Weinstein J.E. and Polk K.D. (2001). phototoxicity of anthracene and pyrene to glochidia of the freshwater mussel *Utterbackia imbecillis*. *Environ. Toxicol. Chem.* 20(9):2021-2028.
- Weisbrod C.J., Kunz P.Y., Zenker A.K., Fent K. (2007). Effects of the UV filter benzophenone-2 on reproduction in fish. *Toxicol. Appl. Pharmacol.* 225(3):255-266.
- Wenzel A., Schafers C., Vollmer G., Michna H., Diel P. (2001). Research efforts towards the development and validation of a test method for the identification of endocrine disrupting chemicals. Final Rep., Contract B6-7920/98/000015, Fraunhofer-Inst.fur Umweltchemie und Okotoxikologie, Schmallenberg, Germany:81p.
- Werner J. (2006). Development of methods to assess metallothionein expression in lake trout (*Salvelinus namaycush*) during a reproductive cycle and the effects of cadmium and ethynylestradiol. Ph.D Thesis, University of Manitoba, Canada:194p.
- Wernersson A.S. and Dave G. (1997). Phototoxicity identification by solid phase extraction and photoinduced toxicity to *Daphnia magna*. *Arch. Environ. Contam. Toxicol.* 32(3):268-273.
- Wernersson A.S., Maggi C., Carere M. (2014) Technical report on aquatic effect-based monitoring tools. Technical Report 2014-077. Luxembourg: Office for Official. Publications of the European Communities.
- Wester P.W. and Canton J.H. (1991). The usefulness of histopathology in aquatic toxicity studies. *Comp. Biochem. Physiol. C Comp. Pharmacol.* 100(1-2):115-117.
- Wijesinghe M.R., Bandara M.G., Ratnasooriya W.D., Lakraj G.P. (2011). Chlorpyrifos-induced toxicity in *Duttaphrynus melanostictus* (Schneider 1799) larvae. *Arch. Environ. Contam. Toxicol.* 60(4):690-696.
- Williams C.R. and Gallagher E.P. (2013). Effects of cadmium on olfactory mediated behaviors and molecular biomarkers in coho salmon

- (*Oncorhynchus kisutch*). *Aquat. Toxicol.* 140/141:295-302.
- Wirth M.C. (1998). Isolation and characterization of two novel organophosphate resistance mechanisms in *Culex pipiens* from Cyprus. *J. Am. Mosq. Control Assoc.* 14(4):397-405.
- Wirth M.C. and Georghiou G.P. (1996). Organophosphate resistance in *Culex pipiens* from Cyprus. *J. Am. Mosq. Control Assoc.* 12(1):112-118.
- Wisk J.D. and Cooper K.R. (1990a). Comparison of the toxicity of several polychlorinated dibenzo-p-dioxins and 2,3,7,8-tetrachlorodibenzofuran in embryos of the Japanese Medaka (*Oryzias latipes*). *Chemosphere* 20(3-4):361-377.
- Wisk J.D. and Cooper K.R. (1990b). The stage specific toxicity of 2,3,7,8-tetrachlorodibenzo-p-dioxin in embryos of the Japanese medaka (*Oryzias latipes*). *Environmental Toxicology and Chemistry* 9(9):1159-1169.
- Wisk J.D. and K.R. Cooper (1992). Effect of 2,3,7,8-tetrachlorodibenzo-p-dioxin on benzo(a)pyrene hydroxylase activity in embryos of the Japanese medaka (*Oryzias latipes*). *Arch. Toxicol.* 66(4):245-249.
- Wollenberger L., Halling-Sorensen B., Kusk K.O. (2000). Acute and chronic toxicity of veterinary antibiotics to *Daphnia magna*. *Chemosphere* 40:723-730.
- Wolska L., Sagajdakow A., Kuczynska A., Namiesnik J. (2007). Application of ecotoxicological studies in integrated environmental monitoring: Possibilities and problems. *Trends in Analytical Chemistry* 26(4):332-344.
- Wood R.J., Pasteur N., Sinigre G. (1984). Carbamate and organophosphate resistance in *Culex pipiens* L. (Diptera: Culicidae) in southern France and the significance of Est-3A. *Bull. Entomol. Res.* 74(4):677-687.
- Woods M., Kumar A., Correll R. (2002). Acute toxicity of mixtures of chlorpyrifos, profenofos, and endosulfan to *Ceriodaphnia dubia*. *Bull. Environ. Contam. Toxicol.* 68(6):801-808.
- Woods R., Davi R., Arnold W. (2004). Toxicity of vanadium to the estuarine mysid, *Americamysis bahia* (Molenock) (formerly *Mysidopsis bahia*). *Bull. Environ. Contam. Toxicol.* 73(4):635-643.
- Wu W.Z., Li W., Xu Y., Wang J.W. (2001). Long-term toxic impact of 2,3,7,8-tetrachlorodibenzo-p-dioxin on the reproduction, sexual differentiation, and development of different life stages of *Gobiocypris rarus* and *Daphnia magna*. *Ecotoxicol. Environ. Saf.* 48(3):293-300.
- Xie L., Flippin J.L., Deighton N., Funk D.H., Dickey D.A., Buchwalter D.B. (2009). Mercury(II) bioaccumulation and antioxidant physiology in four aquatic insects. *Environ. Sci. Technol.* 43(3):934-940.
- Xing H., Li S., Wang Z., Gao X., Xu S., Wang X. (2012a). Histopathological changes and antioxidant response in brain and kidney of common carp exposed to atrazine and chlorpyrifos. *Chemosphere* 88(4):377-383.
- Xing H., Li S., Wang Z., Gao X., Xu S., Wang X. (2012b). Oxidative stress response and histopathological changes due to atrazine and chlorpyrifos exposure in common carp. *Pestic. Biochem. Physiol.* 103(1):74-80.
- Xu H., Yang J., Wang Y., Jiang Q., Chen H., Song H. (2008). Exposure to 17 $\alpha$ -ethynylestradiol impairs reproductive functions of both male and female zebrafish (*Danio rerio*). *Aquat. Toxicol.* 88(1):1-8.
- Yang J.-F., Ying G.G., Zhao J.L., Tao R., Su H.C., Liu Y.S. (2011). Spatial and seasonal distribution of selected antibiotics in surface waters of the Pearl Rivers, China. *J. Environ. Sci. Health. B.* 46:272-80.
- Yang L.H., Ying G.G., Su H.C., Stauber J.L., Adams M.S., Binet M.T. (2008). Growth-inhibiting effects of 12 antibacterial agents and their mixtures on the freshwater microalga *Pseudokirchneriella subcapitata*. *Environ. Toxicol. Chem.* 27(5):1201-1208.
- Yoon C.S., Jin J.H., Park J.H., Yeo C.Y., Kim S.J., Hwang Y.G., Hong S.J., Cheong S.W. (2008). Toxic effects of carbendazim and n-butyl isocyanate, metabolites of the fungicide benomyl, on early development in the African clawed frog, *Xenopus laevis*. *Environ. Toxicol.* 23(1): 131-144.
- Young W.F., Whitehouse P., Johnson I., Sorokin N. (2004). Proposed Predicted-No-Effect-Concentrations (PNECs) for Natural and Synthetic Steroid Oestrogens in Surface Waters. R&D Technical Report P2-T04/1
- Yuan X., Lia T., Zhou L., Zhao X. (2014). Characteristics and risk assessment of estrogenic compounds in rivers of southern Jiangsu province, China. *IERI Procedia* 9:176-184
- Zavala-Aguirre J.L., Torres-Bugarin O., Zamora-Perez A.L. (2007). Aquatic ecotoxicology approaches in western Mexico. *Journal of Environmental Science and Health, Part A: Toxic/Hazardous Substances and Environmental Engineering* 42(10):1503-1511.
- Zegura B., Heath E., Cernoša A., Filipic M. (2009). Combination of in vitro bioassays for the determination of cytotoxic and genotoxic potential of wastewater, surface water and drinking water samples. *Chemosphere* 75:1453-1460.

- Zhang X., Hecker M., Park J.W., Tompsett A.R., Newsted J., Nakayama K., Jones P.D., Au D., Kong R., Wu R.S., Giesy J.P. (2008). Real-time PCR array to study effects of chemicals on the hypothalamic-pituitary-gonadal axis of the japanese medaka. *Aquat. Toxicol.* 88(3):173-182.
- Zou E. and Fingerman M. (1997). Synthetic estrogenic agents do not interfere with sex differentiation but do inhibit molting of the cladoceran *Daphnia magna*. *Bull. Environ. Contam. Toxicol.* 58(4):596-602.
- Holbech H., Kinnberg K., Petersen G.I., Jackson P., Hylland K., Norrgren L., Bjerregaard P. (2006). Detection of endocrine disrupters: evaluation of a fish sexual development test (FSDT). *Comp. Biochem. Physiol. C Comp. Pharmacol. Toxicol.* 144(1):57-66.

# APPENDIX SI-I

Relative effect potencies (REP) of all compounds investigated FOR EBT development in all bioassays

For antibiotics, the REPs were calculated using detection limits of RIKILT WaterScan method, except for \*: RIKILT MeatScan method; and \*\*: NDKT method.

For antibiotics, the REPs were calculated using detection limits of RIKILT WaterScan method, except for \*: RIKILT MeatScan method; and \*\*: NDKT method.

CAS number	Compound	Erb		antiAR		GR		DR		PPARY		PAH		NfZ		PXR		Macrolides & β-Lactams		Antibiotic activity		Sulfonamides		Tetracyclines	
		CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX	CALUX
57-63-6	17α-Ethinyl estradiol	1.560	50.119																						
50-28-2	17β-estradiol	1.000	31.623												0.063										
1689-82-3	4-hydroxazobenzene			0.010	501.187																				
14938-35-3	4-n-amylophenol																								
1806-26-4	4-n-octylphenol																								
140-66-9	4-tert-octylphenol																								
15-972-60-8	Alachlor																								
52-39-1	Aldosterone							0.008																	
1263-89-4	Amnosidine																								
2678778-0	Amoxicillin																								
69-53-4	Ampicillin																								
120-12-7	Anthracene																								
1912-24-9	Atrazine																								
1405-87-4	Bacitracin																								
25057-89-0	Bentazon																								
71-43-2	Benzene																								
50-32-8	Benzo(a)pyrene																								
207-08-9	Benzo(k)fluoranthene																								
61-33-6	Benzylpenicillin								0.005																
80-05-7	Bisphenol-A																								
85-68-7	Butyl benzyl phthalate																								
10108-64-2	Cadmium chloride																								
133-06-2	Captan																								
68-25-2	Carbaryl																								
10605-21-7	Carbendazim																								
56-75-7	Chloramphenicol																								
2921-88-2	Chlorpyrifos-ethyl																								
57-62-5	Chlortetracycline																								
85721-33-1	Ciprofloxacin																								
50-22-6	Corticosterone																								
2242-98-0	Cortisol																								
6055-19-2	Cyclophosphamide																								
50-02-2	Dexamethasone																								
53-70-3	Dibenz(a,h)anthracene																								
84-74-2	Dibutylphthalate																								
683-18-1	Dibutyltin dichloride																								
115-32-2	Dicofol																								
56-53-1	Diethylstilbestrol																								
128-46-1	Dihydrostreptomycin																								
330-54-1	Duron																								
564-25-0	Doxycycline																								
115-29-7	Endosulfan																								
93106-60-6	Enrofloxacin																								
114-07-8	Erythromycin																								
50-27-1	Estril																								
53-16-7	Estrone																								



# APPENDIX SI-II

## Toxicological data considered for EBT design of bioassays

### A: ER CALUX

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference	
57-63-6	17a-Ethinyl estradiol	Algae	<i>Scenedesmus subspicatus</i>	EC50	840000	131323	3	Kopf, 1997
			Amphibia	<i>Lithobates clamitans ssp. Clamitans</i>	NOEC	5.80	9.07	119
		5.80			9.07	119	Park, 2003	
		6.10	9.54	25	Park, 2003			
		6.10	9.54	105	Park and Kidd, 2005			
		6.10	9.54	105	Park and Kidd, 2005			
		6.10	9.54	105	Park and Kidd, 2005			
		10.03	15.68	189	Park, 2003			
		1000	1563	189	Park, 2003			
		LOEC	5.80	9.07	119	Park, 2003		
		6.10	9.54	105	Park and Kidd, 2005			
		6.10	9.54	105	Park and Kidd, 2005			
		82.50	129	189	Park, 2003			
		<i>Lithobates septentrionalis</i>	NOEC	5.00	7.82	119	Park and Kidd, 2005	
			5.00	7.82	119	Park and Kidd, 2005		
			5.00	7.82	119	Park and Kidd, 2005		
			5.00	7.82	119	Park and Kidd, 2005		
			NOEC	115	180	124	Brande-Lavridsen et al., (2010)	
			115	180	124	Brande-Lavridsen et al., (2010)		
			115	180	124	Brande-Lavridsen et al., (2010)		
			115	180	124	Brande-Lavridsen et al., (2010)		
			115	180	124	Brande-Lavridsen et al., (2010)		
			LOEC	6.00	9.38	124	Brande-Lavridsen et al., (2010)	
		<i>Rana temporaria</i>	NOEC	165	2.58	61	Gyllenhammar et al., 2009	
			165	2.58	61	Gyllenhammar et al., 2009		
			17.50	27.36	61	Gyllenhammar et al., 2009		
			180	281	61	Gyllenhammar et al., 2009		
			180	281	61	Gyllenhammar et al., 2009		
			180	281	61	Gyllenhammar et al., 2009		
			180	281	61	Gyllenhammar et al., 2009		
			180	281	61	Gyllenhammar et al., 2009		
			180	281	61	Gyllenhammar et al., 2009		
			180	281	61	Gyllenhammar et al., 2009		
		<i>Xenopus tropicalis</i>	NOEC	500000	781684	7	Jukosky et al., 2008a	
			500000	781684	7	Jukosky et al., 2008a		
			EC50	935500	1462530	7	Cho, 2005	
			NOEC	0.10	0.16	7	Dietrich et al., 2010	
			0.10	0.16	7	Dietrich et al., 2010		
			10000	15634	21	Kopf, 1997		
			500000	781684	21	Clubbs and Brooks, 2007		
			500000	781684	21	Clubbs and Brooks, 2007		
			1000000	1563367	21	Clubbs and Brooks, 2007		
			1000000	1563367	21	Clubbs and Brooks, 2007		
		1000000	1563367	21	Clubbs and Brooks, 2007			
		1000000	1563367	21	Clubbs and Brooks, 2007			
		LOEC	0.10	0.16	7	Dietrich et al., 2010		
		0.10	0.16	7	Dietrich et al., 2010			
		62500	97710	21	Clubbs and Brooks, 2007			
		1000000	1563367	21	Clubbs and Brooks, 2007			
		1000000	1563367	21	Clubbs and Brooks, 2007			
EC50	105000	164154	21	Kopf, 1997				
2590400	4049746	4	Clubbs and Brooks, 2007					
5700000	891119	1	Kopf, 1997					
<i>Hyalella azteca</i>	NOEC	70000	109436	63	Dussault et al., 2008b			
	70000	109436	63	Dussault et al., 2008b				
	70000	109436	63	Dussault et al., 2008b				
	70000	109436	63	Dussault et al., 2008b				
	740000	1156892	63	Dussault et al., 2008b				
	910000	1422664	21	Dussault et al., 2009				
	LOEC	740000	1156892	63	Dussault et al., 2008b			
	740000	1156892	63	Dussault et al., 2008b				
	740000	1156892	63	Dussault et al., 2008b				
	740000	1156892	63	Dussault et al., 2008b				
Crustacea	<i>Ceriodaphnia dubia</i>	NOEC	500000	781684	7	Jukosky et al., 2008a		
		500000	781684	7	Jukosky et al., 2008a			
	<i>Daphnia magna</i>	EC50	935500	1462530	7	Cho, 2005		
		NOEC	0.10	0.16	7	Dietrich et al., 2010		
	0.10	0.16	7	Dietrich et al., 2010				
	10000	15634	21	Kopf, 1997				
	500000	781684	21	Clubbs and Brooks, 2007				
	500000	781684	21	Clubbs and Brooks, 2007				
	1000000	1563367	21	Clubbs and Brooks, 2007				
	1000000	1563367	21	Clubbs and Brooks, 2007				
1000000	1563367	21	Clubbs and Brooks, 2007					
1000000	1563367	21	Clubbs and Brooks, 2007					
Fish	<i>Catostomus commersoni</i>	LC50	1100000	1719704	10	Dussault et al., 2008a		
		NOEC	5.45	8.52	1095	Palace et al., 2009		
		5.55	8.68	1095	Palace et al., 2009			
		6.10	9.54	1095	Palace et al., 2009			
		LOEC	5.45	8.52	1095	Palace et al., 2009		
		5.45	8.52	1095	Palace et al., 2009			
		5.55	8.68	1095	Palace et al., 2009			
		6.10	9.54	1095	Palace et al., 2009			
		6.10	9.54	1095	Palace et al., 2009			
		6.10	9.54	1095	Palace et al., 2009			
<i>Clarias gariepinus</i>	NOEC	1000	1563	21	Swapna and Senthilkumar, 2009			
	LOEC	0.65	1.02	7	Braathen et al., 2009			
	1000	1563	21	Swapna and Senthilkumar, 2009				
	1000	1563	21	Swapna and Senthilkumar, 2009				
<i>Cyprinus carpio</i>	NOEC	1000	1563	21	Swapna and Senthilkumar, 2009			
	1000	1563	21	Swapna and Senthilkumar, 2009				
	1.00	1.56	10	Purdom et al., 1994				
	3.00	4.69	7	Lange et al., 2012				
9.30	14.54	7	Lange et al., 2012					



CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference		
57-63-6	17 $\alpha$ -Ethinyl estradiol	Fish	<i>Cyprinus carpio</i>	LOEC	1.70	2.66	7	Lange et al., 2012	
					10.00	15.63	10	Purdom et al., 1994	
					5000	78.17	30	Ebrahimi, 2007	
				<i>Danio rerio</i>	NOEC	0.050	0.078	56	Schulz et al., 2007
						0.075	0.12	322	Wenzel et al., 2001
						0.30	0.47	322	Wenzel et al., 2001
						0.30	0.47	322	Wenzel et al., 2001
						0.40	0.63	88	Xu et al., 2008
						0.40	0.63	88	Xu et al., 2008
					LOEC	0.40	0.63	88	Xu et al., 2008
						0.50	0.078	2	Colman et al., 2009
						0.50	0.78	56	Schulz et al., 2007
						0.50	0.78	56	Schulz et al., 2007
						0.50	0.78	56	Schulz et al., 2007
						1.55	2.42	322	Wenzel et al., 2001
			1.55	2.42		322	Wenzel et al., 2001		
			1.90	2.97		7	Lange et al., 2012		
			2.00	3.13		88	Xu et al., 2008		
			2.90	4.53		7	Lange et al., 2012		
			5.00	7.82		14	Reyhanian et al., 2011		
			5.00	7.82		56	Schulz et al., 2007		
			5.00	7.82		56	Schulz et al., 2007		
			5.58	8.72		122	Larsen et al., 2009		
			5.80	0.91		2	Colman et al., 2009		
			Fish	<i>Fundulus heteroclitus</i>	NOEC	8.36	13.07	14	Coe et al., 2009
						8.36	13.07	14	Coe et al., 2009
						10.00	15.63	7	Lister et al., 2009
						10.00	15.63	7	Lister et al., 2009
						10.00	15.63	7	Lister et al., 2009
						10.00	15.63	7	Lister et al., 2009
		LOEC			10.00	15.63	10	Filby et al., 2012	
					10.00	15.63	10	Filby et al., 2012	
					10.00	15.63	10	Filby et al., 2012	
					10.00	15.63	10	Filby et al., 2012	
					10.00	15.63	322	Wenzel et al., 2001	
					10.00	15.63	322	Wenzel et al., 2001	
					10.60	16.57	17	Coe et al., 2009	
					10.60	16.57	17	Coe et al., 2009	
					25.00	39.08	14	Reyhanian et al., 2011	
		EC50		25.00	39.08	14	Reyhanian et al., 2011		
				47.30	7.39	2	Colman et al., 2009		
				47.30	7.39	2	Colman et al., 2009		
				100	156	14	Stromqvist et al., 2010		
				0.050	0.078	56	Schulz et al., 2007		
				0.30	0.47	322	Wenzel et al., 2001		
				0.50	0.078	2	Colman et al., 2009		
				0.50	0.78	56	Schulz et al., 2007		
				1.10	1.72	322	Wenzel et al., 2001		
				1.55	2.42	322	Wenzel et al., 2001		
				1.55	2.42	322	Wenzel et al., 2001		
				2.00	3.13	88	Xu et al., 2008		
				2.00	3.13	88	Xu et al., 2008		
				2.00	3.13	88	Xu et al., 2008		
				EC50	3.85	6.02	122	Micael et al., 2007	
		5.00	7.82		14	Reyhanian et al., 2011			
		5.00	7.82		14	Reyhanian et al., 2011			
		5.00	7.82		14	Reyhanian et al., 2011			
		5.00	7.82		56	Schulz et al., 2007			
		5.00	7.82		56	Schulz et al., 2007			
		5.00	7.82		56	Schulz et al., 2007			
5.00	7.82	56	Schulz et al., 2007						
5.58	8.72	122	Larsen et al., 2009						
5.58	8.72	122	Larsen et al., 2009						
5.80	0.91	2	Colman et al., 2009						
8.36	13.07	14	Coe et al., 2009						
9.20	14.38	7	Lange et al., 2012						
10.00	15.63	7	Lister et al., 2009						
EC50	10.00	15.63	10		Filby et al., 2012				
	10.00	15.63	10	Filby et al., 2012					
	10.00	15.63	10	Filby et al., 2012					
	10.00	15.63	10	Filby et al., 2012					
	10.00	15.63	10	Filby et al., 2012					
	10.00	15.63	10	Filby et al., 2012					
	10.00	15.63	21	Martyniuk et al., 2007					
	10.00	15.63	88	Xu et al., 2008					
	10.00	15.63	322	Wenzel et al., 2001					
	10.60	16.57	17	Coe et al., 2009					
	25.00	39.08	14	Reyhanian et al., 2011					
	30.00	4.69	2	Biales et al., 2007					
	100	156	14	Stromqvist et al., 2010					
	EC50	1.10	1.72	322	Wenzel et al., 2001				
		6.22	9.73	21	Van den Belt et al., 2004				
8.00		12.51	21	Van den Belt et al., 2004					
LC50		100	156	28	Wenzel et al., 2001				
		1700000	265772	4	Wenzel et al., 2001				
		Fish	<i>Gasterosteus aculeatus</i>	NOEC	67.90	106	14	Hogan et al., 2010	
248	388				14	Hogan et al., 2010			
248	388				14	Hogan et al., 2010			
LOEC	248			388	14	Hogan et al., 2010			
	67.90			106	14	Hogan et al., 2010			
	248			388	14	Hogan et al., 2010			
NOEC	1.75	2.74	28	Mauder et al., 2007					

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference	
57-63-6	17 $\alpha$ -Ethinyl estradiol	Fish	<i>Gasterosteus aculeatus</i>	NOEC	1.75	2.74	28	Maunder et al., 2007
					1.75	2.74	28	Maunder et al., 2007
					1.75	2.74	28	Maunder et al., 2007
					1.80	2.81	7	Lange et al., 2012
					2.40	3.75	21	Katsiadaki et al., 2010
					3.00	4.69	7	Lange et al., 2012
					6.35	0.99	4	Katsiadaki et al., 2010
					7.30	11.41	58	Hahlbeck et al., 2004b
					7.30	11.41	58	Hahlbeck et al., 2004b
					7.30	11.41	58	Hahlbeck et al., 2004b
					10.00	15.63	58	Hahlbeck, 2004
					10.00	15.63	58	Hahlbeck, 2004
					10.15	15.87	28	Bjorkblom et al., 2009
					10.15	15.87	28	Bjorkblom et al., 2009
					10.15	15.87	28	Bjorkblom et al., 2009
					10.15	15.87	28	Bjorkblom et al., 2009
					10.15	15.87	28	Bjorkblom et al., 2009
					15.00	2.35	3	Dziewieczynski, 2011
					15.00	2.35	3	Dziewieczynski, 2011
					27.70	43.31	28	Maunder et al., 2007
				27.70	43.31	28	Maunder et al., 2007	
				50.00	78.17	42	Le Mer et al., 2013	
				50.00	78.17	58	Hahlbeck et al., 2004a	
				50.00	78.17	58	Hahlbeck, 2004	
				78.90	12.33	4	Katsiadaki et al., 2010	
				100	156	58	Hahlbeck, 2004	
				LOEC	1.01	0.16	4	Katsiadaki et al., 2010
					1.75	2.74	28	Maunder et al., 2007
					1.75	2.74	28	Maunder et al., 2007
					4.00	6.25	58	Hahlbeck, 2004
					4.10	6.41	21	Katsiadaki et al., 2010
					7.30	11.41	58	Hahlbeck et al., 2004b
					7.30	11.41	58	Hahlbeck et al., 2004b
					9.50	14.85	7	Lange et al., 2012
					10.15	15.87	28	Bjorkblom et al., 2009
					10.15	15.87	28	Bjorkblom et al., 2009
					15.00	2.35	3	Dziewieczynski, 2011
					15.00	2.35	3	Dziewieczynski, 2011
					15.00	2.35	3	Dziewieczynski, 2011
					15.00	2.35	3	Dziewieczynski, 2011
					15.00	2.35	3	Dziewieczynski, 2011
					17.65	2.76	4	Katsiadaki et al., 2010
					27.70	43.31	28	Maunder et al., 2007
					27.70	43.31	28	Maunder et al., 2007
					27.70	43.31	28	Maunder et al., 2007
					27.70	43.31	28	Maunder et al., 2007
					50.00	78.17	42	Le Mer et al., 2013
					50.00	78.17	58	Hahlbeck et al., 2004a
					50.00	78.17	58	Hahlbeck et al., 2004a
					50.00	78.17	58	Hahlbeck, 2004
	100	156	58	Hahlbeck, 2004				
	<i>Gobiocypris rarus</i>	NOEC	100	156	21	Ma et al., 2007		
		LOEC	0.20	0.31	21	Ma et al., 2007		
			0.20	0.31	21	Ma et al., 2007		
	<i>Oncorhynchus mykiss</i>	NOEC	0.21	0.33	14	Thorpe et al., 2003		
			0.78	1.22	7	Lange et al., 2012		
			1.00	1.56	10	Purdom et al., 1994		
			1.10	1.72	14	Thorpe et al., 2003		
			7.85	12.27	56	Brown et al., 2007		
			10.00	15.63	14	Albertsson et al., (2007)		
			26.00	40.65	14	Thorpe et al., 2003		
			37.00	57.84	7	Hook et al., 2006		
			59.65	93.25	56	Brown et al., 2007		
			59.65	93.25	56	Brown et al., 2007		
		LOEC	0.10	0.16	10	Purdom et al., 1994		
			1.00	1.56	14	Thorpe et al., 2003		
			2.50	0.39	1	Biales et al., 2007		
			3.00	4.69	7	Lange et al., 2012		
			7.60	11.88	14	Thorpe et al., 2003		
			10.00	15.63	10	Purdom et al., 1994		
			12.50	19.54	50	Brown et al., 2008		
			59.65	93.25	56	Brown et al., 2007		
	<i>Oryzias latipes</i>	NOEC	0.20	0.31	14	Thompson, 2000		
			0.20	0.31	14	Tilton et al., 2005		
			0.20	0.31	14	Tilton et al., 2005		
			0.20	0.31	21	Ma et al., 2007		
			0.20	0.31	21	Ma et al., 2007		
			1.90	2.97	7	Lange et al., 2012		
			5.00	7.82	7	Zhang et al., 2008		
			5.00	7.82	14	Tilton et al., 2005		
			5.00	7.82	14	Tilton et al., 2005		
			5.00	7.82	14	Tilton et al., 2005		
			5.00	7.82	14	Tilton et al., 2005		
			10.00	15.6	1	Biales et al., 2007		
			20.00	31.27	21	Ma et al., 2007		
			50.00	78.17	7	Park et al., 2009		
			50.00	78.17	7	Zhang et al., 2008		
			50.00	78.17	7	Zhang et al., 2008		

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference				
57-63-6	17 $\alpha$ -Ethinyl estradiol	Fish	<i>Oryzias latipes</i>	NOEC	94.80	148	28	Hano et al., 2007			
					94.80	148	28	Hano et al., 2007			
					94.80	148	28	Hano et al., 2007			
					94.80	148	28	Hano et al., 2007			
					100	156	21	Ma et al., 2007			
					216	338	28	Hano et al., 2007			
					216	338	28	Hano et al., 2007			
					216	338	28	Hano et al., 2007			
					480	750	21	Hashimoto et al., 2009			
					480	750	21	Hashimoto et al., 2009			
					500	782	7	Park et al., 2009			
					500	782	7	Park et al., 2009			
					500	782	7	Park et al., 2009			
					500	782	7	Park et al., 2009			
					500	782	7	Zhang et al., 2008			
					500	782	7	Zhang et al., 2008			
					500	782	7	Zhang et al., 2008			
					500	782	7	Zhang et al., 2008			
					500	782	14	Thompson, 2000			
					500	782	14	Tilton et al., 2005			
				500	782	14	Tilton et al., 2005				
				522	816	28	Hano et al., 2007				
				LOEC	0.20	0.31	14	Tilton et al., 2005			
				2.00	3.13	21	Ma et al., 2007				
				2.00	3.13	21	Ma et al., 2007				
				5.00	7.82	7	Park et al., 2009				
				5.00	7.82	7	Park et al., 2009				
				5.00	7.82	7	Zhang et al., 2008				
				5.00	7.82	14	Thompson, 2000				
				5.00	7.82	14	Tilton et al., 2005				
				9.20	14.38	7	Lange et al., 2012				
				50.00	78.17	7	Zhang et al., 2008				
				50.00	78.17	21	Ma et al., 2007				
				60.00	93.80	21	Hashimoto et al., 2009				
				100	15.63	1	Biales et al., 2007				
				216	338	28	Hano et al., 2007				
				216	338	28	Hano et al., 2007				
				216	338	28	Hano et al., 2007				
				500	782	7	Park et al., 2009				
				500	782	7	Zhang et al., 2008				
				500	782	14	Tilton et al., 2005				
				500	782	14	Tilton et al., 2005				
				500	782	14	Tilton et al., 2005				
				500	782	14	Tilton et al., 2005				
				522	816	28	Hano et al., 2007				
				522	816	28	Hano et al., 2007				
				522	816	28	Hano et al., 2007				
				522	816	28	Hano et al., 2007				
				<i>Pimephales promelas</i>			NOEC	0.98	1.53	7	Lange et al., 2012
								1.60	2.50	14	Ankley et al., 2010
5.40	0.84	2	Martyniuk et al., 2010								
5.40	0.84	2	Martyniuk et al., 2010								
5.45	8.52	1095	Palace et al., 2009								
5.55	8.68	1095	Palace et al., 2009								
5.55	8.68	1095	Palace et al., 2009								
5.55	8.68	1095	Palace et al., 2009								
6.10	9.54	1095	Palace et al., 2009								
6.10	9.54	1095	Palace et al., 2009								
6.10	9.54	1095	Palace et al., 2009								
7.00	10.94	8	Ekman et al., 2008								
7.00	10.94	8	Ekman et al., 2008								
7.80	12.19	14	Ankley et al., 2010								
10.00	15.63	17	McGee et al., 2009								
10.00	15.63	21	Filby and Tyler, 2007								
10.00	15.63	21	Panter et al., 2004								
10.00	15.63	21	Panter et al., 2004								
10.00	15.63	21	Salierno and Kane, 2009								
10.00	15.63	21	Salierno and Kane, 2009								
10.60	16.57	21	Filby et al., 2007								
10.60	16.57	21	Filby et al., 2007								
10.60	16.57	21	Filby et al., 2007								
10.60	16.57	21	Filby et al., 2007								
20.00	31.27	21	Salierno and Kane, 2009								
20.00	31.27	21	Salierno and Kane, 2009								
40.00	62.53	21	Salierno and Kane, 2009								
40.00	62.53	21	Salierno and Kane, 2009								
50.00	78.17	15	Weisbrod et al., 2007								
50.00	78.17	15	Weisbrod et al., 2007								
50.00	78.17	15	Weisbrod et al., 2007								
50.00	78.17	15	Weisbrod et al., 2007								
50.00	78.17	15	Weisbrod et al., 2007								
50.00	78.17	15	Weisbrod et al., 2007								
83.00	130	8	Ekman et al., 2008								
83.00	130	8	Ekman et al., 2008								
100	156	30	Warner, 2006								
LOEC	100000	156337	30	Warner, 2006							
LOEC	1.60	2.50	14	Ankley et al., 2010							

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference				
57-63-6	17 $\alpha$ -Ethinyl estradiol	Fish	<i>Pimephales promelas</i>	LOEC	2.90	4.53	7	Lange et al., 2012			
					5.40	0.84	2	Martyniuk et al., 2010			
					5.40	0.84	2	Martyniuk et al., 2010			
					5.40	0.84	2	Martyniuk et al., 2010			
					5.45	8.52	1095	Palace et al., 2009			
					5.55	8.68	1095	Palace et al., 2009			
					5.55	8.68	1095	Palace et al., 2009			
					6.10	9.54	1095	Palace et al., 2009			
					6.10	9.54	1095	Palace et al., 2009			
					7.00	10.94	8	Ekman et al., 2008			
					7.80	12.19	14	Ankley et al., 2010			
					10.00	15.63	21	Filby and Tyler, 2007			
					10.00	15.63	21	Panter et al., 2004			
					10.00	15.63	21	Salierno and Kane, 2009			
					10.00	15.63	21	Salierno and Kane, 2009			
					10.00	15.63	21	Salierno and Kane, 2009			
					10.00	15.63	21	Salierno and Kane, 2009			
					10.60	16.57	21	Filby et al., 2007			
					10.60	16.57	21	Filby et al., 2007			
					10.60	16.57	21	Filby et al., 2007			
					10.60	16.57	21	Filby et al., 2007			
					20.00	31.27	21	Salierno and Kane, 2009			
					40.00	62.53	21	Salierno and Kane, 2009			
					40.00	62.53	21	Salierno and Kane, 2009			
					50.00	78.17	15	Weisbrod et al., 2007			
					50.00	78.17	15	Weisbrod et al., 2007			
					50.00	78.17	15	Weisbrod et al., 2007			
					50.00	78.17	15	Weisbrod et al., 2007			
					83.00	130	8	Ekman et al., 2008			
					83.00	130	8	Ekman et al., 2008			
					100	156	30	Warner, 2006			
					1000	1563	30	Warner, 2006			
					Fish	<i>Poecilia reticulata</i>	NOEC	10.00	15.63	14	Hallgren and Olsen, 2010
								10.00	15.63	14	Hallgren and Olsen, 2010
								10.00	15.63	14	Hallgren and Olsen, 2010
								2000	3127	112	Shenoy, 2012
								2000	3127	112	Shenoy, 2012
							LOEC	50000	78168	14	Hallgren and Olsen, 2010
								10.00	15.63	14	Hallgren and Olsen, 2010
								2000	3127	112	Shenoy, 2012
		2000	3127	112				Shenoy, 2012			
		50000	78168	14				Hallgren and Olsen, 2010			
		Fish	<i>Rutilus rutilus</i>	NOEC	50000	78168	14	Hallgren and Olsen, 2010			
					0.040	0.063	720	Lange et al., 2009			
					0.040	0.063	720	Lange et al., 2009			
					0.10	0.16	94	Katsu et al., 2007			
					0.30	0.47	94	Katsu et al., 2007			
					0.30	0.47	122	Lange et al., 2008			
					0.30	0.47	122	Lange et al., 2008			
					0.30	0.47	720	Lange et al., 2009			
					0.30	0.47	720	Lange et al., 2009			
					0.78	1.22	7	Lange et al., 2012			
					4.00	6.25	122	Lange et al., 2008			
					4.00	6.25	122	Lange et al., 2008			
					4.00	6.25	122	Lange et al., 2008			
					4.00	6.25	720	Lange et al., 2009			
					4.00	6.25	720	Lange et al., 2009			
					28.50	44.56	18	Flores-Valverde et al., 2010			
					28.50	44.56	18	Flores-Valverde et al., 2010			
					Fish	<i>Salmo trutta</i>	LOEC	0.10	0.16	94	Katsu et al., 2007
								0.30	0.47	94	Katsu et al., 2007
								0.30	0.47	122	Lange et al., 2008
		0.30	0.47	720				Lange et al., 2009			
		1.10	1.72	18				Flores-Valverde et al., 2010			
		NOEC	3.00	4.69			7	Lange et al., 2012			
			4.00	6.25			94	Katsu et al., 2007			
			4.00	6.25			122	Lange et al., 2008			
			4.00	6.25			122	Lange et al., 2008			
			4.00	6.25			720	Lange et al., 2009			
		Fish	<i>Salmo trutta</i>	NOEC	1.10	1.72	12	Bjerregaard et al., 2008			
					2.08	3.25	21	Korner, 2008			
					2.08	3.25	21	Korner, 2008			
					2.08	3.25	21	Korner, 2008			
					2.08	3.25	21	Korner, 2008			
					2.08	3.25	21	Korner, 2008			
					2.12	3.31	21	Korner, 2008			
					2.12	3.31	21	Korner, 2008			
					2.12	3.31	21	Korner, 2008			
					2.12	3.31	21	Korner, 2008			
					2.12	3.31	21	Korner, 2008			
					2.40	3.75	21	Korner, 2008			
					2.40	3.75	21	Korner, 2008			
					2.40	3.75	21	Korner, 2008			
					12.00	18.76	12	Bjerregaard et al., 2008			
		Fish	<i>Salmo trutta</i>	LOEC	1.10	1.72	12	Bjerregaard et al., 2008			
					2.08	3.25	21	Korner, 2008			

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference			
57-63-6	17 $\alpha$ -Ethinyl estradiol	Fish	<i>Salmo trutta</i>	LOEC	2.12	3.31	21	Korner, 2008		
					2.12	3.31	21	Korner, 2008		
					2.40	3.75	21	Korner, 2008		
					2.40	3.75	21	Korner, 2008		
					5.10	7.97	12	Bjerregaard et al., 2008		
			EC50	3.70	5.78	12	Bjerregaard et al., 2008			
				5.20	8.13	12	Bjerregaard et al., 2008			
				<i>Salvelinus namaycush</i>	NOEC	5.45	8.52	1095	Palace et al., 2009	
						5.45	8.52	1095	Palace et al., 2009	
						5.55	8.68	1095	Palace et al., 2009	
			5.55			8.68	1095	Palace et al., 2009		
			6.10			9.54	1095	Palace et al., 2009		
			LOEC	6.10	9.54	1095	Palace et al., 2009			
				6.30	9.85	365	Werner, 2006.			
				6.30	9.85	365	Werner, 2006.			
				6.30	9.85	365	Werner, 2006.			
				6.30	9.85	365	Werner, 2006.			
				15.00	23.45	21	Werner, 2006.			
				373	583	21	Werner, 2006.			
				5.45	8.52	1095	Palace et al., 2009			
		5.45		8.52	1095	Palace et al., 2009				
		5.55		8.68	1095	Palace et al., 2009				
		NOEC	5.55	8.68	1095	Palace et al., 2009				
			6.10	9.54	1095	Palace et al., 2009				
			6.10	9.54	1095	Palace et al., 2009				
			6.30	9.85	365	Werner, 2006.				
			6.30	9.85	365	Werner, 2006.				
			6.30	9.85	365	Werner, 2006.				
			35.00	54.72	21	Werner, 2006.				
			Insecta	<i>Syrnathus scovelli</i>	NOEC	1.00	1.56	10	Partridge et al., 2010	
						100	156	10	Partridge et al., 2010	
						1.00	1.56	10	Partridge et al., 2010	
		1.00				1.56	10	Partridge et al., 2010		
		100				156	10	Partridge et al., 2010		
		Insecta	<i>Chironomus tentans</i>	NOEC	20000	31267	47	Dussault et al., 2008b		
					70000	109436	47	Dussault et al., 2008b		
					550000	859852	47	Dussault et al., 2008b		
					550000	859852	47	Dussault et al., 2008b		
					560000	875486	21	Dussault et al., 2009		
					560000	875486	47	Dussault et al., 2008b		
					560000	875486	47	Dussault et al., 2008b		
					560000	875486	47	Dussault et al., 2008b		
					560000	875486	47	Dussault et al., 2008b		
					1000000	166337	2	Cho, 2005		
					20000	31267	47	Dussault et al., 2008b		
					140000	218871	47	Dussault et al., 2008b		
					3100000	4846438	21	Dussault et al., 2009		
					3100000	4846438	47	Dussault et al., 2008b		
					3100000	4846438	47	Dussault et al., 2008b		
					1510000	2360684	47	Dussault et al., 2008b		
1530000	2391952				47	Dussault et al., 2008b				
1550000	2423219				47	Dussault et al., 2008b				
1960000	3064199				47	Dussault et al., 2008b				
6600000	10318222				10	Dussault et al., 2008a				
Mollusca	<i>Bithynia tentaculata</i>	NOEC	2160000	3376873	47	Dussault et al., 2008b				
			9.00	14.07	284	Hallgren et al., 2012				
			44950	70273	284	Hallgren et al., 2012				
			44950	70273	284	Hallgren et al., 2012				
			44950	70273	284	Hallgren et al., 2012				
			44950	70273	284	Hallgren et al., 2012				
			44950	70273	284	Hallgren et al., 2012				
			LOEC	9.00	14.07	284	Hallgren et al., 2012			
				Potamopyrgus antipodarum	NOEC	25.00	39.08	28	Sieratowicz et al., 2011	
						100	156	28	Sieratowicz et al., 2011	
						50.00	78.17	28	Sieratowicz et al., 2011	
						Radix balthica	NOEC	9.00	14.07	153
			5130					8020	153	Hallgren et al., 2012
			5130	8020	153			Hallgren et al., 2012		
			5130	8020	153			Hallgren et al., 2012		
			44950	70273	153			Hallgren et al., 2012		
			Aquatic community	PNEC	9.00	14.07	153	Hallgren et al., 2012		
					0.035	0.055		James et al., 2014		
					0.037	0.058		Oekotoxzentrum, Centre Ecotox		
					0.10	0.16		James et al., 2014		
0.10	0.16				James et al., 2014					
0.35	0.55				James et al., 2014					
0.50	0.78				James et al., 2014					
0.57	0.89				Johnson et al., 2007					
20000	20000	10			Julius et al., 2007					
400000	400000	10			Julius et al., 2007					
50-28-2	17 $\beta$ -estradiol	Algae	<i>Melosira varians</i>	NOEC	800000	800000	10	Julius et al., 2007		
					80000	80000	10	Julius et al., 2007		
					200000	200000	10	Julius et al., 2007		
					100000	100000	506	Coady et al., 2004		
					100000	100000	506	Coady et al., 2004		
		Amphibia	<i>Lithobates clamitans ssp. Clamitans</i>	NOEC	100000	100000	506	Coady et al., 2004		
					100000	100000	506	Coady et al., 2004		
					<i>Lithobates pipiens</i>	NOEC	100000	100000	124	Mackenzie et al., 2003
							100000	100000	124	Mackenzie et al., 2003
							1000	1000	124	Mackenzie et al., 2003
1000	1000	124	Mackenzie et al., 2003							
LC50	1242098	1242098	14	Hogan et al., 2006						

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference		
50-28-2	17β-estradiol	Amphibia	<i>Lithobates pipiens</i>	LC50	15172.12	15172.12	14	Hogan et al., 2006	
			<i>Lithobates sylvaticus</i>	LC50	680975	680975	14	Hogan et al., 2006	
			<i>Rhinella arenarum</i>	NOEC	100000	100000	200	Brodeur et al., 2013	
				LOEC	100000	100000	200	Brodeur et al., 2013	
					100000	100000	200	Brodeur et al., 2013	
			<i>Xenopus laevis</i>	NOEC	100000	100000	78	Carr et al., 2003	
				LOEC	100000	100000	34	Cong et al., 2006	
					100000	100000	34	Cong et al., 2006	
					100000	100000	78	Carr et al., 2003	
					100000	100000	78	Carr et al., 2003	
		Crustacea	<i>Americamysis bahia</i>	LC50	890000	890000	4	Hirano et al., 2004	
					1690000	169000	2	Hirano et al., 2004	
			<i>Ceriodaphnia dubia</i>	NOEC	1000000	1000000	7	Jukosky et al., 2008a	
			<i>Daphnia magna</i>	NOEC	200000	200000	21	Brennan et al., 2006	
					1000000	1000000	21	Brennan et al., 2006	
					1000000	1000000	21	Brennan et al., 2006	
				LOEC	400000	400000	21	Brennan et al., 2006	
				EC50	1550000	155000	1	Brennan et al., 2006	
					2040000	204000	2	Brennan et al., 2006	
					2870000	287000	2	Brennan et al., 2006	
					2970000	297000	2	Hirano et al., 2004	
					3670000	367000	1	Brennan et al., 2006	
			<i>Eurytemora affinis</i>	NOEC	6000	6000	10	Forget-Leray et al., 2005	
				LOEC	18000	18000	10	Forget-Leray et al., 2005	
				LC50	45000	45000	4	Forget-Leray et al., 2005	
			<i>Neocaridina denticulata</i>	LOEC	10000	10000	28	Huang et al., 2006	
					10000	10000	28	Huang et al., 2006	
					10000	10000	28	Huang et al., 2006	
			Fish	<i>Cyprinus carpio</i>	NOEC	100	100	90	Gimeno et al., 1998
					LOEC	100	100	90	Gimeno et al., 1998
					1000	1000	90	Gimeno et al., 1998	
		<i>Danio rerio</i>		NOEC	12.90	12.90	8	Rose et al., 2002	
					24.00	24.00	18	Holbech et al., 2006	
					24.00	24.00	18	Holbech et al., 2006	
					25.00	2.50	2	Jin et al., 2009	
					250	25000	2	Jin et al., 2009	
					250	250	18	Holbech et al., 2006	
					2500	250	3	Jin et al., 2010	
					12500	1250	3	Jin et al., 2010	
				LOEC	2100	2100	8	Rose et al., 2002	
					25.00	2.50	2	Jin et al., 2009	
					54.00	54.00	18	Holbech et al., 2006	
					54.00	54.00	18	Holbech et al., 2006	
					1000	1000	16	Peute et al., 1985	
					12500	1250	3	Jin et al., 2010	
				EC50	4120	4120	8	Rose et al., 2002	
					55.00	55.00	18	Holbech et al., 2006	
					175	175	21	Van den Belt et al., 2004	
					240	240	21	Van den Belt et al., 2004	
		<i>Gambusia affinis</i>		NOEC	1000	1000	8	Huang et al., 2013	
					2500000	25000	3	Kamata et al., 2011	
				LOEC	1000	1000	8	Huang et al., 2012b	
					1000	1000	8	Huang et al., 2013	
					500000	50000	3	Kamata et al., 2011	
		<i>Gambusia holbrooki</i>		NOEC	20.00	20.00	28	Doyle and Lim, 2005	
					100	100	84	Rawson et al., 2006	
				LOEC	100	100	28	Doyle and Lim, 2005	
					500	500	28	Doyle and Lim, 2005	
					500	500	84	Rawson et al., 2006	
		<i>Gasterosteus aculeatus</i>		NOEC	1.00	1.00	7	Hogan et al., 2008	
					10.00	10.00	21	Allen et al., 2008	
					10.00	10.00	58	Hahlbeck et al., 2004a	
					20.00	20.00	21	Allen et al., 2008	
					32.00	32.00	21	Allen et al., 2008	
					10000	10000	58	Hahlbeck et al., 2004a	
				LOEC	10.00	10.00	7	Hogan et al., 2008	
					50.00	50.00	21	Allen et al., 2008	
					70.00	70.00	21	Allen et al., 2008	
					100	100	21	Allen et al., 2008	
					1000	1000	58	Hahlbeck et al., 2004a	
		<i>Gobiocypris rarus</i>		NOEC	100	10.00	4	Ma et al., 2009	
				LOEC	100	10.00	4	Ma et al., 2009	
		<i>Ictalurus punctatus</i>		NOEC	100	100	21	Thompson et al., 2000	
				LOEC	1000	1000	21	Thompson et al., 2000	
				EC50	170	170	21	Thompson et al., 2000	
		<i>Morone saxatilis</i>		NOEC	1000	1000	21	Thompson et al., 2000	
				LOEC	10000	10000	21	Thompson et al., 2000	
				EC50	1560	1560	21	Thompson et al., 2000	
		<i>Oncorhynchus mykiss</i>	NOEC	3.20	3.20	21	Thorpe et al., 2000		
				4.80	4.80	14	Thorpe et al., 2003		
				9.60	9.60	14	Thorpe et al., 2003		
				100	10.00	5	Ward et al., 2006		
				100	100	21	Thorpe et al., 2000		
				100	100	21	Tremblay and Van der Kraak, 1999		
				100	100	21	Tremblay and Van der Kraak, 1999		
				247	247	21	Thorpe et al., 2000		
				250	250	21	Tremblay and Van der Kraak, 1999		
				463	463	14	Thorpe et al., 2003		

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference				
50-28-2	17β-estradiol	Fish	<i>Oncorhynchus mykiss</i>	LOEC	8.90	8.90	21	Thorpe et al., 2000			
					14.00	14.00	14	Thorpe et al., 2003			
					22.00	22.00	14	Thorpe et al., 2003			
					100	100	21	Tremblay and Van der Kraak, 1999			
					247	247	21	Thorpe et al., 2000			
					250	250	21	Tremblay and Van der Kraak, 1999			
					250	250	21	Tremblay and Van der Kraak, 1999			
					15.00	15.00	21	Thorpe et al., 2000			
					EC50	400000	400000	8 h immersion	Piferrer and Donaldson, 1992		
					NOEC	1.00	0.10		Lee et al., 2012		
						1.00	0.10		Lee et al., 2012		
						10.00	1.00	5	Kang et al., 2005		
						100	100	21	Thompson et al., 2000		
						154	154	14	Jukosky et al., 2008b		
						227	227	25	Kang et al., 2002		
						1000	100		Lee et al., 2012		
						2528	2528	14	Jukosky et al., 2008b		
						10000	1000	5	Kang, et al., 2006b		
						5.00	5.00	21	Kashiwada et al., 2002		
						10.00	1.00	5	Kang et al., 2005		
						10.00	1.00		Lee et al., 2012		
						10.00	1.00		Lee et al., 2012		
						29.30	29.30	21	Kang et al., 2002		
						55.70	55.70	21	Kang et al., 2002		
						56.27	56.27	14	Jukosky et al., 2008b		
						100	10.00		Lee et al., 2012		
						463	463	25	Kang et al., 2002		
						1000	1000	21	Thompson et al., 2000		
						1000	100		Lee et al., 2012		
						1000	100		Lee et al., 2012		
						2528	2528	14	Jukosky et al., 2008b		
						10000	1000	5	Kang, et al., 2006b		
						EC50	200	200	21	Thompson et al., 2000	
							225	225	14	Sun et al., 2009	
							470000	470000	21	Kashiwada et al., 2002	
							LC50	460000	46000	3	Kashiwada et al., 2002
								460000	46000	3	Tabata et al., 2001
								2000000	200000	4	Kang et al., 2002
								3500000	350000	3	Kashiwada et al., 2002
								3500000	350000	3	Tabata et al., 2001
						<i>Pimephales promelas</i>	NOEC	10.00	10.00	14	Thorpe et al., 2007
								27.00	27.00	9	Cline et al., 2003
								28.00	28.00	17	McGee et al., 2009
								30.00	30.00	21	Schultz et al., 2012
								30.00	30.00	21	Schultz et al., 2012
								30.00	30.00	21	Schultz et al., 2012
								30.00	30.00	21	Schultz et al., 2012
								30.00	30.00	21	Schultz et al., 2012
								100	100	14	Thorpe et al., 2007
								500	500	21	Bringolf et al., 2004
			500	500	21		Bringolf et al., 2004				
			22.00	22.00	14		Thorpe et al., 2007				
			30.00	30.00	21		Schultz et al., 2012				
			270	270	9		Cline et al., 2003				
			500	500	21		Bringolf et al., 2004				
			500	500	21		Bringolf et al., 2004				
			500	500	21		Bringolf et al., 2004				
			EC50	25.00	25.00		14	Brian et al., 2005			
				120	120		19	Kramer et al., 1998			
				251	251		19	Kramer et al., 1998			
			LC50	1150	1150	19	Kramer et al., 1998				
		<i>Poecilia reticulata</i>	NOEC	50.00	50.00	120	Nielsen and Baatrup, 2006				
				1000	1000	21	Li and Wang, 2005				
				1000	1000	21	Li and Wang, 2005				
		<i>Pomatoschistus minutus</i>	LOEC	1000	1000	21	Li and Wang, 2005				
				71.00	71.00	243	Robinson et al., 2004				
				EC50	87.00	87.00	243	Robinson et al., 2004			
		<i>Salmo trutta</i>		127	127	243	Robinson et al., 2004				
				165	165	243	Robinson et al., 2004				
				NOEC	15.30	15.30	8	Bjerregaard et al., 2008			
			LOEC	20.00	20.00	8	Bjerregaard et al., 2008				
			EC50	15.10	15.10	8	Bjerregaard et al., 2008				
		<i>Thymallus thymallus</i>	LOEC	1.00	1.00	50	Lahnsteiner et al., 2006				
				1.00	1.00	50	Lahnsteiner et al., 2006				
				LC50	2740000	274000		Nendza and Wenzel, 2006			
		Invertebrates	<i>Brachionus calyciflorus</i>	LOEC	1.00	1.00	10	Huang et al., 2012a			
					100000	100000	10	Huang et al., 2012a			
		Mollusca	<i>Elliptio complanata</i>	NOEC	100000	10000	0.25	Flynn et al., 2013			
					LOEC	100000	10000	0.25	Flynn et al., 2013		
		Aquatic community	PNEC		0.40	0.40		Oekotoxzentrum, Centre Ecotox			
						1.00	1.00		Gross-Sorokin et al., 2006		
						1.00	1.00		Young et al., 2004		
						2.00	2.00		Anderson et al., 2012 ; Caldwell et al., 2012		
						2.27	2.27		Yuan et al., 2014		
14938-35-3	4-n-amyphenol	Algae	<i>Chlorella pyrenoidosa</i>	NOEC	980000	980	3	Ramos et al., 1999			
				LOEC	2300000	2300	3	Ramos et al., 1999			
				EC50	2600000	2600	3	Ramos et al., 1999			
					1330000	1330	2	Ramos et al., 1998			
		Crustacea	<i>Daphnia magna</i>	EC50	1330000	1330	2	Ramos et al., 1998			



CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference			
14938-35-3	4-n-amyphenol	Crustacea	<i>Daphnia magna</i>	EC50	2040000	2040	1	Ramos et al., 1998		
			<i>Poecilia reticulata</i>	LC50	1250000	1250	4	Ramos et al., 1998		
		Fish				1360000	1360	3	Ramos et al., 1998	
						1920000	1920	2	Ramos et al., 1998	
			Mollusca	<i>Lymnaea stagnalis</i>			2490000	2490	1	Ramos et al., 1998
							3700000	3700	3	Ramos et al., 1998
							3710000	37100	4	Ramos et al., 1998
							4600000	4600	2	Ramos et al., 1998
							5380000	5380	1	Ramos et al., 1998
56-53-1	Diethylstilbestrol	Amphibia	<i>Xenopus laevis</i>	LOEC	2684	44.01	3	Nishimura et al., 1997		
					2684	44.01	3	Nishimura et al., 1997		
		Crustacea	<i>Daphnia magna</i>	NOEC	100000	16401	6	Kashian and Dodson, 2004		
					100000	16401	6	Kashian and Dodson, 2004		
					100000	16401	21	Brennan et al., 2006		
					200000	3280	2	Zou and Fingerman, 1997		
					500000	82007	21	Baldwin et al., 1995		
					500000	82007	21	Baldwin et al., 1995		
					500000	82007	21	Baldwin et al., 1995		
					500000	82007	21	Brennan et al., 2006		
					500000	82007	21	Brennan et al., 2006		
					500000	82007	21	Brennan et al., 2006		
					540000	8857	2	Baldwin et al., 1995		
					900000	14761	2	Baldwin et al., 1995		
				LOEC	100000	16401	6	Kashian and Dodson, 2004		
					200000	32803	21	Brennan et al., 2006		
					500000	8201	2	Baldwin et al., 1995		
					500000	82007	21	Baldwin et al., 1995		
					500000	82007	21	Baldwin et al., 1995		
					500000	82007	21	Brennan et al., 2006		
					500000	82007	21	Brennan et al., 2006		
					500000	8201	2	Hannas et al., 2011		
					EC50	1090000	17878	2	Zou and Fingerman, 1997	
						1200000	19682	2	Baldwin et al., 1995	
						1550000	25422	2	Brennan et al., 2006	
						1870000	30671	2	Brennan et al., 2006	
						2030000	33295	1	Brennan et al., 2006	
						3710000	60849	1	Brennan et al., 2006	
NOEC	30000	4920	18	Breitholtz and Bengtsson, 2001						
	30000	4920	18	Breitholtz and Bengtsson, 2001						
	30000	4920	18	Breitholtz and Bengtsson, 2001						
	LC50	290000	47564	4	Breitholtz and Bengtsson, 2001					
	Fish	<i>Pimephales promelas</i>	NOEC	3200	525	21	Panter et al., 2002			
			NOEC	3200	525	21	Panter et al., 2002			
	Worm	<i>Dugesia japonica</i>	LC50	500000	82007	4	Li, 2013			
				600000	9841	3	Li, 2013			
				700000	11481	2	Li, 2013			
				800000	13121	1	Li, 2013			
50-27-1			Estrilol	Fish	<i>Danio rerio</i>	NOEC	300	94.87	20 to 38 dph	Holbech et al., 2006
							6700	219	20 to 60 dph	Holbech et al., 2006
		21700				6862		Holbech et al., 2006		
	LOEC	600				190	20 to 38 dph	Holbech et al., 2006		
		21700				6862	20 to 60 dph	Holbech et al., 2006		
	NOEC	<i>Oryzias latipes</i>				4.30	1.36	15	Lei et al., 2014	
						46.50	14.70	15	Lei et al., 2014	
						46.50	14.70	90	Lei et al., 2014	
						100	31.62	110	Metcalfe et al., 2001	
						462	146	15	Lei et al., 2014	
				462	146	90	Lei et al., 2014			
				462	146	90	Lei et al., 2014			
				462	146	90	Lei et al., 2014			
				1000	3.16	110	Metcalfe et al., 2001			
				4517	1428	90	Lei et al., 2014			
	LOEC			46.50	14.70	15	Lei et al., 2014			
				462	146	15	Lei et al., 2014			
				462	146	90	Lei et al., 2014			
				462	146	90	Lei et al., 2014			
				1000	3.16	110	Metcalfe et al., 2001			
		4517	1428	15	Lei et al., 2014					
		4517	1428	90	Lei et al., 2014					
		4517	1428	90	Lei et al., 2014					
		4517	1428	90	Lei et al., 2014					
		4517	1428	90	Lei et al., 2014					
53-16-7	Estrone	Invertebrates	<i>Dugesia japonica</i>	NOEC	100000000	31622800	4	Li, 2013		
				NOEC	100	1.00	4	Ghekierre et al., 2006		
		Crustacea	<i>Neomysis integer</i>	NOEC	100000000	100000	4	Ghekierre et al., 2006		
				LOEC	10000	100	4	Ghekierre et al., 2006		
				Fish	<i>Danio rerio</i>	NOEC	35.50	0.36	40	Holbech et al., 2006
						NOEC	97.70	0.98	40	Holbech et al., 2006
					LOEC	14.00	0.14	18	Holbech et al., 2006	
					LOEC	49.80	0.50	40	Holbech et al., 2006	
					EC50	78.00	0.78	18	Holbech et al., 2006	
						204	2.04	21	Van den Belt et al., 2004	
						465	4.65	21	Van den Belt et al., 2004	
				NOEC	<i>Oncorhynchus mykiss</i>	0.74	0.0074	14	Thorpe et al., 2003	
						3.19	3.19	14	Thorpe et al., 2003	
						3.30	0.033	14	Thorpe et al., 2003	
					<i>Oryzias latipes</i>	NOEC	100	1.00	85 to 110	Metcalfe et al., 2001
							1000	10.00	85 to 110	Metcalfe et al., 2001
						LOEC	1000	10.00	85 to 110	Metcalfe et al., 2001
							10000	100	85 to 110	Metcalfe et al., 2001
					<i>Pimephales promelas</i>	NOEC	781	7.81	21	Thorpe et al., 2007
				LOEC		34.00	0.34	21	Thorpe et al., 2007	

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference		
53-16-7	Estrone	Fish	<i>Pimephales promelas</i>	LOEC	307	3.07	21	Thorpe et al., 2007	
					307	3.07	21	Thorpe et al., 2007	
			<i>Salmo trutta</i>	NOEC	63.00	0.63	10	Bjerregaard et al., 2008	
					89.00	0.89	10	Bjerregaard et al., 2008	
				LOEC	87.00	0.87	10	Bjerregaard et al., 2008	
					134	1.34	10	Bjerregaard et al., 2008	
				EC50	85.00	0.85	10	Bjerregaard et al., 2008	
					88.00	0.88	10	Bjerregaard et al., 2008	
			Aquatic community	PNEC	3.00	0.030		Johnson et al., 2007	
					3.60	0.036		Oekotoxzentrum, Centre Ecotox	
68-22-4	Norethindrone	Crustacea	<i>Daphnia magna</i>	NOEC	500000000	3155000	25	Goto and Hiromi, 2003	
					6410000	4045	2	Goto and Hiromi, 2003	
		Fish	<i>Ictalurus punctatus</i>	NOEC	82500	521	7	Nallani et al., 2012	
			<i>Pimephales promelas</i>	NOEC	370	2.33	28 days ph	Overturf et al., 2012	
					1500	9.47	28 days ph	Overturf et al., 2012	
					35400	223	28	Nallani et al., 2012	
					LOEC	740	4.67	28 days ph	Overturf et al., 2012
						14800	93.39	28 days ph	Overturf et al., 2012

**B: ANTI-AR CALUX**

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference	
57-63-6	17α-Ethinyl estradiol	Algae	<i>Scenedesmus subspicatus</i>	EC50	840	42.10	3	Kopf, 1997
				NOEC	0.0058	0.29	119	Park, 2003
					0.0058	0.29	119	Park, 2003
					0.0061	0.31	25	Park, 2003
					0.0061	0.31	105	Park and Kidd, 2005
					0.0061	0.31	105	Park and Kidd, 2005
					0.0061	0.31	105	Park and Kidd, 2005
					0.10	0.50	189	Park, 2003
					1.00	50.12	189	Park, 2003
					LOEC	0.0058	0.29	119
		0.0061	0.31	105	Park and Kidd, 2005			
		Amphibia	<i>Lithobates clamitans</i>	NOEC	0.0061	0.31	105	Park and Kidd, 2005
					0.083	4.13	189	Park, 2003
					0.0050	0.25	119	Park and Kidd, 2005
					0.0050	0.25	119	Park and Kidd, 2005
					0.0050	0.25	119	Park and Kidd, 2005
					0.0050	0.25	119	Park and Kidd, 2005
					0.12	5.76	124	Brande-Lavridsen et al., (2010)
					0.12	5.76	124	Brande-Lavridsen et al., (2010)
					0.12	5.76	124	Brande-Lavridsen et al., (2010)
				0.12	5.76	124	Brande-Lavridsen et al., (2010)	
		<i>Lithobates septentrionalis</i>	NOEC	0.0060	0.30	124	Brande-Lavridsen et al., (2010)	
				0.0060	0.30	124	Brande-Lavridsen et al., (2010)	
				0.0017	0.083	61	Gyllenhammar et al., 2009	
				0.0017	0.083	61	Gyllenhammar et al., 2009	
				0.018	0.88	61	Gyllenhammar et al., 2009	
				0.18	9.00	61	Gyllenhammar et al., 2009	
				0.18	9.00	61	Gyllenhammar et al., 2009	
				0.18	9.00	61	Gyllenhammar et al., 2009	
				0.18	9.00	61	Gyllenhammar et al., 2009	
			LOEC	0.0017	0.083	61	Gyllenhammar et al., 2009	
		<i>Rana temporaria</i>	NOEC	0.0017	0.083	61	Gyllenhammar et al., 2009	
				0.0018	0.090	61	Gyllenhammar et al., 2009	
				0.0018	0.090	61	Gyllenhammar et al., 2009	
				0.18	9.00	61	Gyllenhammar et al., 2009	
				0.18	9.00	61	Gyllenhammar et al., 2009	
				0.18	9.00	61	Gyllenhammar et al., 2009	
				0.18	9.00	61	Gyllenhammar et al., 2009	
				0.18	9.00	61	Gyllenhammar et al., 2009	
				0.18	9.00	61	Gyllenhammar et al., 2009	
			LOEC	0.0017	0.083	61	Gyllenhammar et al., 2009	
		Crustacea	<i>Ceriodaphnia dubia</i>	NOEC	0.0018	0.090	61	Gyllenhammar et al., 2009
					0.18	9.00	61	Gyllenhammar et al., 2009
					500	25059	7	Jukosky et al., 2008a
					500	25059	7	Jukosky et al., 2008a
					936	46886	7	Cho, 2005
					0.0001	0.0050	7	Dietrich et al., 2010
					0.0001	0.0050	7	Dietrich et al., 2010
					10.00	501	21	Kopf, 1997
					500	25059	21	Clubbs and Brooks, 2007
				500	25059	21	Clubbs and Brooks, 2007	
		<i>Daphnia magna</i>	NOEC	1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
			LOEC	0.0001	0.0050	7	Dietrich et al., 2010	
		<i>Hyalella azteca</i>	NOEC	0.0001	0.0050	7	Dietrich et al., 2010	
				62.50	3132	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
				1000	5019	21	Clubbs and Brooks, 2007	
			EC50	105	5262	21	Kopf, 1997	
		<i>Xenopus tropicalis</i>	NOEC	5700	28568	1	Kopf, 1997	
				2590	129828	4	Clubbs and Brooks, 2007	
				70.00	3508	63	Dussault et al., 2008b	
				70.00	3508	63	Dussault et al., 2008b	
				70.00	3508	63	Dussault et al., 2008b	
				70.00	3508	63	Dussault et al., 2008b	
				70.00	3508	63	Dussault et al., 2008b	
				740	37088	63	Dussault et al., 2008b	
				910	45608	21	Dussault et al., 2009	
			740	37088	63	Dussault et al., 2008b		
		Fish	<i>Catostomus commersoni</i>	NOEC	740	37088	63	Dussault et al., 2008b
					740	37088	63	Dussault et al., 2008b
					740	37088	63	Dussault et al., 2008b
					740	37088	63	Dussault et al., 2008b
					740	37088	63	Dussault et al., 2008b
					740	37088	63	Dussault et al., 2008b
					740	37088	63	Dussault et al., 2008b
					740	37088	63	Dussault et al., 2008b
					740	37088	63	Dussault et al., 2008b
				EC50	360	18043	63	Dussault et al., 2008b
		<i>Cyprinus carpio</i>	NOEC	770	38591	63	Dussault et al., 2008b	
				1300	65154	10	Dussault et al., 2008a	
				1100	55131	10	Dussault et al., 2008a	
				0.0055	0.27	1095	Palace et al., 2009	
				0.0056	0.28	1095	Palace et al., 2009	
				0.0061	0.31	1095	Palace et al., 2009	
				0.0055	0.27	1095	Palace et al., 2009	
				0.0055	0.27	1095	Palace et al., 2009	
				0.0056	0.28	1095	Palace et al., 2009	
0.0061	0.31		1095	Palace et al., 2009				
<i>Clarias gariepinus</i>	NOEC	0.0061	0.31	1095	Palace et al., 2009			
		1.00	50.12	21	Swapna and Senthilkumaran, 2009			
		LOEC	0.0007	0.033	7	Braathen et al., 2009		
		1.00	50.12	21	Swapna and Senthilkumaran, 2009			
		1.00	50.12	21	Swapna and Senthilkumaran, 2009			
		1.00	50.12	21	Swapna and Senthilkumaran, 2009			
		1.00	50.12	21	Swapna and Senthilkumaran, 2009			
		1.00	50.12	21	Swapna and Senthilkumaran, 2009			
		1.00	50.12	21	Swapna and Senthilkumaran, 2009			
	NOEC	0.0010	0.050	10	Purdom et al., 1994			
<i>Danio rerio</i>	NOEC	0.0030	0.15	7	Lange et al., 2012			
		0.0093	0.47	7	Lange et al., 2012			
		0.0017	0.085	7	Lange et al., 2012			
		0.010	0.50	10	Purdom et al., 1994			
		5.00	251	30	Ebrahimi, 2007			
		0.00005	0.0025	56	Schulz et al., 2007			
		0.00008	0.0038	322	Wenzel et al., 2001			

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference			
57-63-6	17α-Ethinyl estradiol	Fish	<i>Danio rerio</i>	NOEC	0.0005	0.0025	2	Colman et al., 2009		
					0.0003	0.015	322	Wenzel et al., 2001		
					0.0003	0.015	322	Wenzel et al., 2001		
					0.0004	0.020	88	Xu et al., 2008		
					0.0004	0.020	88	Xu et al., 2008		
					0.0004	0.020	88	Xu et al., 2008		
					0.0005	0.025	56	Schulz et al., 2007		
					0.0005	0.025	56	Schulz et al., 2007		
					0.0005	0.025	56	Schulz et al., 2007		
					0.0058	0.029	2	Colman et al., 2009		
					0.0016	0.078	322	Wenzel et al., 2001		
					0.0016	0.078	322	Wenzel et al., 2001		
					0.0019	0.10	7	Lange et al., 2012		
					0.0020	0.10	88	Xu et al., 2008		
					0.0029	0.15	7	Lange et al., 2012		
					0.047	0.24	2	Colman et al., 2009		
					0.047	0.24	2	Colman et al., 2009		
					0.0050	0.25	14	Reyhani et al., 2011		
					0.0050	0.25	56	Schulz et al., 2007		
					0.0050	0.25	56	Schulz et al., 2007		
					0.0056	0.28	122	Larsen et al., 2009		
					0.0084	0.42	14	Coe et al., 2009		
					0.0084	0.42	14	Coe et al., 2009		
					0.010	0.50	7	Lister et al., 2009		
					0.010	0.50	7	Lister et al., 2009		
					0.010	0.50	7	Lister et al., 2009		
					0.010	0.50	10	Filby et al., 2012		
					0.010	0.50	10	Filby et al., 2012		
					0.010	0.50	10	Filby et al., 2012		
					0.010	0.50	322	Wenzel et al., 2001		
					0.010	0.50	322	Wenzel et al., 2001		
					0.011	0.53	17	Coe et al., 2009		
					0.011	0.53	17	Coe et al., 2009		
					0.025	1.25	14	Reyhani et al., 2011		
					0.025	1.25	14	Reyhani et al., 2011		
					0.10	5.01	14	Stromqvist et al., 2010		
					LOEC	0.0005	0.0025	2	Colman et al., 2009	
						0.00005	0.0025	56	Schulz et al., 2007	
						0.0003	0.015	322	Wenzel et al., 2001	
						0.0005	0.025	56	Schulz et al., 2007	
				0.0058		0.029	2	Colman et al., 2009		
				0.0011		0.055	322	Wenzel et al., 2001		
				0.0016		0.078	322	Wenzel et al., 2001		
				0.0016		0.078	322	Wenzel et al., 2001		
				0.0020		0.10	88	Xu et al., 2008		
				0.0020		0.10	88	Xu et al., 2008		
				0.0020		0.10	88	Xu et al., 2008		
				0.030		0.15	2	Biales et al., 2007		
				0.0039		0.19	122	Micael et al., 2007		
				0.0050		0.25	14	Reyhani et al., 2011		
				0.0050		0.25	14	Reyhani et al., 2011		
				0.0050		0.25	14	Reyhani et al., 2011		
				0.0050		0.25	56	Schulz et al., 2007		
				0.0050		0.25	56	Schulz et al., 2007		
				0.0050		0.25	56	Schulz et al., 2007		
				0.0056		0.28	122	Larsen et al., 2009		
				0.0056		0.28	122	Larsen et al., 2009		
				0.0084		0.42	14	Coe et al., 2009		
				0.0092		0.46	7	Lange et al., 2012		
				0.010		0.50	7	Lister et al., 2009		
				0.010		0.50	10	Filby et al., 2012		
				0.010		0.50	10	Filby et al., 2012		
				0.010		0.50	10	Filby et al., 2012		
				0.010		0.50	10	Filby et al., 2012		
				0.010		0.50	10	Filby et al., 2012		
				0.010		0.50	21	Martyniuk et al., 2007		
				0.010		0.50	88	Xu et al., 2008		
				0.010		0.50	322	Wenzel et al., 2001		
				0.011		0.53	17	Coe et al., 2009		
				0.025		1.25	14	Reyhani et al., 2011		
				0.10		5.01	14	Stromqvist et al., 2010		
				EC50		0.0011	0.055	322	Wenzel et al., 2001	
						0.0062	0.31	21	Van den Belt et al., 2004	
						0.0080	0.40	21	Van den Belt et al., 2004	
						LC50	0.10	5.01	28	Wenzel et al., 2001
							1700	8520	4	Wenzel et al., 2001
					<i>Fundulus heteroclitus</i>	NOEC	0.068	3.40	14	Hogan et al., 2010
							0.25	12.42	14	Hogan et al., 2010
							0.25	12.42	14	Hogan et al., 2010
							0.25	12.42	14	Hogan et al., 2010
							0.25	12.42	14	Hogan et al., 2010
				LOEC	0.068	3.40	14	Hogan et al., 2010		
					0.25	12.42	14	Hogan et al., 2010		
				<i>Gasterosteus aculeatus</i>	NOEC	0.0064	0.032	4	Katsiadaki et al., 2010	
						0.015	0.075	3	Dziewieczynski, 2011	
						0.0018	0.088	28	Mauder et al., 2007	
						0.0018	0.088	28	Mauder et al., 2007	
						0.0018	0.088	28	Mauder et al., 2007	
						0.0018	0.088	28	Mauder et al., 2007	
						0.0018	0.088	28	Mauder et al., 2007	
						0.0018	0.090	7	Lange et al., 2012	
						0.0024	0.12	21	Katsiadaki et al., 2010	
						0.0030	0.15	7	Lange et al., 2012	
						0.0073	0.37	58	Hahlbeck et al., 2004b	
						0.0073	0.37	58	Hahlbeck et al., 2004b	
						0.0073	0.37	58	Hahlbeck et al., 2004b	
						0.010	0.50	58	Hahlbeck, 2004	
						0.010	0.50	58	Hahlbeck, 2004	
						0.010	0.51	28	Bjorkblom et al., 2009	
						0.010	0.51	28	Bjorkblom et al., 2009	
0.010	0.51	28	Bjorkblom et al., 2009							
0.010	0.51	28	Bjorkblom et al., 2009							
0.010	0.51	28	Bjorkblom et al., 2009							

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference			
57-63-6	17α-Ethinyl estradiol	Fish	<i>Gasterosteus aculeatus</i>	NOEC	0.015	0.075	3	Dziewieczynski, 2011		
					0.015	0.075	3	Dziewieczynski, 2011		
					0.079	0.40	4	Katsiadaki et al., 2010		
					0.028	1.39	28	Mauder et al., 2007		
					0.028	1.39	28	Mauder et al., 2007		
					0.050	2.51	42	Le Mer et al., 2013		
					0.050	2.51	58	Hahlbeck et al., 2004a		
					0.050	2.51	58	Hahlbeck, 2004		
					0.10	5.01	58	Hahlbeck, 2004		
					LOEC	0.0010	0.0051	4	Katsiadaki et al., 2010	
					0.015	0.075	3	Dziewieczynski, 2011		
					0.015	0.075	3	Dziewieczynski, 2011		
					0.015	0.075	3	Dziewieczynski, 2011		
					0.015	0.075	3	Dziewieczynski, 2011		
					0.015	0.075	3	Dziewieczynski, 2011		
					0.0018	0.088	28	Mauder et al., 2007		
					0.0018	0.088	28	Mauder et al., 2007		
					0.018	0.088	4	Katsiadaki et al., 2010		
					0.0040	0.20	58	Hahlbeck, 2004		
					0.0041	0.21	21	Katsiadaki et al., 2010		
					0.0073	0.37	58	Hahlbeck et al., 2004b		
					0.0073	0.37	58	Hahlbeck et al., 2004b		
					0.010	0.48	7	Lange et al., 2012		
					0.010	0.51	28	Bjorkblom et al., 2009		
					0.010	0.51	28	Bjorkblom et al., 2009		
					0.028	1.39	28	Mauder et al., 2007		
					0.028	1.39	28	Mauder et al., 2007		
					0.028	1.39	28	Mauder et al., 2007		
					0.028	1.39	28	Mauder et al., 2007		
					0.050	2.51	42	Le Mer et al., 2013		
					0.050	2.51	58	Hahlbeck et al., 2004a		
					0.050	2.51	58	Hahlbeck et al., 2004a		
					0.050	2.51	58	Hahlbeck, 2004		
					0.10	5.01	58	Hahlbeck, 2004		
					<i>Gobiocypris rarus</i>	NOEC	0.10	5.01	21	Ma et al., 2007
					LOEC	0.0002	0.010	21	Ma et al., 2007	
					0.0002	0.010	21	Ma et al., 2007		
					<i>Oncorhynchus mykiss</i>	NOEC	0.0002	0.011	14	Thorpe et al., 2003
					0.0008	0.039	7	Lange et al., 2012		
					0.0010	0.050	10	Purdom et al., 1994		
					0.0011	0.055	14	Thorpe et al., 2003		
					0.0079	0.39	56	Brown et al., 2007		
					0.010	0.50	14	Albertsson et al., 2007		
					0.026	1.30	14	Thorpe et al., 2003		
					0.037	1.85	7	Hook et al., 2006		
					0.060	2.99	56	Brown et al., 2007		
					0.060	2.99	56	Brown et al., 2007		
					LOEC	0.0001	0.0050	10	Purdom et al., 1994	
					0.0025	0.013	1	Biales et al., 2007		
					0.0010	0.050	14	Thorpe et al., 2003		
					0.0030	0.15	7	Lange et al., 2012		
					0.0076	0.38	14	Thorpe et al., 2003		
					0.010	0.50	10	Purdom et al., 1994		
					0.013	0.63	50	Brown et al., 2008		
					0.060	2.99	56	Brown et al., 2007		
					<i>Oryzias latipes</i>	NOEC	0.0002	0.010	14	Thompson, 2000
					0.0002	0.010	14	Tilton et al., 2005		
					0.0002	0.010	14	Tilton et al., 2005		
					0.0002	0.010	21	Ma et al., 2007		
					0.0002	0.010	21	Ma et al., 2007		
					0.010	0.050	1	Biales et al., 2007		
					0.0019	0.10	7	Lange et al., 2012		
					0.0050	0.25	7	Zhang et al., 2008		
					0.0050	0.25	14	Tilton et al., 2005		
					0.0050	0.25	14	Tilton et al., 2005		
					0.0050	0.25	14	Tilton et al., 2005		
					0.0050	0.25	14	Tilton et al., 2005		
					0.020	1.00	21	Ma et al., 2007		
					0.050	2.51	7	Park et al., 2009		
					0.050	2.51	7	Zhang et al., 2008		
					0.050	2.51	7	Zhang et al., 2008		
					0.095	4.75	28	Hano et al., 2007		
					0.095	4.75	28	Hano et al., 2007		
					0.095	4.75	28	Hano et al., 2007		
					0.095	4.75	28	Hano et al., 2007		
					0.10	5.01	21	Ma et al., 2007		
					0.22	10.83	28	Hano et al., 2007		
					0.22	10.83	28	Hano et al., 2007		
					0.22	10.83	28	Hano et al., 2007		
					0.48	24.06	21	Hashimoto et al., 2009		
					0.48	24.06	21	Hashimoto et al., 2009		
					0.50	25.06	7	Park et al., 2009		
					0.50	25.06	7	Park et al., 2009		
					0.50	25.06	7	Park et al., 2009		
					0.50	25.06	7	Park et al., 2009		
					0.50	25.06	7	Park et al., 2009		
					0.50	25.06	7	Zhang et al., 2008		
					0.50	25.06	7	Zhang et al., 2008		
					0.50	25.06	7	Zhang et al., 2008		
					0.50	25.06	7	Zhang et al., 2008		
					0.50	25.06	14	Thompson, 2000		
					0.50	25.06	14	Tilton et al., 2005		
					0.50	25.06	14	Tilton et al., 2005		
					0.52	26.16	28	Hano et al., 2007		
					LOEC	0.0002	0.010	14	Tilton et al., 2005	
					0.0020	0.10	21	Ma et al., 2007		
					0.0020	0.10	21	Ma et al., 2007		
					0.0050	0.25	7	Park et al., 2009		
					0.0050	0.25	7	Park et al., 2009		
					0.0050	0.25	7	Zhang et al., 2008		
0.0050	0.25	14	Thompson, 2000							
0.0050	0.25	14	Tilton et al., 2005							
0.0050	0.25	14	Tilton et al., 2005							

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference
57-63-6	17α-Ethinyl estradiol	Fish	LOEC	0.0092	0.46	7	Lange et al., 2012
				0.10	0.50	1	Biales et al., 2007
				0.050	2.51	7	Zhang et al., 2008
				0.050	2.51	21	Ma et al., 2007
				0.060	3.01	21	Hashimoto et al., 2009
				0.22	10.83	28	Hano et al., 2007
				0.22	10.83	28	Hano et al., 2007
				0.22	10.83	28	Hano et al., 2007
				0.50	25.06	7	Park et al., 2009
				0.50	25.06	7	Zhang et al., 2008
				0.50	25.06	14	Tilton et al., 2005
				0.50	25.06	14	Tilton et al., 2005
				0.50	25.06	14	Tilton et al., 2005
				0.50	25.06	14	Tilton et al., 2005
				0.52	26.16	28	Hano et al., 2007
				0.52	26.16	28	Hano et al., 2007
				0.52	26.16	28	Hano et al., 2007
				0.52	26.16	28	Hano et al., 2007
				0.52	26.16	28	Hano et al., 2007
				0.0054	0.027	2	Martyniuk et al., 2010
				0.0054	0.027	2	Martyniuk et al., 2010
				0.0016	0.049	7	Lange et al., 2012
				0.0016	0.080	14	Ankley et al., 2010
				0.0055	0.27	1095	Palace et al., 2009
				0.0056	0.28	1095	Palace et al., 2009
				0.0056	0.28	1095	Palace et al., 2009
				0.0056	0.28	1095	Palace et al., 2009
				0.0061	0.31	1095	Palace et al., 2009
				0.0061	0.31	1095	Palace et al., 2009
				0.0061	0.31	1095	Palace et al., 2009
				0.0070	0.35	8	Ekman et al., 2008
				0.0070	0.35	8	Ekman et al., 2008
				0.0078	0.39	14	Ankley et al., 2010
				0.010	0.50	17	McGee et al., 2009
				0.010	0.50	21	Filby and Tyler, 2007
				0.010	0.50	21	Panter et al., 2004
				0.010	0.50	21	Panter et al., 2004
				0.010	0.50	21	Salierno and Kane, 2009
				0.010	0.50	21	Salierno and Kane, 2009
				0.011	0.53	21	Filby et al., 2007
				0.011	0.53	21	Filby et al., 2007
				0.011	0.53	21	Filby et al., 2007
				0.011	0.53	21	Filby et al., 2007
				0.011	0.53	21	Filby et al., 2007
				0.020	1.00	21	Salierno and Kane, 2009
				0.020	1.00	21	Salierno and Kane, 2009
				0.040	2.00	21	Salierno and Kane, 2009
				0.040	2.00	21	Salierno and Kane, 2009
				0.050	2.51	15	Weisbrod et al., 2007
				0.050	2.51	15	Weisbrod et al., 2007
		0.050	2.51	15	Weisbrod et al., 2007		
		0.050	2.51	15	Weisbrod et al., 2007		
		0.050	2.51	15	Weisbrod et al., 2007		
		0.050	2.51	15	Weisbrod et al., 2007		
		0.083	4.16	8	Ekman et al., 2008		
		0.083	4.16	8	Ekman et al., 2008		
		0.10	5.01	30	Warner, 2006		
		0.10	50.12	30	Warner, 2006		
		0.0054	0.027	2	Martyniuk et al., 2010		
		0.0054	0.027	2	Martyniuk et al., 2010		
		0.0054	0.027	2	Martyniuk et al., 2010		
		0.0016	0.080	14	Ankley et al., 2010		
		0.0029	0.15	7	Lange et al., 2012		
		0.0055	0.27	1095	Palace et al., 2009		
		0.0056	0.28	1095	Palace et al., 2009		
		0.0056	0.28	1095	Palace et al., 2009		
		0.0061	0.31	1095	Palace et al., 2009		
		0.0061	0.31	1095	Palace et al., 2009		
		0.0070	0.35	8	Ekman et al., 2008		
		0.0078	0.39	14	Ankley et al., 2010		
		0.010	0.50	21	Filby and Tyler, 2007		
		0.010	0.50	21	Panter et al., 2004		
		0.010	0.50	21	Panter et al., 2004		
		0.010	0.50	21	Salierno and Kane, 2009		
		0.010	0.50	21	Salierno and Kane, 2009		
		0.010	0.50	21	Salierno and Kane, 2009		
		0.010	0.50	21	Salierno and Kane, 2009		
		0.011	0.53	21	Filby et al., 2007		
		0.011	0.53	21	Filby et al., 2007		
		0.011	0.53	21	Filby et al., 2007		
		0.011	0.53	21	Filby et al., 2007		
		0.011	0.53	21	Filby et al., 2007		
		0.020	1.00	21	Salierno and Kane, 2009		
		0.040	2.00	21	Salierno and Kane, 2009		
		0.040	2.00	21	Salierno and Kane, 2009		
		0.050	2.51	15	Weisbrod et al., 2007		
		0.050	2.51	15	Weisbrod et al., 2007		
		0.050	2.51	15	Weisbrod et al., 2007		
		0.050	2.51	15	Weisbrod et al., 2007		
		0.083	4.16	8	Ekman et al., 2008		
		0.083	4.16	8	Ekman et al., 2008		
		0.10	5.01	30	Warner, 2006		
		0.10	50.12	30	Warner, 2006		
		0.010	0.50	14	Hallgren and Olsen, 2010		
		0.010	0.50	14	Hallgren and Olsen, 2010		
		0.010	0.50	14	Hallgren and Olsen, 2010		
		2.00	100	112	Shenoy, 2012		
		2.00	100	112	Shenoy, 2012		
		50.00	2506	14	Hallgren and Olsen, 2010		
		0.010	0.50	14	Hallgren and Olsen, 2010		
2.00	100	112	Shenoy, 2012				
2.00	100	112	Shenoy, 2012				
50.00	2506	14	Hallgren and Olsen, 2010				
50.00	2506	14	Hallgren and Olsen, 2010				





CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference			
57-63-6	17α-Ethinyl estradiol	Mollusca	<i>Bithynia tentaculata</i>	NOEC	44.95	2253	284	Hallgren et al., 2012		
					44.95	2253	284	Hallgren et al., 2012		
			<i>Potamopyrgus antipodarum</i>	LOEC	0.0090	0.451	284	Hallgren et al., 2012		
				NOEC	0.025	1.3	28	Sieratowicz et al., 2011		
				LOEC	0.10	5.0	28	Sieratowicz et al., 2011		
				NOEC	0.050	2.5	28	Sieratowicz et al., 2011		
			<i>Radix balthica</i>	NOEC	0.0090	0.451	153	Hallgren et al., 2012		
					5.13	257	153	Hallgren et al., 2012		
					5.13	257	153	Hallgren et al., 2012		
					5.13	257	153	Hallgren et al., 2012		
		Aquatic community	LOEC	44.95	2253	153	Hallgren et al., 2012			
			NOEC	0.0090	0.451	153	Hallgren et al., 2012			
			PNEC	0.00004	0.0018		James et al., 2014			
				0.00004	0.0019		Oekotoxzentrum, Centre Ecotox			
				0.0001	0.0050		James et al., 2014			
				0.0001	0.0050		James et al., 2014			
		50-28-2	17β-estradiol	Algae	<i>Melosira varians</i>	NOEC	20.00	632	10	Julius et al., 2007
							400	12649	10	Julius et al., 2007
						LOEC	800	25298	10	Julius et al., 2007
							80	2530	10	Julius et al., 2007
	NOEC				200	6325	10	Julius et al., 2007		
					100	3162	506	Coady et al., 2004		
<i>Lithobates clamitans</i>	LOEC				100	3162	506	Coady et al., 2004		
	NOEC				100	3162	506	Coady et al., 2004		
<i>Lithobates pipiens</i>	NOEC				100	3162	124	Mackenzie et al., 2003		
					100	3162	124	Mackenzie et al., 2003		
	LOEC			100	316	124	Mackenzie et al., 2003			
				100	316	124	Mackenzie et al., 2003			
	LC50			1242	39279	14	Hogan et al., 2006			
				1517	47978	14	Hogan et al., 2006			
<i>Lithobates sylvaticus</i>	LC50			681	21534	14	Hogan et al., 2006			
	NOEC			100	3162	200	Brodeur et al., 2013			
<i>Rhinella arenarum</i>	LOEC			100	3162	200	Brodeur et al., 2013			
				100	3162	200	Brodeur et al., 2013			
<i>Xenopus laevis</i>	NOEC			100	3162	78	Carr et al., 2003			
	LOEC			100	3162	34	Cong et al., 2006			
		100	3162	34	Cong et al., 2006					
		100	3162	78	Carr et al., 2003					
Crustacea	<i>Americamysis bahia</i>	LC50	1690	5344	2	Hirano et al., 2004				
			890	28144	4	Hirano et al., 2004				
	<i>Ceriodaphnia dubia</i>	NOEC	1000	31623	7	Jukosky et al., 2008a				
		NOEC	200	6325	21	Brennan et al., 2006				
			1000	31623	21	Brennan et al., 2006				
			1000	31623	21	Brennan et al., 2006				
		LOEC	400	12649	21	Brennan et al., 2006				
		EC50	1550	4902	1	Brennan et al., 2006				
			2040	6451	2	Brennan et al., 2006				
			2870	9076	2	Brennan et al., 2006				
<i>Eurytemora affinis</i>	NOEC	2970	9392	2	Hirano et al., 2004					
	LOEC	3670	11606	1	Brennan et al., 2006					
	NOEC	6.00	190	10	Forget-Leray et al., 2005					
	LOEC	18.00	569	10	Forget-Leray et al., 2005					
<i>Neocaridina denticulata</i>	LC50	45.00	1423	4	Forget-Leray et al., 2005					
	LOEC	10.00	316	28	Huang et al., 2006					
		10.00	316	28	Huang et al., 2006					
		10.00	316	28	Huang et al., 2006					
Fish	<i>Cyprinus carpio</i>	NOEC	0.10	3.2	90	Gimeno et al., 1998				
		LOEC	0.10	3.2	90	Gimeno et al., 1998				
			1.00	31.6	90	Gimeno et al., 1998				
		NOEC	0.025	0.079	2	Jin et al., 2009				
			0.013	0.408	8	Rose et al., 2002				
			0.024	0.759	18	Holbech et al., 2006				
			0.024	0.759	18	Holbech et al., 2006				
			0.25	0.791	2	Jin et al., 2009				
			2.50	7.9	3	Jin et al., 2010				
			0.25	7.9	18	Holbech et al., 2006				
	<i>Danio rerio</i>	LOEC	12.50	39.5	3	Jin et al., 2010				
			0.025	0.079	2	Jin et al., 2009				
			0.021	0.664	8	Rose et al., 2002				
			0.054	1.7	18	Holbech et al., 2006				
			0.054	1.7	18	Holbech et al., 2006				
			1.00	31.6	16	Peute et al., 1985				
			12.50	39.5	3	Jin et al., 2010				
		EC50	0.041	1.3	8	Rose et al., 2002				
			0.055	1.7	18	Holbech et al., 2006				
			0.17	5.5	21	Van den Belt et al., 2004				
<i>Gambusia affinis</i>	NOEC	0.24	7.6	21	Van den Belt et al., 2004					
		1.00	31.6	8	Huang et al., 2013					
	LOEC	250	791	3	Kamata et al., 2011					
		1.00	31.6	8	Huang et al., 2012b					
		1.00	31.6	8	Huang et al., 2013					
		500	1581	3	Kamata et al., 2011					
<i>Gambusia holbrooki</i>	NOEC	0.020	0.632	28	Doyle and Lim, 2005					
	LOEC	0.10	3.2	84	Rawson et al., 2006					
		0.10	3.2	28	Doyle and Lim, 2005					
		0.50	15.8	28	Doyle and Lim, 2005					
<i>Gasterosteus aculeatus</i>	NOEC	0.50	15.8	84	Rawson et al., 2006					
		0.010	0.032	7	Hogan et al., 2008					
		0.010	0.316	21	Allen et al., 2008					
		0.010	0.316	58	Hahlbeck et al., 2004a					
		0.020	0.632	21	Allen et al., 2008					
		0.032	1.0	21	Allen et al., 2008					
		10.00	31.6	58	Hahlbeck et al., 2004a					
	LOEC	0.010	0.316	7	Hogan et al., 2008					
		0.050	1.6	21	Allen et al., 2008					
		0.070	2.2	21	Allen et al., 2008					
	0.10	3.2	21	Allen et al., 2008						

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs	Exposure time (days)	Reference			
50-28-2	17β-estradiol	Fish	<i>Gasterosteus aculeatus</i>	LOEC	1.00	3.162	58	Hahlbeck et al., 2004a		
			<i>Gobiocypris rarus</i>	NOEC	0.10	0.32	4	Ma et al., 2009		
				LOEC	0.10	0.32	4	Ma et al., 2009		
			<i>Ictalurus punctatus</i>	NOEC	0.10	3.16	21	Thompson et al., 2000		
				LOEC	1.00	3.162	21	Thompson et al., 2000		
				EC50	0.17	5.38	21	Thompson et al., 2000		
			<i>Morone saxatilis</i>	NOEC	1.00	3.162	21	Thompson et al., 2000		
				LOEC	10.00	3.16	21	Thompson et al., 2000		
				EC50	1.56	49.33	21	Thompson et al., 2000		
			<i>Oncorhynchus mykiss</i>	NOEC	0.0032	0.10	21	Thorpe et al., 2000		
					0.0048	0.15	14	Thorpe et al., 2003		
					0.0096	0.30	14	Thorpe et al., 2003		
					0.10	0.32	5	Ward et al., 2006		
					0.10	3.16	21	Thorpe et al., 2000		
					0.10	3.16	21	Tremblay and Van der Kraak, 1999		
					0.10	3.16	21	Tremblay and Van der Kraak, 1999		
					0.25	7.81	21	Thorpe et al., 2000		
					0.25	7.91	21	Tremblay and Van der Kraak, 1999		
					0.46	14.64	14	Thorpe et al., 2003		
					LOEC	0.0089	0.28	21	Thorpe et al., 2000	
						0.014	0.44	14	Thorpe et al., 2003	
						0.022	0.70	14	Thorpe et al., 2003	
						0.10	3.16	21	Tremblay and Van der Kraak, 1999	
						0.25	7.81	21	Thorpe et al., 2000	
						0.25	7.91	21	Tremblay and Van der Kraak, 1999	
						0.25	7.91	21	Tremblay and Van der Kraak, 1999	
					EC50	0.015	0.47	21	Thorpe et al., 2000	
				<i>Oncorhynchus tshawytscha</i>	LOEC	400	12649	8 h immersion	Piferrer and Donaldson, 1992	
				<i>Oryzias latipes</i>	NOEC	0.0010	0.0032		Lee et al., 2012	
						0.0010	0.0032		Lee et al., 2012	
						0.010	0.032	5	Kang et al., 2005	
						0.10	3.16	21	Thompson et al., 2000	
						1.00	3.16		Lee et al., 2012	
						0.15	4.86	14	Jukosky et al., 2008b	
						0.23	7.18	25	Kang et al., 2002	
						10.00	3.162	5	Kang et al., 2006b	
						2.53	79.95	14	Jukosky et al., 2008b	
					LOEC	0.010	0.032	5	Kang et al., 2005	
						0.010	0.032		Lee et al., 2012	
						0.010	0.032		Lee et al., 2012	
						0.0050	0.16	21	Kashiwada et al., 2002	
						0.10	0.32		Lee et al., 2012	
						0.029	0.93	21	Kang et al., 2002	
						0.056	1.76	21	Kang et al., 2002	
						0.056	1.78	14	Jukosky et al., 2008b	
						1.00	3.16		Lee et al., 2012	
						1.00	3.16		Lee et al., 2012	
						0.46	14.64	25	Kang et al., 2002	
						10.00	3.162	5	Kang et al., 2006b	
						1.00	3.162	21	Thompson et al., 2000	
						2.53	79.95	14	Jukosky et al., 2008b	
					EC50	0.20	6.32	21	Thompson et al., 2000	
						0.23	7.12	14	Sun et al., 2009	
						470	14863	21	Kashiwada et al., 2002	
					LC50	460	1455	3	Kashiwada et al., 2002	
						460	1455	3	Tabata et al., 2001	
						2000	6325	4	Kang et al., 2002	
						3500	11068	3	Kashiwada et al., 2002	
						3500	11068	3	Tabata et al., 2001	
				<i>Pimephales promelas</i>	NOEC	0.010	0.32	14	Thorpe et al., 2007	
						0.027	0.85	9	Cline et al., 2003	
						0.028	0.89	17	McCree et al., 2009	
						0.030	0.95	21	Schultz et al., 2012	
						0.030	0.95	21	Schultz et al., 2012	
						0.030	0.95	21	Schultz et al., 2012	
						0.030	0.95	21	Schultz et al., 2012	
						0.030	0.95	21	Schultz et al., 2012	
						0.030	0.95	21	Schultz et al., 2012	
						0.10	3.16	14	Thorpe et al., 2007	
						0.50	15.81	21	Bringolf et al., 2004	
						0.50	15.81	21	Bringolf et al., 2004	
					LOEC	0.022	0.70	14	Thorpe et al., 2007	
						0.030	0.95	21	Schultz et al., 2012	
						0.27	8.54	9	Cline et al., 2003	
						0.50	15.81	21	Bringolf et al., 2004	
						0.50	15.81	21	Bringolf et al., 2004	
						0.50	15.81	21	Bringolf et al., 2004	
					EC50	0.025	0.79	14	Brian et al., 2005	
						0.12	3.79	19	Kramer et al., 1998	
						0.25	7.94	19	Kramer et al., 1998	
					LC50	1.15	36.37	19	Kramer et al., 1998	
				<i>Poecilia reticulata</i>	NOEC	0.050	1.58	120	Nielsen and Baatrup, 2006	
						1.00	3.162	21	Li and Wang, 2005	
					LOEC	1.00	3.162	21	Li and Wang, 2005	
						1.00	3.162	21	Li and Wang, 2005	
				<i>Pomatoschistus minutus</i>	LOEC	0.071	2.25	243	Robinson et al., 2004	
					EC50	0.087	2.75	243	Robinson et al., 2004	
						0.13	4.02	243	Robinson et al., 2004	
						0.17	5.22	243	Robinson et al., 2004	
				<i>Salmo trutta</i>	NOEC	0.015	0.48	8	Bjerregaard et al., 2008	
					LOEC	0.020	0.63	8	Bjerregaard et al., 2008	
					EC50	0.015	0.48	8	Bjerregaard et al., 2008	
				<i>Thymallus thymallus</i>	LOEC	0.0010	0.032	50	Lahnsteiner et al., 2006	
						0.0010	0.032	50	Lahnsteiner et al., 2006	
				Fish		LC50	2740	8665	Nendza and Wenzel, 2006	
				Invertebrates	<i>Brachionus calyciflorus</i>	LOEC	0.0010	0.032	10	Huang et al., 2012a
							100	3.162	10	Huang et al., 2012a
				Mollusca	<i>Elliptio complanata</i>	NOEC	100	3.16	0.25	Flynn et al., 2013
						LOEC	100	3.16	0.25	Flynn et al., 2013
					Aquatic community	PNEC	0.0004	0.013		Oekotoxentrum, Centre Ecotox
				0.0010	0.032		Gross-Sorokin et al., 2006			
				0.0010	0.032		Young et al., 2004			

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference
50-28-2	17β-estradiol	Aquatic community	PNEC	0.0020	0.063		Anderson et al., 2012 ; Caldwell et al., 2012
				0.0023	0.072		Yuan et al., 2014
1689-82-3	4-hydroxyazobenzene	Algae	EC50	9600	606	Not specified	Nendza and Wenzel, 2006
		Bacteria	EC50	930	58.68	Not specified	Nendza and Wenzel, 2006
		Fish	LC50	1170	73.82	Not specified	Nendza and Wenzel, 2006
14938-35-3	4-n-amyphenol	Algae	NOEC	980	49116	3	Ramos et al., 1999
			LOEC	2300	115273	3	Ramos et al., 1999
			EC50	2600	130309	3	Ramos et al., 1999
		Crustacea	EC50	1330	66658	2	Ramos et al., 1998
				2040	102242	1	Ramos et al., 1998
		Fish	LC50	1250	62648	4	Ramos et al., 1998
				1360	68161	3	Ramos et al., 1998
				1920	96228	2	Ramos et al., 1998
				2490	124796	1	Ramos et al., 1998
		Mollusca	LC50	3700	185439	3	Ramos et al., 1998
				4600	230546	2	Ramos et al., 1998
				5380	269639	1	Ramos et al., 1998
				3710	1859405	4	Ramos et al., 1998
1806-26-4	4-n-octylphenol	Amphibia	NOEC	206	41.17	7	Crump et al., 2002
				206	41.17	10	Crump et al., 2002
			LC50	578	115	7	Crump et al., 2002
		Fish	LOEC	2.00	0.40	98	Knorr and Braunbeck, 2002
				20.00	3.99	98	Knorr and Braunbeck, 2002
140-66-9	4-tert-octylphenol	Crustacea	EC50	11.00	0.87	2	Ra et al., 2008
		Fish	NOEC	12.00	9.53	185	Wenzel et al., 2001
				12.00	9.53	185	Wenzel et al., 2001
				35.00	27.80	185	Wenzel et al., 2001
				35.00	27.80	185	Wenzel et al., 2001
				35.00	27.80	185	Wenzel et al., 2001
			LOEC	35.00	27.80	185	Wenzel et al., 2001
				35.00	27.80	185	Wenzel et al., 2001
			EC50	28.00	22.24	185	Wenzel et al., 2001
			LC50	370	29.39	4	Wenzel et al., 2001
15972-60-8	Alachlor	Algae	NOEC	100	15.85	4	Garten and Frank, 1984
			LOEC	1000	158	4	Garten and Frank, 1984
		Algae	NOEC	0.35	0.055	5	U.S. EPA, 2013
			LOEC	10.00	1.58	4	Garten and Frank, 1984
			EC50	1.64	0.26	5	U.S. EPA, 2013
		Algae	NOEC	5.35	0.85	28	Carder et al., 1998
		Algae		78.55	12.45	28	Carder et al., 1998
		Algae	LOEC	78.55	12.45	28	Carder et al., 1998
		Crustacea	NOEC	5600	88.75	2	U.S. EPA, 2013
				12000	190	2	U.S. EPA, 2013
				14000	222	2	U.S. EPA, 2013
				18000	285	2	U.S. EPA, 2013
			LOEC	230	36.45	21	U.S. EPA, 2013
				430	68.15	21	U.S. EPA, 2013
				1700	269	21	U.S. EPA, 2013
		Fish	NOEC	2600	4121	4	U.S. EPA, 2013
			LC50	3900	6181	4	U.S. EPA, 2013
			LC50	6500	103	4	U.S. EPA, 2013
			NOEC	1800	28.53	4	U.S. EPA, 2013
				3700	58.64	4	U.S. EPA, 2013
				4200	66.57	4	U.S. EPA, 2013
				5600	88.75	4	U.S. EPA, 2013
			LC50	2800	44.38	4	U.S. EPA, 2013
				6200	98.26	4	U.S. EPA, 2013
				6400	101	4	U.S. EPA, 2013
				7600	120	4	U.S. EPA, 2013
				12400	197	4	U.S. EPA, 2013
			NOEC	1000	15.85	4	U.S. EPA, 2013
				1800	28.53	4	U.S. EPA, 2013
				2400	38.04	4	U.S. EPA, 2013
				2400	38.04	4	U.S. EPA, 2013
			LOEC	388	61.49	96	U.S. EPA, 2013
				390	61.81	96	U.S. EPA, 2013
			LC50	240	3.80	4	U.S. EPA, 2013
				1800	28.53	4	U.S. EPA, 2013
				3600	57.06	4	U.S. EPA, 2013
				3700	58.64	4	U.S. EPA, 2013
				4200	66.57	4	U.S. EPA, 2013
1912-24-9	Atrazine	Algae	EC50	50.00	0.050		Nendza and Wenzel, 2006
		Bacteria	EC50	24200	24.20		Nendza and Wenzel, 2006
		Crustacea	EC50	240	0.24		Nendza and Wenzel, 2006
		Fish	LC50	6300	6.30		Nendza and Wenzel, 2006
71-43-2	Benzene	Crustacea	NOEC	98000	19554	21	LeBlanc and Surprenant, 1980
				98000	19554	21	LeBlanc and Surprenant, 1980
		Fish	LC50	21639	432	4	Hodson et al., 1984
		Aquatic community	PNEC	8.00	1.60		OSPAR Agreement, 2014-05
				46.00	9.18		U.S. EPA, 1996
50-32-8	Benzo [a] pyrene	Algae	NOEC	4000	400	3	Schoeny et al., 1988
			EC50	4000	400	3	Schoeny et al., 1988
			NOEC	4000	400	3	Schoeny et al., 1988
			EC50	1300	130	3	Schoeny et al., 1988
			NOEC	4000	400	3	Schoeny et al., 1988
			EC50	4000	400	3	Schoeny et al., 1988
			NOEC	4000	400	3	Schoeny et al., 1988
			EC50	4000	400	3	Schoeny et al., 1988
			NOEC	4000	400	3	Schoeny et al., 1988
			EC50	4000	400	3	Schoeny et al., 1988
			NOEC	4000	400	3	Schoeny et al., 1988
			EC50	5.00	0.50	3	Schoeny et al., 1988
			NOEC	4000	400	3	Schoeny et al., 1988
			EC50	15.00	1.50	3	Schoeny et al., 1988
			LC50	400	40.00	1	Warsawskyyet al., 1995
				400	400	6	Warsawskyyet al., 1995
		Algae	EC50	5.00	0.50	Not specified	Nendza and Wenzel, 2006
		Amphibia	LOEC	10.00	1.00	6	Reynaud et al., 2012
			NOEC	500	500	16	Jaylet et al., 1986
			LOEC	125	125	12	Mouchet et al., 2006
			NOEC	500	50.00	6	Marquiset al., 2009
			LOEC	50.00	5.00	6	Marquiset al., 2009



CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference				
50-32-8	Benzo [a] pyrene	Amphibia	<i>Xenopus laevis</i>	EC50	8700	870	4	Propst et al., 1997			
				LC50	13400	1340	4	Propst et al., 1997			
			Crustacea	<i>Daphnia magna</i>	NOEC	25.00	25.00	14	Atienzar et al., 1999		
					LOEC	0.020	0.0020	1	Ha and Choi, 2009		
				<i>Eurytemora affinis</i>	NOEC	12.00	12.00	10	Forget-Leray et al., 2005		
					LOEC	12.00	12.00	10	Forget-Leray et al., 2005		
					LC50	58.00	58.00	4	Forget-Leray et al., 2005		
					<i>Gammarus duebeni</i>	LC50	11.00	1.10	2	Lawrence and Poulter, 1998	
						LC50	371	37.10	1	Lawrence and Poulter, 1998	
					Fish	<i>Palaemonetes pugio</i>	LC50	102	102	4	Weinstein and Garner, 2008
		NOEC	0.76	0.76			60	Chang et al., 2005			
		<i>Cyprinus carpio</i>	LOEC	0.76		0.76	60	Chang et al., 2005			
			NOEC	63.08		6.31	3	Weight et al., 2011			
		<i>Danio rerio</i>	NOEC	252		25.23	1	Jonsson et al., 2009			
			LOEC	252		25.23	3	Kazeto et al., 2004			
			LC50	2523		252	3	Kazeto et al., 2004			
			LC50	2523		2.52	1	Jonsson et al., 2009			
			LC50	2523		252	3	Kazeto et al., 2004			
			EC50	131		13.12	3	Weight et al., 2011			
		<i>Oncorhynchus kisutch</i>	LC50	1287	129	3	Weight et al., 2011				
			NOEC	1.00	0.10	7	Hook et al., 2006				
		<i>Oncorhynchus mykiss</i>	LOEC	0.25	0.025	1	Ostrander et al., 1988				
			LOEC	1.00	0.10	7	Hook et al., 2006				
		Insecta	<i>Oryzias latipes</i>	NOEC	25.23	2.52	1	Jonsson et al., 2009			
				NOEC	10.00	10.00	16	Shugart et al., 1991			
			<i>Chironomus riparius</i>	LC50	1.00	0.10	Not specified	Nendza and Wenzel, 2006			
				LOEC	10.00	1.00	1	Ha and Choi, 2008a			
			<i>Chironomus tentans</i>	LC50	31590	3159	1	Ha and Choi, 2008a			
				NOEC	500	50.00	1	Lee et al., 2006			
			<i>Chironomus tentans</i>	LOEC	5.00	0.50	1	Lee et al., 2006			
				LC50	9873	987	1	Ha and Choi, 2008b			
			<i>Physella acuta</i>	LOEC	5.00	5.00	7	Sanchez-Arguello et al., 2012			
				LOEC	20.00	20.00	7	Sanchez-Arguello et al., 2012			
		Mollusca	Aquatic community	LOEC	40.00	40.00	7	Sanchez-Arguello et al., 2012			
				PNEC	0.0002	0.0002		OSPAR Agreement, 2014-05			
		207-08-9	Benzo(k)fluoranthene	Crustacea	<i>Daphnia magna</i>	EC50	140	0.44	0.5	EU Risk Assessment Report, 2008	
						NOEC	300	94.87	5	Newsted and Giesy, 1987	
					Fish	<i>Fundulus heteroclitus</i>	LOEC	300	94.87	5	Clark et al., 2010
							NOEC	300	94.87	5	Clark, 2010
							NOEC	300	94.87	5	Clark, 2010
Aquatic community	PNEC			0.0002	0.0005		OSPAR Agreement, 2014-05				
	PNEC			0.017	0.054		EU Risk Assessment Report, 2008				
80-05-7	Bisphenol-A			Algae	EC50	1000	200	Not specified	Nendza and Wenzel, 2006		
				Crustacea	EC50	7050	1407	Not specified	Nendza and Wenzel, 2006		
				Fish	LC50	6010	1199	Not specified	Nendza and Wenzel, 2006		
85-68-7	Butyl benzyl phthalate	Aquatic community	PNEC	0.002	0.0033		Yuan et al., 2014				
		Algae	EC50	150	2.99	Not specified	Oekotoxzentrum, Centre Ecotox				
		Crustacea	EC50	190	2.39	Not specified	Nendza and Wenzel, 2006				
133-06-2	Captan	Crustacea	<i>Daphnia magna</i>	EC50	1640	20.65	Not specified	Nendza and Wenzel, 2006			
				LC50	1250	15.74	Not specified	Nendza and Wenzel, 2006			
		Algae	<i>Chlorella pyrenoidosa</i>	NOEC	6020	1904	4	Anton, 1993			
				EC50	44500	14072	4	Anton, 1993			
			<i>Selenastrum capricornutum</i>	EC50	2400	75.89	3	Kikuchi, 1993			
				NOEC	3125	9.88	12	Mouchet et al., 2006			
			<i>Pleurodeles waltl</i>	NOEC	125	39.53	12	Mouchet et al., 2006			
				NOEC	125	39.53	12	Mouchet et al., 2006			
				LOEC	62.50	19.76	12	Mouchet et al., 2006			
				LOEC	250	79.06	12	Mouchet et al., 2006			
NOEC	3125	9.88		12	Mouchet et al., 2006						
LOEC	62.50	19.76		12	Mouchet et al., 2006						
Fish	<i>Carassius auratus</i>	LC50	15.60	4.93	12	Mouchet et al., 2006					
		LC50	125	39.53	12	Mouchet et al., 2006					
	<i>Danio rerio</i>	LC50	890	28.14	4	Anton, 1993					
		EC50	358	11.34	5	Padilla et al., 2012					
	<i>Ictalurus punctatus</i>	LC50	77.50	2.45	4	Johnson and Finley, 1980					
		LC50	141	4.46	4	Johnson and Finley, 1980					
	<i>Lepomis macrochirus</i>	LC50	56.40	1.78	4	Johnson and Finley, 1980					
		LC50	138	4.36	4	Johnson and Finley, 1980					
	<i>Oncorhynchus mykiss</i>	LC50	73.20	2.31	4	Johnson and Finley, 1980					
		LC50	56.50	1.79	4	Johnson and Finley, 1980					
<i>Oncorhynchus tshawytscha</i>	LC50	500	15.81	2	Tsuji et al., 1986						
	LC50	610	19.29	2	Tsuji et al., 1986						
	LC50	800	25.30	2	Tsuji et al., 1986						
	LC50	120	3.79	4	Johnson and Finley, 1980						
	LC50	200	6.32	4	Johnson and Finley, 1980						
	LC50	80.00	2.53	4	Johnson and Finley, 1980						
63-25-2	Carbaryl	Algae	EC50	49.00	1.55	4	Johnson and Finley, 1980				
		Bacteria	EC50	4000	126	Not specified	Nendza and Wenzel, 2006				
		Crustacea	EC50	636	20.11	Not specified	Nendza and Wenzel, 2006				
10605-21-7	Carbendazim	Fish	<i>Daphnia magna</i>	EC50	6.40	0.20	Not specified	Nendza and Wenzel, 2006			
				LC50	8870	280	Not specified	Nendza and Wenzel, 2006			
		Algae	NOEC	330	0.66	28	Van den Brink et al., 2000				

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs	Exposure time (days)	Reference		
10605-21-7	Carbendazim	Algae	<i>Chara sp.</i>	NOEC	1000	2.00	28	Van den Brink et al., 2000	
			<i>Chlomonas sp.</i>	NOEC	330	0.66	28	Van den Brink et al., 2000	
			<i>Chlamydomonas sp.</i>	NOEC	100	0.20	28	Van den Brink et al., 2000	
			<i>Chlorella pyrenoidosa</i>	EC50	340	0.068	2	Canton, 1976	
					34650	69.14	4	Ma et al., 2002a	
			<i>Cryptomonas sp.</i>	NOEC	100	0.20	28	Van den Brink et al., 2000	
		<i>Cyclotella sp.</i>	NOEC	100	0.20	28	Van den Brink et al., 2000		
		<i>Epithemia sp.</i>	NOEC	1000	2.00	28	Van den Brink et al., 2000		
		<i>Monoraphidium sp.</i>	NOEC	1000	2.00	28	Van den Brink et al., 2000		
		<i>Oedogonium sp.</i>	NOEC	1000	2.00	28	Van den Brink et al., 2000		
		<i>Scenedesmus acutus</i>	NOEC	1000	2.00	28	Van den Brink et al., 2000		
		<i>Scenedesmus obliquus</i>	EC50	19050	38.01	4	Ma et al., 2002a		
		<i>Stephanodiscus sp.</i>	NOEC	100	0.20	28	Van den Brink et al., 2000		
		Amphibia	<i>Xenopus laevis</i>	NOEC	191	0.038	4	Yoon et al., 2008	
				LOEC	382	0.076	4	Yoon et al., 2008	
				LC50	574	0.11	4	Yoon et al., 2008	
		Crustacea	<i>Acroperus harpae</i>	NOEC	1072	0.21	4	Yoon et al., 2008	
				NOEC	3.30	0.0066	28	Van den Brink et al., 2000	
		Algae	<i>Alona rectangula</i>	NOEC	33.00	0.066	21	Daam and Van den Brink, 2007	
			<i>Alonella exigua</i>	NOEC	33.00	0.066	28	Van den Brink et al., 2000	
			<i>Cyclopoida</i>	NOEC	100	0.20	28	Van den Brink et al., 2000	
			<i>Daphnia magna</i>	NOEC	10.00	0.0020	1	Canton, 1976	
					60.00	0.12	1	Ferreira et al., 2008	
					33.00	0.066	28	Van den Brink et al., 2000	
					100	0.20	4	Van den Brink et al., 2000	
					70.00	0.014	1	Ferreira et al., 2008	
					EC50	3.50	0.0007	1	Ferreira et al., 2008
					20.00	0.0040		Canton, 1976	
					22.90	0.0046	1	Ferreira et al., 2008	
					24.40	0.0049	1	Ferreira et al., 2008	
					28.20	0.0056	2	Ferreira et al., 2008	
					28.60	0.0057	1	Ferreira et al., 2008	
					54.10	0.011	2	Ferreira et al., 2008	
					68.70	0.014	2	Ferreira et al., 2008	
					73.10	0.015	2	Ferreira et al., 2008	
					97.54	0.019	1	Ferreira et al., 2008	
					103	0.021	2	Ferreira et al., 2008	
					137	0.027	1	Ferreira et al., 2008	
					145	0.029	2	Ferreira et al., 2008	
					157	0.031	2	Ferreira et al., 2008	
					37.00	0.074	28	Van den Brink et al., 2000	
					460	0.092	2	Canton, 1976	
					113	0.23	4	Van den Brink et al., 2000	
				<i>Graptoleberis testudinaria</i>	NOEC	33.00	0.066	21	Daam and Van den Brink, 2007
				<i>Macrobrachium ferreirai</i>	LC50	16767	33.45	4	Rico et al., 2011
				<i>Simocephalus vetulus</i>	NOEC	33.00	0.066	21	Daam and Van den Brink, 2007
			Fish	<i>Colossoma macropomum</i>	LC50	33.00	0.066	28	Van den Brink et al., 2000
					LC50	4162	0.83	4	Rico et al., 2011
		LC50			3690	0.74	4	Rico et al., 2011	
		LC50			4138	0.83	4	Rico et al., 2011	
		LC50			1800	0.36	2	Canton, 1976	
		LC50			Otocinclus affinis	4238	0.85	4	Rico et al., 2011
		LC50			Paracheirodon axelrodi	1648	0.33	4	Rico et al., 2011
		NOEC			Fusarium sporotrichioides	2000	3.99	14	Dijksterhuis et al., 2011
		NOEC			Trichoderma hamatum	260	0.52	14	Dijksterhuis et al., 2011
		Insecta			<i>Kiefferulus calligaster</i>	NOEC	1700	0.34	3
				1700		0.34	3	Domingues et al., 2009	
				15000		2.99	3	Domingues et al., 2009	
				15000		2.99	3	Domingues et al., 2009	
				15000		2.99	3	Domingues et al., 2009	
	1700		3.39	6		Domingues et al., 2009			
	5000		1.00	3		Domingues et al., 2009			
	5000		1.00	3		Domingues et al., 2009			
	1700		3.39	6		Domingues et al., 2009			
	5000		9.98	6		Domingues et al., 2009			
Invertebrates	<i>Colurella uncinata</i>	NOEC	100	0.20	21	Daam and Van den Brink, 2007			
		NOEC	33.00	0.066	21	Daam and Van den Brink, 2007			
		NOEC	330	0.66	28	Van den Brink et al., 2000			
		NOEC	330	0.66	28	Van den Brink et al., 2000			
		NOEC	33.00	0.066	21	Daam and Van den Brink, 2007			
		NOEC	330	0.66	28	Van den Brink et al., 2000			
Mollusca	<i>Lepadella patella</i>	NOEC	33.00	0.066	21	Daam and Van den Brink, 2007			
		NOEC	330	0.66	28	Van den Brink et al., 2000			
		NOEC	330	0.66	28	Van den Brink et al., 2000			
		NOEC	330	0.66	28	Van den Brink et al., 2000			
		NOEC	330	0.66	28	Van den Brink et al., 2000			
		NOEC	330	0.66	28	Van den Brink et al., 2000			
Plants	<i>Pomacea doliooides</i>	LC50	1758576	3509	4	Rico et al., 2011			
		LC50	73822	14.73	3	Rico et al., 2011			
		NOEC	10000	19.95	21	Belgers et al., 2009			
		EC50	9743	19.44	21	Belgers et al., 2009			
		NOEC	10000	19.95	21	Belgers et al., 2009			
		LC50	80669	16.10	4	Rico et al., 2011			
		NOEC	10000	19.95	21	Belgers et al., 2009			
		LC50	11329	22.21	4	Rico et al., 2011			
		NOEC	10000	19.95	21	Belgers et al., 2009			
		PNEC	Aquatic community	0.34	0.0007		Oekotoxzentrum, Centre Ecotox		
50-22-6	Corticosterone	Amphibia	<i>Xenopus laevis</i>	LOEC	0.57	0.0011		Oekotoxzentrum, Centre Ecotox	
					34.65	43.62	21	Lorenz et al., 2009	
					34.65	43.62	21	Lorenz et al., 2009	
84-74-2	Dibutylphthalate	Algae	<i>Xenopus laevis</i>	EC50	173	2.8	21	Lorenz et al., 2009	
				EC50	1200	12.00	Not specified	Nendza and Wenzel, 2006	
				EC50	10900	109	Not specified	Nendza and Wenzel, 2006	
Crustacea	<i>Daphnia magna</i>	EC50	3400	34.00	Not specified	Nendza and Wenzel, 2006			
		LC50	980	9.80	Not specified	Nendza and Wenzel, 2006			
		EC50	0.031	0.16	4	Huang et al., 1996			
683-18-1	Dibutyltin dichloride	Algae	<i>Scenedesmus acutus</i>	EC50	0.031	0.16	4	Huang et al., 1996	
				EC50	900	451	1	Vighi and Calamari, 1985	
		Crustacea	<i>Daphnia magna</i>	EC50	900	451	1	Vighi and Calamari, 1985	
				NOEC	48.61	244	110	De Vries et al., 1991	
		Fish	<i>Oncorhynchus mykiss</i>	NOEC	48.61	244	110	De Vries et al., 1991	
					48.61	244	110	De Vries et al., 1991	
					48.61	244	110	De Vries et al., 1991	
					48.61	244	110	De Vries et al., 1991	
					48.61	244	110	De Vries et al., 1991	
					48.61	244	110	De Vries et al., 1991	
	LOEC	243	12.18	110	De Vries et al., 1991				
		243	12.18	110	De Vries et al., 1991				
		243	12.18	110	De Vries et al., 1991				
		243	12.18	110	De Vries et al., 1991				



CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference				
115-29-7	Endosulfan	Crustacea	<i>Hyalella curvispina</i>	LC50	17.20	5.44	2	Mugni et al., 2012			
			<i>Mesocyclops longisetus</i>	NOEC	0.0040	0.0013	1	Gutierrez et al., 2013			
				LOEC	0.0060	0.0019	1	Gutierrez et al., 2013			
			<i>Palaemonetes pugio</i>	LOEC	0.0040	0.0013	1	Gutierrez et al., 2013			
				LC50	0.0060	0.0019	1	Gutierrez et al., 2013			
					0.016	0.0051	2	Gutierrez et al., 2013			
					0.020	0.0063	2	Gutierrez et al., 2013			
				NOEC	0.18	0.56	4	Pennington, 2002			
				LOEC	0.52	1.65	4	Pennington, 2002			
				LC50	0.37	1.16	4	Pennington, 2002			
				0.51	1.62	4	Pennington, 2002				
				0.55	1.74	4	Pennington, 2002				
				1.02	3.23	4	Pennington, 2002				
		<i>Penaeus aztecus</i>	LC50	0.24	0.076	2	U.S. EPA, 2013				
			<i>Uca pugilator</i>	NOEC	2.99	9.45	4	Pennington, 2002			
				LC50	100	31.62	2	U.S. EPA, 2013			
				3.34	10.56	4	Pennington, 2002				
				790	250	2	U.S. EPA, 2013				
			Fish	<i>Bidyanus bidyanus</i>	NOEC	10.00	3.16	0.75	Patra et al., 2009		
				<i>Channa punctata</i>	NOEC	8.10	25.61	90	Sarma et al., 2009		
						8.10	25.61	90	Sarma et al., 2009		
					LOEC	8.10	25.61	90	Sarma et al., 2009		
					8.10	25.61	90	Sarma et al., 2009			
				8.10	25.61	90	Sarma et al., 2009				
				8.10	25.61	90	Sarma et al., 2009				
		<i>Cnesterodon decemmaculatus</i>		NOEC	0.0040	0.0013	1	Gutierrez et al., 2013			
				0.0060	0.0019	1	Gutierrez et al., 2013				
				1.00	0.32	4	Mugni et al., 2012				
		<i>Cyprinodon variegatus</i>	LOEC	0.0060	0.0019	1	Gutierrez et al., 2013				
			NOEC	0.60	0.19	7	Hemmer et al., 2011				
				0.27	0.85	28	U.S. EPA, 2013				
		<i>Cyprinus carpio</i>	LOEC	0.60	1.90	28	U.S. EPA, 2013				
			NOEC	0.50	0.16	7	U.S. EPA, 2013				
				0.90	0.28	4	U.S. EPA, 2013				
		<i>Fundulus heteroclitus</i>	LC50	0.90	0.28	7	U.S. EPA, 2013				
				2.20	0.70	4	U.S. EPA, 2013				
			NOEC	0.52	0.16	4	Pennington, 2002				
		<i>Labeo rohita</i>	LOEC	2.99	0.94	4	Pennington, 2002				
			LC50	2.23	0.71	4	Pennington, 2002				
				2.55	0.81	4	Pennington, 2002				
		<i>Leostomus xanthurus</i>	LC50	400	126	4	Alam et al., 2010				
				500	158	4	Alam et al., 2010				
				750	237	4	Alam et al., 2010				
		<i>Lepomis macrochirus</i>		1100	348	4	Alam et al., 2010				
			LC50	0.32	0.10	2	U.S. EPA, 2013				
			NOEC	1.00	0.32	4	U.S. EPA, 2013				
		<i>Morone saxatilis</i>		1.80	0.57	4	U.S. EPA, 2013				
			LC50	1.70	0.54	4	U.S. EPA, 2013				
				2.08	0.66	4	U.S. EPA, 2013				
		<i>Mugil cephalus</i>		3.90	1.23	4	U.S. EPA, 2013				
			LC50	5.60	1.77	4	U.S. EPA, 2013				
				1000	3.16	4	U.S. EPA, 2013				
		<i>Oncorhynchus kisutch</i>	LC50	0.32	0.10	2	U.S. EPA, 2013				
			NOEC	10.00	3.16	0.020833333	Tierney et al., 2006				
				100	31.62	0.020833333	Tierney et al., 2006				
		<i>Oncorhynchus mykiss</i>	LOEC	100	31.62	0.02	Tierney et al., 2006				
			NOEC	0.32	0.10	4	U.S. EPA, 2013				
				1.00	0.32	4	U.S. EPA, 2013				
				1.80	0.57	4	U.S. EPA, 2013				
			LC50	0.37	0.12	4	U.S. EPA, 2013				
				0.47	0.15	4	U.S. EPA, 2013				
				0.83	0.26	4	U.S. EPA, 2013				
				2.30	0.73	4	U.S. EPA, 2013				
				2.70	0.85	4	U.S. EPA, 2013				
		<i>Pimephales promelas</i>		28.00	8.85	4	U.S. EPA, 2013				
			NOEC	0.030	0.095	260	U.S. EPA, 2013				
			LOEC	0.030	0.095	260	U.S. EPA, 2013				
				0.11	0.35	260	U.S. EPA, 2013				
				0.11	0.35	260	U.S. EPA, 2013				
				0.46	1.45	260	U.S. EPA, 2013				
		Insecta	<i>Enallagma cyathigerum</i>	LOEC	50.00	15.81	1	Janssens and Stoks, 2012			
				50.00	15.81	1	Janssens and Stoks, 2012				
		Mollusca	<i>Crassostrea virginica</i>	NOEC	0.10	0.032	2	U.S. EPA, 2013			
				2.99	9.45	4	Pennington, 2002				
			EC50	460	145	2	U.S. EPA, 2013				
		Plants	<i>Bidens laevis</i>	LC50	3.34	10.56	4	Pennington, 2002			
				NOEC	0.50	0.16	2	Perez et al., 2011a			
					10.00	3.16	2	Perez et al., 2011a			
			100	31.62	2	Perez et al., 2011a					
			5.00	1.58	2	Perez et al., 2011a					
			50.00	15.81	2	Perez et al., 2011a					
							U.S. EPA, 1996				
		13311-84-7	Flutamide	Crustacea	Aquatic community	PNEC	0.051	0.16			
					<i>Acartia tonsa</i>	EC50	480	480	5	Andersen et al., 2001	
					<i>Daphnia magna</i>	LC50	5400	540	2	Andersen et al., 2001	
						NOEC	100	100	21	Haeba et al., 2008	
							1000	1000	21	Haeba et al., 2008	
						LOEC	1000	1000	21	Haeba et al., 2008	
						EC50	2700	270	2	Haeba et al., 2008	
						7800	780	1	Haeba et al., 2008		
					Fish	<i>Neomysis integer</i>	LC50	1380	1380	4	Verslycke et al., 2004
						<i>Danio rerio</i>	NOEC	100	10.00	7	Andersen et al., 2003
				LOEC			100	10.00	7	Andersen et al., 2003	
				<i>Gasterosteus aculeatus</i>		LOEC	50.00	50.00	21	Jolly et al., 2009	
						NOEC	32.00	3.20	4	León et al., 2007	
				<i>Oryzias latipes</i>			1560	1560	21	Kang et al., 2006a	
							1560	1560	21	Kang et al., 2006a	
						LOEC	32.00	3.20	4	León et al., 2007	
							320	32.00	4	León et al., 2007	
							320	32.00	4	León et al., 2007	
	202				202	21	Kang et al., 2006a				
	1560			1560	21	Kang et al., 2006a					



CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs	Exposure time (days)	Reference		
118-74-1	Hexachlorobenzene	Fish	<i>Lepomis macrochirus</i>	NOEC	2800	88.54	4	U.S. EPA, 2013	
				LC50	3400	108	4	U.S. EPA, 2013	
					7600	240	4	U.S. EPA, 2013	
			<i>Oncorhynchus mykiss</i>	NOEC	1000	3162	4	U.S. EPA, 2013	
				LC50	2300	72.73	4	U.S. EPA, 2013	
13311-84-7	Flutamide	Fish	<i>Pimephales promelas</i>	NOEC	4.80	1.52	32	Carlson et al., 1987	
					4.80	1.52	32	Carlson et al., 1987	
			<i>Oryzias latipes</i>	LC50	1920	192	4	León et al., 2007	
				NOEC	320	320	21	Filby et al., 2007	
					939	939	21	Panter et al., 2004	
					939	939	21	Panter et al., 2004	
					939	939	21	Panter et al., 2004	
				LOEC	50.00	5.00	2	Garcia-Reyero et al., 2009	
					50.00	5.00	2	Garcia-Reyero et al., 2009	
					53.00	5.30	2	Martyniuk et al., 2009	
					500	50.00	2	Garcia-Reyero et al., 2009	
					95.40	95.40	21	Panter et al., 2004	
					320	320	21	Filby et al., 2007	
					320	320	21	Filby et al., 2007	
			330-55-2	Linuron	Invertebrates	<i>Brachionus calyciflorus</i>	NOEC	0.10	0.10
	LOEC	1.00				1.00	4	Preston et al., 2000	
<i>Anabaena flosaquae</i>	NOEC	12.80				8.08	5	U.S. EPA, 2013	
	EC50	38.80				24.48	5	U.S. EPA, 2013	
<i>Chlorella pyrenoidosa</i>	LOEC	1.00				0.063	0.5	Thomas et al., 1973	
	EC50	110				6.94	15	Kratky and Warren, 1971	
		130				8.20	15	Kratky and Warren, 1971	
	EC50	13.70				8.64	5	U.S. EPA, 2013	
	EC50	6.00				0.38	3	Snel et al., 1998	
		17.30				1.09	3	Snel et al., 1998	
	NOEC	0.25			0.016	1	Junghans et al., 2006		
	EC50	19.93			1.26	1	Junghans et al., 2006		
	NOEC	20.00			12.62	5	U.S. EPA, 2013		
	EC50	67.00			42.27	5	U.S. EPA, 2013		
	NOEC	5.35			3.38	5	U.S. EPA, 2013		
	EC50	35.90			22.65	5	U.S. EPA, 2013		
	EC50	25.10			15.84	35	Snel et al., 1998		
Bacteria	Crustacea	<i>Spirogyra sp.</i>			EC50	5500	347	30 min	Hernando et al., 2003
		<i>Vibrio fischeri</i>			NOEC	1200	757	4	U.S. EPA, 2013
		<i>Americanysis bahia</i>			LOEC	297	187	28	U.S. EPA, 2013
						582	367	28	U.S. EPA, 2013
					LC50	3300	2082	4	U.S. EPA, 2013
					NOEC	100	6.31	2	U.S. EPA, 2013
						130	82.02	21	U.S. EPA, 2013
						130	82.02	21	U.S. EPA, 2013
			LOEC	130	82.02	21	U.S. EPA, 2013		
				240	151	21	U.S. EPA, 2013		
				240	151	21	U.S. EPA, 2013		
			EC50	120	7.57	2	U.S. EPA, 2013		
				310	19.56	1	Stephenson and Kane, 1984		
				360	22.71	1	Stephenson and Kane, 1984		
				590	37.23	1	Stephenson and Kane, 1984		
		1100	69.41	2	U.S. EPA, 2013				
		1910	121	2	U.S. EPA, 2013				
		3000	189	15 min	Martins et al., 2007				
		7000	447	4	Hernando et al., 2003				
		330	20.82	1	Stephenson and Kane, 1984				
Fish		<i>Cyprinodon variegatus</i>	NOEC	498	31.42	4	U.S. EPA, 2013		
			LOEC	357	225	35	U.S. EPA, 2013		
				760	480	35	U.S. EPA, 2013		
				760	480	35	U.S. EPA, 2013		
				766	483	35	U.S. EPA, 2013		
			LC50	890	56.16	4	U.S. EPA, 2013		
			EC50	8895	561	5	Padilla et al., 2012		
		<i>Danio rerio</i>	LOEC	100	63.10	21	Jolly et al., 2009		
		<i>Gasterosteus aculeatus</i>	NOEC	4900	309	4	U.S. EPA, 2013		
				5600	353	4	U.S. EPA, 2013		
				7500	473	4	U.S. EPA, 2013		
			LC50	9200	580	4	U.S. EPA, 2013		
				9600	606	4	U.S. EPA, 2013		
				16200	1022	4	U.S. EPA, 2013		
			<i>Oncorhynchus mykiss</i>	NOEC	2090	132	4	U.S. EPA, 2013	
		5600	353	4	U.S. EPA, 2013				
	LOEC	30.00	18.93	28	Bruggemann et al., 1995				
		40.00	25.24	80	U.S. EPA, 2013				
		60.00	37.86	28	Bruggemann et al., 1995				
		390	246	80	U.S. EPA, 2013				
		3085	195	4	U.S. EPA, 2013				
	LC50	16400	1035	4	U.S. EPA, 2013				
Mollusca		<i>Crassostrea virginica</i>	NOEC	3600	227	2	U.S. EPA, 2013		
			EC50	5400	341	2	U.S. EPA, 2013		
		<i>Ceratophyllum demersum</i>	EC50	8.70	5.49	35	Snel et al., 1998		
		<i>Chara globularis</i>	EC50	12.10	0.76	1	Snel et al., 1998		
		<i>Elodea nuttallii</i>	EC50	13.40	0.85	1	Snel et al., 1998		
				10.50	6.63	56	Snel et al., 1998		
		<i>Lemna gibba</i>	NOEC	9.65	6.09	14	U.S. EPA, 2013		
			LOEC	19.93	12.57	7	Hulsen et al., 2002		
			EC50	27.30	17.23	14	U.S. EPA, 2013		
			EC50	11.80	0.74	1	Snel et al., 1998		
	EC50	12.90	0.81	1	Snel et al., 1998				
	EC50	13.20	0.83	1	Snel et al., 1998				
	Aquatic community	PNEC	0.26	0.16		Oekotoxzentrum, Centre Ecotox			
			1.37	0.86		Oekotoxzentrum, Centre Ecotox			
65277-42-1	Ketoconazole	Crustacea	<i>Daphnia magna</i>	NOEC	1000	100	21	Haeba et al., 2008	
				EC50	150	15.10	2	Haeba et al., 2008	
					8100	8100	1	Haeba et al., 2008	
					400	40.00	21	Villeneuve et al., 2007	
					10740	33963	1	Pickford and Morris, 1999	
84371-65-3	Mifepristone	Fish	<i>Pimephales promelas</i>	LOEC	400	40.00	21	Villeneuve et al., 2007	
			<i>Xenopus laevis</i>	NOEC	10740	33963	1	Pickford and Morris, 1999	
			<i>Danio rerio</i>	NOEC	107	340	4	Hillegass et al., 2008	

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference				
84371-65-3 104-40-5	Mifepristone Nonylphenol	Fish	<i>Danio rerio</i>	NOEC	108	341	4	Hillegass et al., 2008			
		Algae		EC50	410	4100	Not specified	Nendza and Wenzel, 2006			
		Bacteria	<i>Vibrio fischeri</i>	EC50	1300	130	Not specified	Nendza and Wenzel, 2006			
		Crustacea	<i>Daphnia magna</i>	EC50	220	22.00	Not specified	Nendza and Wenzel, 2006			
		Fish		LC50	140	14.00	Not specified	Nendza and Wenzel, 2006			
84852-15-3	Nonylphenol technical mixture	Fish	<i>Danio rerio</i>	PNEC	0.30	0.30		OSPAR Agreement, 2014-05			
				NOEC	2.50	0.13	3	Jin et al., 2010			
					10.00	0.50	2	Jin et al., 2009			
					12.50	0.63	3	Jin et al., 2010			
					100	5.01	2	Jin et al., 2009			
					1000	50.12	2	Kammann et al., 2009			
					1000	50.12	2	Kammann et al., 2009			
					1200	60.14	2	Kammann et al., 2009			
					1400	70.17	2	Kammann et al., 2009			
					1500	75.18	2	Kammann et al., 2009			
					2000	100	2	Kammann et al., 2009			
					LOEC	10.00	0.50	2	Jin et al., 2009		
						12.50	0.63	3	Jin et al., 2010		
						100	5.01	2	Jin et al., 2009		
						2000	100	2	Kammann et al., 2009		
						2100	105	2	Kammann et al., 2009		
						4400	221	2	Kammann et al., 2009		
						<i>Oncorhynchus mykiss</i>	NOEC	2.30	0.12	4	Shelley et al., 2012
								18.00	0.90	4	Shelley et al., 2012
								18.00	0.90	4	Shelley et al., 2012
				18.00	0.90	4	Shelley et al., 2012				
				18.00	0.90	4	Shelley et al., 2012				
				18.00	0.90	4	Shelley et al., 2012				
				18.00	0.90	4	Shelley et al., 2012				
298-00-0	Parathion-methyl	Algae		EC50	5000	500	Not specified	Nendza and Wenzel, 2006			
		Bacteria	<i>Vibrio fischeri</i>	EC50	510	5100	Not specified	Nendza and Wenzel, 2006			
		Crustacea	<i>Daphnia magna</i>	EC50	0.14	0.014	Not specified	Nendza and Wenzel, 2006			
		Fish		LC50	5400	540	Not specified	Nendza and Wenzel, 2006			
57465-28-8	PCB 126	Fish	<i>Fundulus heteroclitus</i>	LOEC	1.00	0.10	5	Clark et al., 2010			
					1.00	0.10	5	Clark, 2010			
					1.00	0.10	5	Clark, 2010			
					0.15	0.015	until 3 dph	Kim and Cooper, 1998			
					0.15	0.015	until 3 dph	Kim and Cooper, 1998			
			EC50	0.17	0.017	until 3 dph	Kim and Cooper, 1999				
				0.43	0.043	until 3 dph	Kim and Cooper, 1999				
				0.45	0.045	until 3 dph	Kim and Cooper, 1999				
				LC50	0.25	0.025	until 3 dph	Kim and Cooper, 1999			
		2310-17-0	Phosalone	Polyp	<i>Hydra vulgaris</i>	NOEC	1000	1000	4	Becker, 1991	
Algae	<i>Selenastrum capricornutum</i>			EC50	830	4160	3	Graff et al., 2003			
					930	46.61	3	Graff et al., 2003			
Fish	<i>Charina orientalis</i>			LC50	81.00	4.06	4	Verma et al., 1978b			
	<i>Danio rerio</i>			EC50	3443	173	5	Padilla et al., 2012			
114-26-1	Propoxur	Algae	<i>Selenastrum capricornutum</i>	EC50	10000	3162		Nendza and Wenzel, 2006			
302-79-4	Retinoic acid	Crustacea	<i>Daphnia magna</i>	NOEC	100	100	>7	Peterson et al., 2001			
					800	800		Wang et al., 2005			
					0.60	0.060	2	Teixido et al., 2013			
					3.00	0.30	2	Teixido et al., 2013			
					LOEC	0.30	0.030	2	Teixido et al., 2013		
				1.50	0.15	2	Teixido et al., 2013				
				3004351	300435	2	Elo et al., 2007				
				0.42	0.042	6	Selderslaghs et al., 2012				
				1.48	0.15	3	Selderslaghs et al., 2012				
				3.58	0.36	2	Selderslaghs et al., 2012				
				5.71	0.57	2	Teixido et al., 2013				
				234	23.37	1	Selderslaghs et al., 2012				
				LC50	15.32	1.53	2	Teixido et al., 2013			
					40.26	4.03	6	Selderslaghs et al., 2012			
					442	44.16	3	Selderslaghs et al., 2012			
			865	86.52	2	Selderslaghs et al., 2012					
			2121	212	1	Selderslaghs et al., 2012					
122320-73-4 688-73-3	Rosiglitazone Tributyltin hydride	Fish	<i>Danio rerio</i>	LOEC	357	1.13	2	Elo et al., 2007			
		Fish	<i>Salmo salar</i>	NOEC	0.25	0.025	7	Greco et al., 2007			
		Invertebrates	<i>Brachionus calyciflorus</i>	LOEC	0.050	0.0050	7	Greco et al., 2007			
					0.25	0.025	7	Greco et al., 2007			
					19.00	1.90	1	Snell, 1991			
					NOEC	0.019	0.019	21	Giusti et al., 2013		
						0.094	0.094	21	Giusti et al., 2013		
		Mollusca	<i>Lymnaea stagnalis</i>	LOEC	0.094	0.094	21	Giusti et al., 2013			
					0.20	0.20	180	Tillmann et al., 2001			
					NOEC	0.81	2.56	4	Orvos et al., 2002		
					EC50	0.97	3.07	4	Orvos et al., 2002		
						1.20	3.79	4	U.S. EPA, 2013		
3380-34-5	Triclosan (Irgasan)	Algae	<i>Anabaena flosaquae</i>		1.60	5.06	4	Orvos et al., 2002			
					3.55	11.23	4	DeLorenzo et al., 2008			
					EC50	16.00	50.60	4	U.S. EPA, 2013		
						19.10	60.40	4	Orvos et al., 2002		
					NOEC	40.00	12.65	3	U.S. EPA, 2013		
			<i>Scenedesmus subspicatus</i>	EC50	120	37.95	3	U.S. EPA, 2013			
					NOEC	0.20	0.063	3	Yang et al., 2008		
					0.50	0.16	3	Orvos et al., 2002			
					0.69	2.18	4	Orvos et al., 2002			
					2.50	7.91	4	U.S. EPA, 2013			
			<i>Selenastrum capricornutum</i>	LOEC	0.40	0.13	3	Yang et al., 2008			
					1.20	0.38	3	Orvos et al., 2002			
					EC50	0.53	0.17	3	Yang et al., 2008		
					0.70	0.22	3	Orvos et al., 2002			
					2.90	0.92	3	Orvos et al., 2002			
					4.70	1.49	3	Tatarazako et al., 2004			
					1.40	4.43	4	Orvos et al., 2002			
					3.40	10.75	4	U.S. EPA, 2013			
					4.46	14.10	4	Orvos et al., 2002			
					0.10	0.32	24	Fraker and Smith, 2004			
					230	727	25	Fraker and Smith, 2004			
		Amphibia	<i>Lithobates pipiens</i>	LOEC	0.10	0.32	24	Fraker and Smith, 2004			
					230	727	25	Fraker and Smith, 2004			

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference	
3380-34-5	Triclosan (Irgasan)	Amphibia	<i>Xenopus laevis</i>	NOEC	200	632	14	Matsumura et al., 2005
				EC50	200	632	14	Matsumura et al., 2005
		Bacteria	<i>Vibrio fischeri</i>	EC50	150	47.43	15 min	Tatarazako et al., 2004
				NOEC	53000	16760	15 min	DeLorenzo et al., 2008
				EC50	2800000	88544	15-30 min	Farre et al., 2008
				EC50	220	696	7	Tatarazako et al., 2004
				NOEC	100	3162	2	U.S. EPA, 2013
				NOEC	190	56.92	2	U.S. EPA, 2013
				NOEC	240	75.89	2	U.S. EPA, 2013
				NOEC	240	75.89	2	U.S. EPA, 2013
				NOEC	40.00	126	21	Orvos et al., 2002
				NOEC	100	316	6	Flaherty and Dodson, 2005
		Crustacea	<i>Ceriodaphnia dubia</i>	EC50	220	696	7	Tatarazako et al., 2004
				NOEC	100	3162	2	U.S. EPA, 2013
				NOEC	190	56.92	2	U.S. EPA, 2013
				NOEC	240	75.89	2	U.S. EPA, 2013
				NOEC	240	75.89	2	U.S. EPA, 2013
				NOEC	100	316	6	Flaherty and Dodson, 2005
				LOEC	10.00	3162	30	Flaherty and Dodson, 2005
				NOEC	200	632	21	Orvos et al., 2002
				EC50	390	123	2	Orvos et al., 2002
				EC50	390	123	2	U.S. EPA, 2013
		Fish	<i>Daphnia magna</i>	NOEC	420	133	2	U.S. EPA, 2013
				NOEC	420	133	2	U.S. EPA, 2013
				EC50	250	791	10	Dussault et al., 2008a
				LC50	200	632	10	Dussault et al., 2008a
				LC50	470	149	1	Kim et al., 2009
				LC50	220	696	9	Tatarazako et al., 2004
				NOEC	18000	5692	4	U.S. EPA, 2013
				LC50	370	117	4	Orvos et al., 2002
				LC50	410	130	2	Orvos et al., 2002
				LC50	440	139	1	Orvos et al., 2002
		Fish	<i>Danio rerio</i>	NOEC	36200	11447	4	U.S. EPA, 2013
				NOEC	100	3162	4	U.S. EPA, 2013
				NOEC	34.10	108	70	Orvos et al., 2002
				NOEC	18000	5692	4	U.S. EPA, 2013
				LOEC	71.30	225	70	Orvos et al., 2002
				LC50	288	9107	4	U.S. EPA, 2013
				LC50	23400	7400	4	U.S. EPA, 2013
				NOEC	200	632	21	Ishibashi et al., 2004
				NOEC	20.00	63.25	21	Ishibashi et al., 2004
				NOEC	20.00	63.25	21	Ishibashi et al., 2004
		Fish	<i>Lepomis macrochirus</i>	NOEC	100	316	21	Ishibashi et al., 2004
				NOEC	200	632	21	Ishibashi et al., 2004
				NOEC	200	632	21	Ishibashi et al., 2004
				NOEC	313	990	14	Ishibashi et al., 2004
				EC50	170	538	9	Nassef et al., 2010
LC50	350			111	2	Foran et al., 2000		
LC50	399			126	4	Ishibashi et al., 2004		
LC50	600			190	4	Kim et al., 2009		
LC50	602			190	4	Ishibashi et al., 2004		
LC50	400			1265	14	Tatarazako et al., 2004		
Fish	<i>Oncorhynchus mykiss</i>	NOEC	160	5.06	12	Schultz et al., 2012		
		NOEC	160	5.06	21	Schultz et al., 2012		
		NOEC	160	5.06	21	Schultz et al., 2012		
		NOEC	100	3162	4	U.S. EPA, 2013		
		LOEC	160	5.06	21	Schultz et al., 2012		
		LC50	250	79.06	4	U.S. EPA, 2013		
		LC50	260	82.22	4	Orvos et al., 2002		
		LC50	270	85.38	2	Orvos et al., 2002		
		LC50	270	85.38	3	Orvos et al., 2002		
		LC50	360	114	1	Orvos et al., 2002		
Insecta	<i>Chironomus dilutus</i>	EC50	280	885	10	Dussault et al., 2008a		
		LC50	400	1265	10	Dussault et al., 2008a		
		PNEC	0.020	0.063		Oekotoxzentrum, Centre Ecotox		
		EC50	1060	3352	5	U.S. EPA, 2013		
		NOEC	2540	8032	5	U.S. EPA, 2013		
		EC50	1020	3226	5	U.S. EPA, 2013		
		EC50	870	2751	5	U.S. EPA, 2013		
		NOEC	580	1824	4	U.S. EPA, 2013		
		LC50	1800	5692	4	U.S. EPA, 2013		
		NOEC	1000	316	2	U.S. EPA, 2013		
Algae	<i>Selenastrum capricornutum</i>	NOEC	3000	949	1	Haeba et al., 2008		
		NOEC	3000	949	2	Haeba et al., 2008		
		NOEC	790	2498	21	U.S. EPA, 2013		
		NOEC	790	2498	21	U.S. EPA, 2013		
		NOEC	790	2498	21	U.S. EPA, 2013		
		NOEC	1000	3162	21	Haeba et al., 2008		
		LOEC	1000	3162	21	Haeba et al., 2008		
		LOEC	1400	4427	21	U.S. EPA, 2013		
		LOEC	1400	4427	21	U.S. EPA, 2013		
		LOEC	1400	4427	21	U.S. EPA, 2013		
Crustacea	<i>Skeletonema costatum</i>	EC50	2400	7589	21	U.S. EPA, 2013		
		EC50	3650	1154	2	U.S. EPA, 2013		
		NOEC	1100	348	4	U.S. EPA, 2013		
		EC50	10741	3396	5	Padilla et al., 2012		
		LOEC	100	316	21	Jolly et al., 2009		
		NOEC	68100	21535	4	U.S. EPA, 2013		
		LC50	49800	15748	4	U.S. EPA, 2013		
		LC50	47500	15021	4	U.S. EPA, 2013		
		NOEC	1040	329	4	U.S. EPA, 2013		
		NOEC	1800	569	4	U.S. EPA, 2013		
Fish	<i>Cyprinodon variegatus</i>	NOEC	3160	999	4	U.S. EPA, 2013		
		NOEC	2840	898	4	U.S. EPA, 2013		
		NOEC	13600	4301	4	U.S. EPA, 2013		
		NOEC	2500	7906	100	Kiparissis et al., 2003		
		NOEC	2500	7906	100	Kiparissis et al., 2003		
		NOEC	2500	7906	100	Kiparissis et al., 2003		
		NOEC	50.00	158	175	U.S. EPA, 2013		
		NOEC	50.00	158	175	U.S. EPA, 2013		
		NOEC	50.00	158	175	U.S. EPA, 2013		
		NOEC	450	1423	21	Martinovic et al., 2008		
Fish	<i>Danio rerio</i>	NOEC	700	2214	21	Makynen et al., 2000		
		NOEC	700	2214	21	Makynen et al., 2000		
		NOEC	1200	3795	34	Makynen et al., 2000		
		NOEC	1200	3795	34	Makynen et al., 2000		
		NOEC	60.00	190	21	Martinovic et al., 2008		
		NOEC	60.00	190	21	Martinovic et al., 2008		
		NOEC	100	316	21	Villeneuve et al., 2007		
		NOEC	150	474	175	U.S. EPA, 2013		
		NOEC	150	474	175	U.S. EPA, 2013		
		NOEC	150	474	175	U.S. EPA, 2013		
Fish	<i>Gasterosteus aculeatus</i>	NOEC	255	806	21	Martinovic et al., 2008		
		NOEC	255	806	21	Martinovic et al., 2008		
		NOEC	450	1423	21	Martinovic et al., 2008		
		NOEC	450	1423	21	Martinovic et al., 2008		
		NOEC	450	1423	21	Martinovic et al., 2008		
		NOEC	700	2214	21	Makynen et al., 2000		
		NOEC	700	2214	21	Makynen et al., 2000		
		NOEC	1200	3795	34	Makynen et al., 2000		
		NOEC	1200	3795	34	Makynen et al., 2000		
		NOEC	60.00	190	21	Martinovic et al., 2008		
Fish	<i>Lepomis gibbosus</i>	NOEC	3120	987	1	Zavala-Aguirre et al., 2007		
		LOEC	6250	1976	1	Zavala-Aguirre et al., 2007		
		LC50	30500	9645	1	Zavala-Aguirre et al., 2007		
		NOEC	2100	6641	4	U.S. EPA, 2013		
		EC50	3200	10119	4	U.S. EPA, 2013		
		LOEC	0.030	0.095	140	Tillmann et al., 2001		
		LOEC	5000	15811	21	Sanchez-Arguello et al., 2012		
		NOEC	2400	7589	14	U.S. EPA, 2013		
		EC50	900	285	5	U.S. EPA, 2013		
		LOEC	10000	31623	6	Prescott et al., 1977		
Mollusca	<i>Crassostrea virginica</i>	NOEC	100	316	21	U.S. EPA, 2013		
		NOEC	150	474	175	U.S. EPA, 2013		
		NOEC	150	474	175	U.S. EPA, 2013		
		NOEC	150	474	175	U.S. EPA, 2013		
		NOEC	255	806	21	Martinovic et al., 2008		
		NOEC	255	806	21	Martinovic et al., 2008		
		NOEC	450	1423	21	Martinovic et al., 2008		
		NOEC	450	1423	21	Martinovic et al., 2008		
		NOEC	450	1423	21	Martinovic et al., 2008		
		NOEC	700	2214	21	Makynen et al., 2000		
Plants	<i>Physella acuta</i>	NOEC	700	2214	21	Makynen et al., 2000		
		NOEC	700	2214	21	Makynen et al., 2000		
		NOEC	1200	3795	34	Makynen et al., 2000		
		NOEC	3120	987	1	Zavala-Aguirre et al., 2007		
		LOEC	6250	1976	1	Zavala-Aguirre et al., 2007		
		LC50	30500	9645	1	Zavala-Aguirre et al., 2007		
		NOEC	2100	6641	4	U.S. EPA, 2013		
		EC50	3200	10119	4	U.S. EPA, 2013		
		LOEC	0.030	0.095	140	Tillmann et al., 2001		
		LOEC	5000	15811	21	Sanchez-Arguello et al., 2012		
Protozoa	<i>Acanthamoeba castellanii</i>	NOEC	2400	7589	14	U.S. EPA, 2013		
		EC50	900	285	5	U.S. EPA, 2013		
		LOEC	10000	31623	6	Prescott et al., 1977		

C: DR CALUX

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (pg/L)	Exposure time (days)	Reference			
1746-016	2,3,7,8-TCDD	Amphibia	<i>Pseudacris triseriata</i>	NOEC	30000000	3000000	2	Collier et al., 2008		
				LOEC	30000000	3000000	2	Collier et al., 2008		
				LOEC	3000000	300000	2	Collier et al., 2008		
			<i>Xenopus laevis</i>	NOEC	30000000	3000000	3	Collier et al., 2008		
				LOEC	3000000	300000	2	Collier et al., 2008		
				LOEC	30000000	3000000	2	Collier et al., 2008		
			Crustacea	<i>Daphnia magna</i>	NOEC	1000000	1000000	2	Wu et al., 2001	
					LOEC	1030000	103000	2	Adams et al., 1986	
					LOEC	100	100	8	Wu et al., 2001	
			Fish	<i>Esox lucius</i>	LOEC	100	10.00	4	Helder, 1980	
					LOEC	100	10.00	4	Helder, 1980	
					LOEC	1100	110	4	Helder, 1980	
				<i>Fundulus heteroclitus</i>	LC50	20000000	2000000	2h	Toomey et al., 2001	
					LOEC	2.00	2.00	120	Wu et al., 2001	
					LOEC	2.00	2.00	chronic	Wu et al., 2001	
		<i>Gobiocypris rarus</i>		LOEC	1000	100	2	Wu et al., 2001		
				LOEC	1000	1000	120	Wu et al., 2001		
				LOEC	1000	1000	chronic	Wu et al., 2001		
				<i>Oryzias latipes</i>	NOEC	3400	340	until 3 dph	Kim and Cooper, 1998	
					LOEC	12000	12000	10	Wisk and Cooper, 1992	
					LOEC	12000	12000	10	Wisk and Cooper, 1992	
					EC50	1200	120	3	Chen and Cooper, 1999	
					LOEC	2200	2200	11 dph	Wisk and Cooper, 1990b	
					LOEC	2800	280	4	Kim and Cooper, 1998	
			LOEC		3500	350	until 3 dph	Wisk and Cooper, 1990b		
			LOEC		5600	560	until 3 dph	Kim and Cooper, 1999		
			LOEC		6000	600	3	Wisk and Cooper, 1990a		
		LOEC	10100		1010	until 3 dph	Kim and Cooper, 1999			
		LOEC	12500		1250	until 3 dph	Kim and Cooper, 1999			
		LOEC	14000		1400	until 3 dph	Wisk and Cooper, 1990b			
		LOEC	14000	1400	until 3 dph	Wisk and Cooper, 1990b				
		LOEC	15000	1500	3	Wisk and Cooper, 1990a				
		LOEC	15800	1580	until 3 dph	Chen and Cooper, 1999				
		LOEC	18400	1840	until 3 dph	Chen and Cooper, 1999				
		LOEC	26800	2680	until 3 dph	Chen and Cooper, 1999				
		LC50	5700	5700	17	Metcalfe et al., 1997				
		LOEC	8100	810	until 3 dph	Kim and Cooper, 1999				
		LOEC	9000	900	until 3 dph	Wisk and Cooper, 1990b				
		LOEC	13000	1300	3	Wisk and Cooper, 1990a				
		LOEC	13500	1350	until 3 dph	Chen and Cooper, 1999				
		Pimphales promelas	NOEC	700	700	28	Adams et al., 1986			
			NOEC	3800	380	1	Olivieri and Cooper, 1997			
			NOEC	3800	380	1	Olivieri and Cooper, 1997			
			LOEC	10160	1016	until 2 dph	Olivieri and Cooper, 1997			
			LOEC	370	37.00	until 2 dph	Olivieri and Cooper, 1997			
LC50	1700		1700	28	Adams et al., 1986					
NOEC	34000		3400	2	Walker et al., 1991					
LOEC	1000		100	2	Spitsbergen et al., 1991					
LOEC	10000		1000	2	Spitsbergen et al., 1991					
LOEC	55000		5500	2	Walker et al., 1991					
LOEC	55000		5500	2	Walker et al., 1991					
LOEC	226000		22600	2	Walker et al., 1991					
LC50	65000		6500	2	Walker et al., 1991					
NOEC	200000		200000	17	Miller et al., 1973					
207-08-9	Benzo(k)fluoranthene		Insecta	<i>Aedes aegypti</i>	NOEC	200000	200000	17	Miller et al., 1973	
		LOEC			200000	200000	36	Miller et al., 1973		
		LOEC			200000	200000	55	Miller et al., 1973		
		Mollusca	<i>Physa sp.</i>	LOEC	200000	200000	36	Miller et al., 1973		
				LOEC	200000	200000	55	Miller et al., 1973		
				LOEC	200000	200000	55	Miller et al., 1973		
		Crustacea	<i>Daphnia magna</i>	EC50	1400000	702	0.5	Newsted and Giesy, 1987		
				NOEC	300000000	150356	5	Clark et al., 2010		
				LOEC	300000000	150356	5	Clark et al., 2010		
			Fish	<i>Fundulus heteroclitus</i>	NOEC	300000000	150356	5	Clark, 2010	
					LOEC	300000000	150356	5	Clark, 2010	
					LOEC	300000000	150356	5	Clark, 2010	
				Aquatic community	PNEC	170	0.85		OSPAR Agreement, 2014-05	
					PNEC	1700	85.20		EU Risk Assessment Report, 2008	
					PNEC	1700	85.20		EU Risk Assessment Report, 2008	
50-02-2	Dexamethasone	Algae	<i>Selenastrum capricornutum</i>	NOEC	10000000000	10000000	3	DellaGreca et al., 2004		
				LOEC	39246100	39246	21	Lorenz et al., 2009		
				LOEC	39246100	39246	21	Lorenz et al., 2009		
				LOEC	39246100	39246	21	Lorenz et al., 2009		
				LOEC	196230500	196230	21	Lorenz et al., 2009		
		Crustacea	<i>Ceriodaphnia dubia</i>	EC50	50000000	50000	7	DellaGreca et al., 2004		
				EC50	48300000000	4830000	1	DellaGreca et al., 2004		
				LC50	60110000000	6011000	1	DellaGreca et al., 2004		
				NOEC	39246100	3925	5	Gustafson et al., 2012		
				NOEC	392461000	39246	5	Gustafson et al., 2012		
			Fish	<i>Danio rerio</i>	NOEC	392461000	39246	5	Gustafson et al., 2012	
					NOEC	3924610000	392461	5	Gustafson et al., 2012	
					LOEC	39246100000	3924610	4	Sun et al., 2010a	
					LOEC	500000000	500000	21	Lalone et al., 2012	
					LOEC	13736135000	1373614	4	Sun et al., 2010a	
		50-02-2	Dexamethasone	Fish	<i>Danio rerio</i>	LOEC	10000000000	10000000	3	Hillegass et al., 2007
						LOEC	10000000000	10000000	3	Hillegass et al., 2008
						NOEC	254000000	254000	30 dph	Overturf et al., 2012
						NOEC	160000000	160000	31 dph	Overturf et al., 2012
						LOEC	100000	100	21	Lalone et al., 2012
				<i>Pimphales promelas</i>	NOEC	50000000	50000	21	Lalone et al., 2012	
					NOEC	500000000	500000	21	Lalone et al., 2012	
					NOEC	500000000	500000	29	Lalone et al., 2012	
					NOEC	500000000	500000	29	Lalone et al., 2012	
					LC50	577000000	577000	29 dph	Overturf et al., 2012	
Invertebrates	<i>Brachionus calyciflorus</i>	LC50	254000000	254000	28 dph	Overturf et al., 2012				
		LC50	48220000000	4822000	1	DellaGreca et al., 2004				
		LOEC	1000000	31623	5	Clark et al., 2010				
		LOEC	1000000	31623	5	Clark, 2010				
		LOEC	1000000	31623	5	Clark, 2010				
57465-28-8	PCB 126	Fish	<i>Fundulus heteroclitus</i>	NOEC	150000	4743	until 3 dph	Kim and Cooper, 1998		
				NOEC	150000	4743	until 3 dph	Kim and Cooper, 1998		
				EC50	170000	5376	until 3 dph	Kim and Cooper, 1999		
				EC50	430000	13598	until 3 dph	Kim and Cooper, 1999		
				EC50	450000	14230	until 3 dph	Kim and Cooper, 1999		
		Polyp	<i>Hydra vulgaris</i>	LC50	250000	7906	until 3 dph	Kim and Cooper, 1999		
				NOEC	100000000	316227766	4	Becker, 1991		

D: GR CALUX

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference		
52-39-1	Aldosterone	Amphibia	<i>Xenopus laevis</i>	NOEC	36044	288	21	Lorenz et al., 2009	
					36044	288	21	Lorenz et al., 2009	
					36044	288	21	Lorenz et al., 2009	
2242-98-0	Cortisol	Crustacea Fish	<i>Daphnia magna</i> <i>Danio rerio</i>	NOEC	10000	7000	6	Kashian and Dodson, 2004	
				LOEC	100000000	700000	3	Hillegass et al., 2007	
					100000000	700000	3	Hillegass et al., 2008	
50-02-2	Dexamethasone	Algae	<i>Selenastrum capricornutum</i>	NOEC	800000	56000	14	Brodeur et al., 2005	
				NOEC	100000000	10000000	3	DellaGreca et al., 2004	
				LOEC	39246	39246	21	Lorenz et al., 2009	
					39246	39246	21	Lorenz et al., 2009	
					39246	39246	21	Lorenz et al., 2009	
					196231	196231	21	Lorenz et al., 2009	
		Crustacea	<i>Ceriodaphnia dubia</i>	EC50	50000	50000	7	DellaGreca et al., 2004	
				EC50	48300000	48300000	1	DellaGreca et al., 2004	
				LC50	60100000	60100000	1	DellaGreca et al., 2004	
			Fish	<i>Thamnocephalus platyurus</i> <i>Danio rerio</i>	NOEC	39246	3925	5	Gustafson et al., 2012
						392461	39246	5	Gustafson et al., 2012
						392461	39246	5	Gustafson et al., 2012
				3924610	392461	5	Gustafson et al., 2012		
				39246100	3924610	4	Sun et al., 2010a		
			LOEC	500000	500000	21	Lalone et al., 2012		
				13736135	1373614	4	Sun et al., 2010a		
				15698440	1569844	5	To et al., 2007		
				19623050	1962305	5	Liu et al., 2003		
				100000000	10000000	3	Hillegass et al., 2007		
				100000000	10000000	3	Hillegass et al., 2008		
			<i>Pimephales promelas</i>	NOEC	254000	254000	28 dph	Overturf et al., 2012	
	1160000	1160000		28 dph	Overturf et al., 2012				
LOEC	100	100		21	Lalone et al., 2012				
	50000	50000		21	Lalone et al., 2012				
	500000	500000		21	Lalone et al., 2012				
	500000	500000		29	Lalone et al., 2012				
50-24-8	Prednisolone	Invertebrates	<i>Brachionus calyciflorus</i>	LC50	254000	254000	28 dph	Overturf et al., 2012	
				LC50	48220000	4822000	1	DellaGreca et al., 2004	
		Algae	<i>Selenastrum capricornutum</i>	NOEC	160000000	3200000	3	DellaGreca et al., 2004	
				EC50	230000	46000	7	DellaGreca et al., 2004	
		Crustacea	<i>Daphnia magna</i>	NOEC	85000000	1700000	1	DellaGreca et al., 2004	
				LOEC	1000	200	21	Kugathas, 2011	
		Fish	<i>Pimephales promelas</i>	LOEC	1000	200	21	Kugathas, 2011	
				LC50	22290000	445800	1	DellaGreca et al., 2004	
		Invertebrates	<i>Brachionus calyciflorus</i>	LC50	139000	27800		Escher et al., 2011	
			Aquatic community	PNEC					

E: NRF2 CALUX

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference				
50-28-2	17β-estradiol	Algae	<i>Melosira varians</i>	NOEC	20.00	126	10	Julius et al., 2007			
					400	25.24	10	Julius et al., 2007			
					800	50.48	10	Julius et al., 2007			
				LOEC	80.00	5.05	10	Julius et al., 2007			
					200	12.62	10	Julius et al., 2007			
				Amphibia	<i>Lithobates clamitans</i>	NOEC	100	6.31	506	Coady et al., 2004	
							100	6.31	506	Coady et al., 2004	
						LOEC	100	6.31	506	Coady et al., 2004	
						<i>Lithobates pipiens</i>	NOEC	100	6.31	124	Mackenzie et al., 2003
								100	6.31	124	Mackenzie et al., 2003
		LOEC	100		0.063		124	Mackenzie et al., 2003			
		<i>Lithobates sylvaticus</i>	NOEC		100	0.063	124	Mackenzie et al., 2003			
					100	0.063	124	Mackenzie et al., 2003			
			LC50		1242	78.37	14	Hogan et al., 2006			
					1517	95.73	14	Hogan et al., 2006			
			LC50	681	42.97	14	Hogan et al., 2006				
		<i>Rhinella arenarum</i>	NOEC	100	6.31	200	Brodeur et al., 2013				
			LOEC	100	6.31	200	Brodeur et al., 2013				
				100	6.31	200	Brodeur et al., 2013				
			<i>Xenopus laevis</i>	NOEC	100	6.31	78	Carr et al., 2003			
				LOEC	100	6.31	34	Cong et al., 2006			
					100	6.31	34	Cong et al., 2006			
			Crustacea	<i>Americamysis bahia</i>	LC50	890	56.16	4	Hirano et al., 2004		
						1690	10.66	2	Hirano et al., 2004		
					<i>Ceriodaphnia dubia</i>	NOEC	1000	63.10	7	Jukosky et al., 2008a	
							200	12.62	21	Brennan et al., 2006	
		NOEC				1000	63.10	21	Brennan et al., 2006		
		<i>Daphnia magna</i>		NOEC	1000	63.10	21	Brennan et al., 2006			
					400	25.24	21	Brennan et al., 2006			
				LOEC	400	25.24	21	Brennan et al., 2006			
					1500	9.78	1	Brennan et al., 2006			
				EC50	2040	12.87	2	Brennan et al., 2006			
		<i>Eurytemora affinis</i>	NOEC		2870	18.11	2	Brennan et al., 2006			
					2970	18.74	2	Hirano et al., 2004			
					3670	23.36	1	Brennan et al., 2006			
					6.00	0.38	10	Forget-Leray et al., 2005			
					18.00	1.14	10	Forget-Leray et al., 2005			
			LC50		45.00	2.84	4	Forget-Leray et al., 2005			
				<i>Neocaridina denticulata</i>	LOEC	10.00	0.63	28	Huang et al., 2006		
						10.00	0.63	28	Huang et al., 2006		
						10.00	0.63	28	Huang et al., 2006		
						10.00	0.63	28	Huang et al., 2006		
			10.00		0.63	28	Huang et al., 2006				
		Fish	<i>Cyprinus carpio</i>	NOEC	0.10	0.0063	90	Gimeno et al., 1998			
				LOEC	0.10	0.0063	90	Gimeno et al., 1998			
					1.00	0.063	90	Gimeno et al., 1998			
				<i>Danio rerio</i>	NOEC	0.013	0.0008	8	Rose et al., 2002		
						0.024	0.0015	18	Holbech et al., 2006		
						0.024	0.0015	18	Holbech et al., 2006		
						0.025	0.0002	2	Jin et al., 2009		
						0.25	0.0016	2	Jin et al., 2009		
						0.25	0.0016	18	Holbech et al., 2006		
						2.50	0.0016	3	Jin et al., 2010		
					12.50	0.0016	3	Jin et al., 2010			
			LOEC		0.021	0.0013	8	Rose et al., 2002			
					0.025	0.0002	2	Jin et al., 2009			
			<i>Gambusia affinis</i>		0.054	0.0034	18	Holbech et al., 2006			
					0.054	0.0034	18	Holbech et al., 2006			
					1.00	0.063	16	Peute et al., 1985			
					12.50	0.079	3	Jin et al., 2010			
				EC50	0.041	0.0026	8	Rose et al., 2002			
					0.055	0.0035	18	Holbech et al., 2006			
					0.17	0.011	21	Van den Belt et al., 2004			
					0.24	0.015	21	Van den Belt et al., 2004			
				NOEC	1.00	0.063	8	Huang et al., 2013			
				LOEC	250	1.58	3	Kamata et al., 2011			
			<i>Gambusia holbrooki</i>	LOEC	1.00	0.063	8	Huang et al., 2012b			
					1.00	0.063	8	Huang et al., 2013			
					500	3.15	3	Kamata et al., 2011			
				NOEC	0.020	0.0013	28	Doyle and Lim, 2005			
				LOEC	0.10	0.0063	84	Rawson et al., 2006			
			<i>Gasterosteus aculeatus</i>	LOEC	0.10	0.0063	28	Doyle and Lim, 2005			
					0.50	0.032	28	Doyle and Lim, 2005			
				0.50	0.032	84	Rawson et al., 2006				
		NOEC		0.0010	0.00006	7	Hogan et al., 2008				
				0.010	0.0006	21	Allen et al., 2008				
				0.010	0.0006	58	Hahlbeck et al., 2004a				
				0.020	0.0013	21	Allen et al., 2008				
				0.032	0.0020	21	Allen et al., 2008				
				10.00	0.63	58	Hahlbeck et al., 2004a				
		LOEC		0.010	0.0006	7	Hogan et al., 2008				
		<i>Gobiocypris rarus</i>		0.050	0.0032	21	Allen et al., 2008				
				0.070	0.0044	21	Allen et al., 2008				
				0.10	0.0063	21	Allen et al., 2008				
				1.00	0.063	58	Hahlbeck et al., 2004a				
			NOEC	0.10	0.0006	4	Ma et al., 2009				
			LOEC	0.10	0.0006	4	Ma et al., 2009				
			<i>Ictalurus punctatus</i>	NOEC	0.10	0.0063	21	Thompson et al., 2000			
				LOEC	1.00	0.063	21	Thompson et al., 2000			
				EC50	0.17	0.011	21	Thompson et al., 2000			
				NOEC	1.00	0.063	21	Thompson et al., 2000			
		LOEC		10.00	0.63	21	Thompson et al., 2000				
		<i>Morone saxatilis</i>	EC50	1.56	0.10	21	Thompson et al., 2000				
			NOEC	0.0032	0.0002	21	Thorpe et al., 2000				
				0.0048	0.0003	14	Thorpe et al., 2003				
				0.0096	0.0006	14	Thorpe et al., 2003				
				0.10	0.0006	5	Ward et al., 2006				
		<i>Oncorhynchus mykiss</i>		0.10	0.0063	21	Thorpe et al., 2000				
				0.10	0.0063	21	Tremblay and Van der Kraak, 1999				
				0.10	0.0063	21	Tremblay and Van der Kraak, 1999				
	0.25		0.016	21	Thorpe et al., 2000						
	0.25		0.016	21	Tremblay and Van der Kraak, 1999						
	0.46		0.029	14	Thorpe et al., 2003						
LOEC	0.0089		0.0006	21	Thorpe et al., 2000						
	0.014		0.0009	14	Thorpe et al., 2003						
	0.022		0.0014	14	Thorpe et al., 2003						

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference		
50-28-2	17β-estradiol	Fish	<i>Oncorhynchus mykiss</i>	LOEC	0.10	0.0063	21	Tremblay and Van der Kraak, 1999	
					0.25	0.016	21	Thorpe et al., 2000	
					0.25	0.016	21	Tremblay and Van der Kraak, 1999	
					0.25	0.016	21	Tremblay and Van der Kraak, 1999	
					0.25	0.016	21	Thorpe et al., 2000	
					0.015	0.0009	21	Thorpe et al., 2000	
					400	2.52	0.3	Piferrer and Donaldson, 1992	
					0.0010	0.000006	4	Lee et al., 2012	
					0.0010	0.000006	4	Lee et al., 2012	
					0.010	0.00006	5	Kang et al., 2005	
					0.10	0.0063	21	Thompson et al., 2000	
					0.15	0.010	14	Jukosky et al., 2008b	
					0.23	0.014	25	Kang et al., 2002	
					100	0.0063	4	Lee et al., 2012	
					2.53	0.16	14	Jukosky et al., 2008b	
					10.00	0.063	5	Kang et al., 2006b	
					0.0050	0.0003	21	Kashiwada et al., 2002	
					0.010	0.00006	4	Lee et al., 2012	
					0.010	0.00006	4	Lee et al., 2012	
					0.010	0.00006	5	Kang et al., 2005	
					0.029	0.0018	21	Kang et al., 2002	
					0.056	0.0035	21	Kang et al., 2002	
					0.056	0.0036	14	Jukosky et al., 2008b	
					0.10	0.0006	4	Lee et al., 2012	
					0.46	0.029	25	Kang et al., 2002	
					100	0.0063	4	Lee et al., 2012	
					100	0.0063	4	Lee et al., 2012	
					100	0.063	21	Thompson et al., 2000	
					2.53	0.16	14	Jukosky et al., 2008b	
					10.00	0.063	5	Kang et al., 2006b	
					0.20	0.013	21	Thompson et al., 2000	
					0.23	0.014	14	Sun et al., 2009	
					470	29.65	21	Kashiwada et al., 2002	
					460	2.90	3	Kashiwada et al., 2002	
					460	2.90	3	Tabata et al., 2001	
			2000	12.62	4	Kang et al., 2002			
			3500	22.08	3	Kashiwada et al., 2002			
			3500	22.08	3	Tabata et al., 2001			
			0.010	0.0006	14	Thorpe et al., 2007			
			0.027	0.0017	9	Cline et al., 2003			
			0.028	0.0018	17	McGee et al., 2009			
			0.030	0.0019	21	Schultz et al., 2012			
			0.030	0.0019	21	Schultz et al., 2012			
			0.030	0.0019	21	Schultz et al., 2012			
			0.030	0.0019	21	Schultz et al., 2012			
			0.030	0.0019	21	Schultz et al., 2012			
			0.030	0.0019	21	Schultz et al., 2012			
			0.10	0.0063	14	Thorpe et al., 2007			
			0.50	0.032	21	Bringolf et al., 2004			
			0.50	0.032	21	Bringolf et al., 2004			
			0.022	0.0014	14	Thorpe et al., 2007			
			0.030	0.0019	21	Schultz et al., 2012			
			0.27	0.017	9	Cline et al., 2003			
			0.50	0.032	21	Bringolf et al., 2004			
			0.50	0.032	21	Bringolf et al., 2004			
			0.50	0.032	21	Bringolf et al., 2004			
			0.025	0.0016	14	Brian et al., 2005			
			0.12	0.0076	19	Kramer et al., 1998			
			0.25	0.016	19	Kramer et al., 1998			
			1.15	0.073	19	Kramer et al., 1998			
			0.050	0.0032	120	Nielsen and Bastrup, 2006			
			100	0.063	21	Li and Wang, 2005			
			100	0.063	21	Li and Wang, 2005			
			100	0.063	21	Li and Wang, 2005			
			0.071	0.0045	243	Robinson et al., 2004			
	0.087	0.0055	243	Robinson et al., 2004					
	0.13	0.0080	243	Robinson et al., 2004					
	0.17	0.010	243	Robinson et al., 2004					
	0.015	0.0010	14	Bjerregaard et al., 2008					
	0.020	0.0013	8	Bjerregaard et al., 2008					
	0.015	0.0010	8	Bjerregaard et al., 2008					
	0.0010	0.00006	50	Lahnsteiner et al., 2006					
	0.0010	0.00006	50	Lahnsteiner et al., 2006					
	LC50	2740	17.29	Nendza and Wenzel, 2006					
	Invertebrates	<i>Brachionus calyciflorus</i>	LOEC	0.0010	0.00006	10	Huang et al., 2012a		
				100	6.31	10	Huang et al., 2012a		
	Mollusca	<i>Elliptio complanata</i>	NOEC	100	0.63	0.25	Flynn et al., 2013		
			LOEC	100	0.63	0.25	Flynn et al., 2013		
		Aquatic community	PNEC	0.0004	0.00003		Oekotoxzentrum, Centre Ecotox		
				0.0010	0.00006		Gross-Sorokin et al., 2006		
				0.0010	0.00006		Young et al., 2004		
				0.0020	0.0001		Anderson et al., 2012; Caldwell et al., 2012		
				0.0023	0.0001		Yuan et al., 2014		
10108-64-2	Cadmium chloride	Algae	<i>Chlorella protothecoides</i>	EC50	7000	88.12	14	Aliotta et al., 1983	
					14000	176.25	14	Aliotta et al., 1983	
					14000	176.25	14	Aliotta et al., 1983	
					14000	176.25	14	Aliotta et al., 1983	
					220	2.77	14	Aliotta et al., 1983	
					14000	176.25	14	Aliotta et al., 1983	
					300	3.78	4	Qian et al., 2012	
					300	3.78	4	Qian et al., 2012	
					400	5.04	4	Qian et al., 2012	
					400	5.04	4	Qian et al., 2012	
					300	3.78	4	Qian et al., 2012	
					400	5.04	4	Qian et al., 2012	
					400	5.04	4	Qian et al., 2012	
					250	3.15	4	Qian et al., 2012	
					390	4.91	4	Qian et al., 2012	
			3100	39.03	4	Irving et al., 2009			
			9.40	1.18	3	Kallqvist, 2009			
			29.00	3.65	3	Kallqvist, 2009			
			31.00	3.90	3	Kallqvist, 2009			
			43.00	5.41	3	Kallqvist, 2009			
			62.00	7.81	3	Kallqvist, 2009			
			131	16.49	3	Kallqvist, 2009			
			199	25.05	3	Kallqvist, 2009			
			14000	176.25	14	Aliotta et al., 1983			
			122400	15409.2	14	Aliotta et al., 1983			
			Amphibia	<i>Duttaphrynus melanostictus</i>	NOEC	2.00	2.52	10	Ranatunge et al., 2012



CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference									
10108-64-2	Cadmium chloride	Amphibia	<i>Duttaphrynus melanostictus</i>	NOEC	2.00	2.52	10	Ranatunge et al., 2012								
					19.00	23.92	10	Ranatunge et al., 2012								
					19.00	23.92	10	Ranatunge et al., 2012								
					182	229	10	Ranatunge et al., 2012								
					939	182	10	Ranatunge et al., 2012								
					1879	2366	10	Ranatunge et al., 2012								
					LOEC	19.00	23.92	10	Ranatunge et al., 2012							
						19.00	23.92	10	Ranatunge et al., 2012							
						182	229	10	Ranatunge et al., 2012							
						182	229	10	Ranatunge et al., 2012							
				LC50		300	37.77	4	Shuhaimi-Othman et al., 2012							
						500	62.95	4	Shuhaimi-Othman et al., 2012							
						700	88.12	4	Shuhaimi-Othman et al., 2012							
						1000	126	4	Shuhaimi-Othman et al., 2012							
						<i>Rhinella arenarum</i>	LC50	190	23.92	4	Mastrangelo et al., 2011					
								660	83.09	4	Mastrangelo et al., 2011					
					720			90.64	4	Mastrangelo et al., 2011						
					1120			141	4	Mastrangelo et al., 2011						
					210			266	4	Mastrangelo et al., 2011						
					4090			515	4	Mastrangelo et al., 2011						
				4270	538			4	Mastrangelo et al., 2011							
				7700	969			4	Mastrangelo et al., 2011							
				10830	1363			4	Mastrangelo et al., 2011							
				14400	1435			4	Mastrangelo et al., 2011							
				Crustacea				NOEC	7600	9568	7	Woods et al., 2004				
									LOEC	100	126	21	Khaili et al., 2014			
										EC50	821	103	2	Khargarot and Das, 2009		
											3220	405	2	Khargarot and Das, 2009		
											<i>Daphnia magna</i>	NOEC	0.75	0.94	4	Chadwick Ecological Consultants, 2003
													1.67	2.10	21	Chadwick Ecological Consultants, 2003
													1.97	2.48	21	Chadwick Ecological Consultants, 2003
													3.43	4.32	21	Chadwick Ecological Consultants, 2003
													3.43	4.32	21	Chadwick Ecological Consultants, 2003
													14.60	18.38	18	Chadwick Ecological Consultants, 2003
								14.60					18.38	18	Chadwick Ecological Consultants, 2003	
								20.00	2.52				1	Haap and Kohler, 2009		
								100	12.59	1			Haap and Kohler, 2009			
								400	50.36	1			Taylor et al., 2010			
								LOEC	3.43	4.32	21	Chadwick Ecological Consultants, 2003				
									3.43	4.32	21	Chadwick Ecological Consultants, 2003				
									3.43	4.32	21	Chadwick Ecological Consultants, 2003				
									6.85	8.62	21	Chadwick Ecological Consultants, 2003				
									14.60	18.38	18	Chadwick Ecological Consultants, 2003				
									14.60	18.38	18	Chadwick Ecological Consultants, 2003				
									20.00	2.52	1	Haap and Kohler, 2009				
									50.00	6.29	1	Haap and Kohler, 2009				
									300	37.77	1	Haap and Kohler, 2009				
									EC50	3.43	4.32	21	Chadwick Ecological Consultants, 2003			
								6.85		8.62	21	Chadwick Ecological Consultants, 2003				
								7.50		0.94	2	Tan and Wang, 2011				
								14.20		1.79	2	Tan and Wang, 2011				
								14.60		18.38	18	Chadwick Ecological Consultants, 2003				
								17.50		2.20	2	Tan and Wang, 2011				
								20.00		25.18	4	Chadwick Ecological Consultants, 2003				
								24.80		3.12	2	Tan and Wang, 2011				
								46.20		5.82	2	Tan and Wang, 2011				
								170		2.140	2	Tan and Wang, 2011				
								200	25.18	1	Haap and Kohler, 2009					
								250	3147	1	Haap and Kohler, 2009					
								300	37.77	1	Haap and Kohler, 2009					
								350	44.06	1	Haap and Kohler, 2009					
								400	50.36	1	Haap and Kohler, 2009					
								450	56.65	1	Haap and Kohler, 2009					
								550	69.24	1	Haap and Kohler, 2009					
								600	75.54	1	Haap and Kohler, 2009					
								714	89.84	1	Taylor et al., 2010					
								750	94.42	1	Haap and Kohler, 2009					
								<i>Gammarus pseudolimnaeus</i>	LC50	20.00	25.18	4	Call et al., 1983			
										24.00	30.21	4	Call et al., 1983			
										45.00	56.65	4	Call et al., 1983			
										95.00	120	4	Call et al., 1983			
										140	176	4	Call et al., 1983			
										<i>Gammarus pulex</i>	NOEC	3.40	4.28	10	Vellinger et al., 2013	
												3.40	4.28	10	Vellinger et al., 2013	
												6.00	7.55	10	Vellinger et al., 2013	
												6.00	7.55	10	Vellinger et al., 2013	
												30.00	3.78	2	Alonso et al., 2009	
								30.00	3.78			2	Alonso et al., 2009			
								50.00	6.29			2	Alonso et al., 2009			
								50.00	6.29			2	Alonso et al., 2009			
								50.00	6.29			2	Alonso et al., 2009			
								50.00	6.29			2	Alonso et al., 2009			
								LOEC	3.40	4.28	10	Vellinger et al., 2013				
									3.40	4.28	10	Vellinger et al., 2013				
									3.40	4.28	10	Vellinger et al., 2013				
									6.00	7.55	10	Vellinger et al., 2013				
									6.00	7.55	10	Vellinger et al., 2013				
									50.00	6.29	2	Alonso et al., 2009				
									50.00	6.29	2	Alonso et al., 2009				
									<i>Hyalella azteca</i>	NOEC	0.50	0.63	28	Chadwick Ecological Consultants, 2003		
											0.80	101	28	Chadwick Ecological Consultants, 2003		
											1.10	138	28	Mathi, 2012		
								130			164	28	Chadwick Ecological Consultants, 2003			
								4.50			5.67	28	Chadwick Ecological Consultants, 2003			
								LOEC			1.10	138	28	Chadwick Ecological Consultants, 2003		
											130	164	28	Chadwick Ecological Consultants, 2003		
											2.20	2.77	28	Chadwick Ecological Consultants, 2003		
											3.10	3.90	28	Mathi, 2012		
											4.50	5.67	28	Chadwick Ecological Consultants, 2003		
									110	138	4	Call et al., 1983				
<i>Ozotelphusa senex</i>	LC50	240	30.21						1	Reddy et al., 2011						
		13.15	16.55						4	Shuhaimi-Othman et al., 2011a						
		28.76	36.21						4	Shuhaimi-Othman et al., 2011a						
		50.73	63.87						4	Shuhaimi-Othman et al., 2011a						
		125	158					4	Shuhaimi-Othman et al., 2011a							
		Fish	<i>Cirrhinus mrigala</i>					NOEC	98.00	12.34	4	Bhilave et al., 2008				
									98.00	123	30	Bhilave et al., 2008				
									98.00	123	30	Bhilave et al., 2008				

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference	
10108-64-2	Cadmium chloride	Fish	<i>Cirrhinus mrigala</i>	LOEC	132	166	30	Bhilave et al., 2008
				LC50	132	16.62	4	Bhilave et al., 2008
			<i>Danio rerio</i>	NOEC	43.00	5.41	3	Kim et al., 2011
					43.00	5.41	3	Kim et al., 2011
					43.00	5.41	3	Kim et al., 2011
					43.00	5.41	3	Kim et al., 2011
					43.00	5.41	3	Kim et al., 2011
			<i>Gambusia affinis</i>	LOEC	43.00	5.41	3	Kim et al., 2011
				LC50	2360	297	4	Annabi et al., 2009
					3250	409	4	Annabi et al., 2009
					3840	483	4	Annabi et al., 2009
			<i>Halobatrachus didactylus</i>	LC50	14670000	1846844	1	Soares et al., 2008
				LC50	392920	49466	4	Kasherwani et al., 2009
			<i>Heteropneustes fossilis</i>		401310	50522	4	Kasherwani et al., 2009
					409880	51601	4	Kasherwani et al., 2009
			<i>Lepomis macrochirus</i>	LC50	434740	54731	4	Kasherwani et al., 2009
					26000	3273	4	U.S. EPA, 2013
			<i>Oncorhynchus kisutch</i>	NOEC	3.70	0.47	2	Williams and Gallagher, 2013
					3.70	0.47	2	Williams and Gallagher, 2013
					347	43.68	2	Williams and Gallagher, 2013
					347	43.68	2	Williams and Gallagher, 2013
				LOEC	3.70	0.47	2	Williams and Gallagher, 2013
					347	43.68	2	Williams and Gallagher, 2013
			<i>Oncorhynchus mykiss</i>	NOEC	347	43.68	2	Williams and Gallagher, 2013
					0.16	0.20	62	Mebane et al., 2008
					0.60	0.76	69	Mebane et al., 2008
					0.71	0.89	28	Sandhu et al., 2014
					0.71	0.89	28	Sandhu et al., 2014
					0.71	0.89	28	Sandhu et al., 2014
					0.71	0.89	28	Sandhu et al., 2014
					1.10	1.38	62	Mebane et al., 2008
					1.30	1.64	69	Mebane et al., 2008
					1.30	1.64	69	Mebane et al., 2008
					1.60	2.01	62	Mebane et al., 2008
					1.85	2.33	28	Sandhu et al., 2014
					1.85	2.33	28	Sandhu et al., 2014
					1.85	2.33	28	Sandhu et al., 2014
					1.85	2.33	28	Sandhu et al., 2014
					2.90	3.65	69	Mebane et al., 2008
					50.00	6.29	0.125	Baldissarotto et al., 2005
					0.16	0.20	62	Mebane et al., 2008
					0.71	0.89	28	Sandhu et al., 2014
					0.71	0.89	28	Sandhu et al., 2014
					0.71	0.89	28	Sandhu et al., 2014
					0.71	0.89	28	Sandhu et al., 2014
					0.71	0.89	28	Sandhu et al., 2014
					1.30	1.64	69	Mebane et al., 2008
					1.30	1.64	69	Mebane et al., 2008
					1.85	2.33	28	Sandhu et al., 2014
					1.85	2.33	28	Sandhu et al., 2014
					1.85	2.33	28	Sandhu et al., 2014
					2.50	3.15	62	Mebane et al., 2008
					2.90	3.65	69	Mebane et al., 2008
					6.90	8.69	69	Mebane et al., 2008
					50.00	6.29	0.125	Baldissarotto et al., 2005
					0.84	0.11	4	Mebane et al., 2008
					0.89	0.11	4	Mebane et al., 2008
					4.40	0.55	4	Call et al., 1983
					5.70	0.72	4	Call et al., 1983
					7.80	0.98	4	Call et al., 1983
					18.00	2.27	4	Call et al., 1983
					26.00	3.27	4	Call et al., 1983
			<i>Oreochromis niloticus</i>	NOEC	5.00	6.29	28	Silva and Pathiratne, 2008
					15.00	18.88	9	Silva and Pathiratne, 2008
					30.00	37.77	28	Silva and Pathiratne, 2008
				LOEC	5.00	6.29	9	Silva and Pathiratne, 2008
					5.00	6.29	28	Silva and Pathiratne, 2008
			<i>Oryzias latipes</i>	NOEC	15.00	18.88	28	Silva and Pathiratne, 2008
					1.03	1.30	49	Tilton et al., 2004
					8.05	10.13	49	Tilton et al., 2004
					8.05	10.13	49	Tilton et al., 2004
					8.05	10.13	49	Tilton et al., 2004
					8.05	10.13	50	Thompson, 2000
					1.03	1.30	49	Tilton et al., 2004
					1.03	1.30	50	Thompson, 2000
					4.63	5.83	49	Tilton et al., 2004
					8.05	10.13	50	Thompson, 2000
			<i>Pimephales promelas</i>	NOEC	32.40	40.79	21	Sellin and Kolok, 2009
					50.80	63.95	21	Sellin and Kolok, 2009
					50.80	63.95	21	Sellin and Kolok, 2009
					50.80	63.95	21	Sellin and Kolok, 2009
					86.70	109	8	Sellin and Kolok, 2009
					86.70	109	8	Sellin and Kolok, 2009
					86.70	109	8	Sellin and Kolok, 2009
					86.70	109	8	Sellin and Kolok, 2009
					86.70	109	8	Sellin and Kolok, 2009
					86.70	109	8	Sellin and Kolok, 2009
					50.80	63.95	21	Sellin and Kolok, 2009
				LOEC	60.00	75.54	4	Shuhaimi-Othman et al., 2011b
				LC50	90.00	113	4	Shuhaimi-Othman et al., 2011b
					130	164	4	Shuhaimi-Othman et al., 2011b
					190	239	4	Shuhaimi-Othman et al., 2011b
			<i>Chironomus riparius</i>	NOEC	200	25.18	1	Choi and Ha, 2009
					200	25.18	1	Choi and Ha, 2009
					200	25.18	1	Choi and Ha, 2009
					200	25.18	1	Choi and Ha, 2009
					2000	252	1	Choi and Ha, 2009
					2000	252	1	Park et al., 2012
					20000	2518	1	Choi and Ha, 2009
					20000	2518	1	Choi and Ha, 2009
	200	25.18		1	Choi and Ha, 2009			
	2000	252		1	Choi and Ha, 2009			
	2000	252		1	Choi and Ha, 2009			
	2000	252		1	Choi and Ha, 2009			
	2000	252		1	Choi and Ha, 2009			
	2000	252		1	Choi and Ha, 2009			
		10000		1259	1	Park et al., 2012		
		20000	2518	1	Choi and Ha, 2009			

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference				
10108-64-2	Cadmium chloride	Insecta	<i>Chironomus riparius</i>	LC50	212230	2678	1	Choi and Ha, 2009			
			<i>Dreissena polymorpha</i>	NOEC	34.00	42.80	5	Faria et al., 2009			
					34.00	42.80	5	Faria et al., 2009			
					34.00	42.80	5	Faria et al., 2009			
					34.00	42.80	5	Faria et al., 2009			
		Mollusca	<i>Lymnaea stagnalis</i>	LOEC	47.50	59.80	20	Desouky, 2012			
					34.00	42.80	5	Faria et al., 2009			
					34.00	42.80	5	Faria et al., 2009			
					47.50	59.80	20	Desouky, 2012			
					48.00	60.43	20	Desouky, 2012			
		Polyp	<i>Hydra vulgaris</i>	LOEC	48.00	60.43	20	Desouky, 2012			
					48.00	60.43	20	Desouky, 2012			
					64.00	80.57	4	Kar and Aditya, 2010			
					27.00	33.99	4	Shuhaimi-Orhman et al., 2012			
					74.00	93.16	4	Shuhaimi-Orhman et al., 2012			
Worm	<i>Nais elinguis</i>	LC50	74.00	93.16	4	Shuhaimi-Orhman et al., 2012					
			94.00	118	4	Shuhaimi-Orhman et al., 2012					
			158	199	4	Shuhaimi-Orhman et al., 2012					
			158	199	4	Shuhaimi-Orhman et al., 2012					
			158	199	4	Shuhaimi-Orhman et al., 2012					
133-06-2	Captan	Algae	<i>Chlorella pyrenoidosa</i>	NOEC	6020	9541	4	Anton, 1993			
				EC50	44500	70528	4	Anton, 1993			
				2400	380	3	Kikuchi, 1993				
			Amphibia	<i>Selenastrum capricornutum</i>	NOEC	3125	49.53	12	Mouchet et al., 2006		
						125	198	12	Mouchet et al., 2006		
					125	198	12	Mouchet et al., 2006			
					125	198	12	Mouchet et al., 2006			
					125	198	12	Mouchet et al., 2006			
			<i>Pleurodeles waltl</i>	LOEC	62.50	99.06	12	Mouchet et al., 2006			
					250	396	12	Mouchet et al., 2006			
					3125	49.53	12	Mouchet et al., 2006			
					62.50	99.06	12	Mouchet et al., 2006			
					15.60	24.72	12	Mouchet et al., 2006			
		Fish	<i>Xenopus laevis</i>	NOEC	62.50	99.06	12	Mouchet et al., 2006			
					62.50	99.06	12	Mouchet et al., 2006			
					62.50	99.06	12	Mouchet et al., 2006			
					62.50	99.06	12	Mouchet et al., 2006			
					62.50	99.06	12	Mouchet et al., 2006			
					<i>Carassius auratus</i>	LC50	890	141	4	Anton, 1993	
							EC50	358	56.82	5	Padilla et al., 2012
							LC50	77.50	12.28	4	Johnson and Finley, 1980
							LC50	141	22.35	4	Johnson and Finley, 1980
							LC50	56.40	8.94	4	Johnson and Finley, 1980
							LC50	198	21.87	4	Johnson and Finley, 1980
							LC50	73.20	11.60	4	Johnson and Finley, 1980
							LC50	56.50	8.95	4	Johnson and Finley, 1980
							LC50	500	79.24	2	Tsuji et al., 1986
							LC50	610	96.68	2	Tsuji et al., 1986
			<i>Danio rerio</i>	LC50	800	127	2	Tsuji et al., 1986			
					LC50	120	19.02	4	Johnson and Finley, 1980		
					LC50	200	31.70	4	Johnson and Finley, 1980		
					LC50	80.00	12.68	4	Johnson and Finley, 1980		
					LC50	200	31.70	4	Johnson and Finley, 1980		
		10605-21-7	Carbendazim	Algae	<i>Salvelinus namaycush</i>	LC50	49.00	7.77	4	Johnson and Finley, 1980	
					<i>Achnanthes</i> sp.	NOEC	330	131	28	Van den Brink et al., 2000	
	NOEC					1000	398	28	Van den Brink et al., 2000		
	NOEC					330	131	28	Van den Brink et al., 2000		
	NOEC					100	39.81	28	Van den Brink et al., 2000		
	EC50			340		13.54	2	Canton, 1976			
	<i>Chlorella pyrenoidosa</i>			EC50	34650	13794	4	Ma et al., 2002a			
					NOEC	100	39.81	28	Van den Brink et al., 2000		
					NOEC	100	39.81	28	Van den Brink et al., 2000		
					NOEC	1000	398	28	Van den Brink et al., 2000		
					NOEC	1000	398	28	Van den Brink et al., 2000		
					NOEC	1000	398	28	Van den Brink et al., 2000		
					NOEC	1000	398	28	Van den Brink et al., 2000		
					NOEC	1000	398	28	Van den Brink et al., 2000		
					NOEC	1000	398	28	Van den Brink et al., 2000		
			EC50	19050	7584	4	Ma et al., 2002a				
Amphibia	<i>Stephanodiscus</i> sp.	NOEC	100	39.81	28	Van den Brink et al., 2000					
			NOEC	191	7.61	4	Yoon et al., 2008				
			LOEC	382	15.22	4	Yoon et al., 2008				
			LOEC	574	22.83	4	Yoon et al., 2008				
			LOEC	1072	42.67	4	Yoon et al., 2008				
Crustacea	<i>Xenopus laevis</i>	<i>Acroporus herpae</i>	NOEC	3.30	1.31	28	Van den Brink et al., 2000				
				NOEC	33.00	13.14	21	Daam and Van den Brink, 2007			
				NOEC	33.00	13.14	28	Van den Brink et al., 2000			
				NOEC	100	39.81	28	Van den Brink et al., 2000			
				NOEC	10.00	3.98	28	Canton, 1976			
			<i>Alonella exigua</i>	NOEC	33.00	13.14	28	Van den Brink et al., 2000			
					60.00	2.39	1	Ferreira et al., 2008			
					100	39.81	4	Van den Brink et al., 2000			
					LOEC	70.00	2.79	1	Ferreira et al., 2008		
					EC50	3.50	0.14	1	Ferreira et al., 2008		
					20.00	7.96	1	Canton, 1976			
					22.90	0.91	1	Ferreira et al., 2008			
					24.40	0.97	1	Ferreira et al., 2008			
					28.20	1.12	2	Ferreira et al., 2008			
					28.60	1.14	1	Ferreira et al., 2008			
			<i>Cyclopoida</i>	NOEC	37.00	14.73	28	Van den Brink et al., 2000			
					54.10	2.15	2	Ferreira et al., 2008			
					68.70	2.73	2	Ferreira et al., 2008			
					73.10	2.91	2	Ferreira et al., 2008			
					97.54	3.88	1	Ferreira et al., 2008			
					103	4.10	2	Ferreira et al., 2008			
					113	44.99	4	Van den Brink et al., 2000			
					137	5.45	1	Ferreira et al., 2008			
					145	5.78	2	Ferreira et al., 2008			
					157	6.24	2	Ferreira et al., 2008			
Fish	<i>Daphnia magna</i>	<i>Graptoleberis testudinaria</i>	NOEC	460	18.31	2	Canton, 1976				
				NOEC	33.00	13.14	21	Daam and Van den Brink, 2007			
				LC50	16767	6675	4	Rico et al., 2011			
				NOEC	33.00	13.14	21	Daam and Van den Brink, 2007			
				NOEC	33.00	13.14	28	Van den Brink et al., 2000			
			<i>Colossoma macropomum</i>	LC50	4162	166	4	Rico et al., 2011			
					LC50	3690	147	4	Rico et al., 2011		
					LC50	4138	165	4	Rico et al., 2011		
					LC50	1800	71.66	2	Canton, 1976		
					LC50	4238	169	4	Rico et al., 2011		
			<i>Nannostomus unifasciatus</i>	LC50	1648	65.61	4	Rico et al., 2011			
					LC50	2000	79.6	14	Dijksterhuis et al., 2011		
					NOEC	260	10.4	14	Dijksterhuis et al., 2011		
					NOEC	1700	67.68	3	Domingues et al., 2009		
					NOEC	1700	67.68	3	Domingues et al., 2009		
Fungi	<i>Paracheirodon axelrodi</i>	<i>Fusarium sporotrichioides</i>	NOEC	1700	67.68	3	Domingues et al., 2009				
				1700	67.68	3	Domingues et al., 2009				
				1700	67.68	3	Domingues et al., 2009				
				1700	67.68	3	Domingues et al., 2009				
				1700	67.68	3	Domingues et al., 2009				
			<i>Trichoderma hamatum</i>	NOEC	1700	67.68	3	Domingues et al., 2009			
					1700	67.68	3	Domingues et al., 2009			
					1700	67.68	3	Domingues et al., 2009			
					1700	67.68	3	Domingues et al., 2009			
					1700	67.68	3	Domingues et al., 2009			
Insecta	<i>Kiefferulus calligaster</i>		NOEC	5000	199	3	Domingues et al., 2009				
				5000	199	3	Domingues et al., 2009				
				5000	199	3	Domingues et al., 2009				
				5000	199	3	Domingues et al., 2009				
				5000	199	3	Domingues et al., 2009				

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference			
10605-21-7	Carbendazim	Insecta	<i>Kiefferulus calligaster</i>	LOEC	5000	1991	Domingues et al., 2009			
			<i>Colurella uncinata</i>	NOEC	100	39.81	21	Daam and Van den Brink, 2007		
		Invertebrates	<i>Epiphanes brachionus</i>	NOEC	33.00	13.14	21	Daam and Van den Brink, 2007		
			<i>Keratella quadrata</i>	NOEC	330	131	28	Van den Brink et al., 2000		
			<i>Lecane sp.</i>	NOEC	330	131	28	Van den Brink et al., 2000		
			<i>Lepadella patella</i>	NOEC	33.00	13.14	21	Daam and Van den Brink, 2007		
			<i>Testudinella parva</i>	NOEC	330	131	28	Van den Brink et al., 2000		
			<i>Trichocerca sp.</i>	NOEC	330	131	28	Van den Brink et al., 2000		
			Mollusca	<i>Pomacea doliooides</i>	LC50	1758576	700102	4	Rico et al., 2011	
				<i>Buenaia unguis</i>	LC50	73822	2939	3	Rico et al., 2011	
			Plants	<i>Elodea canadensis</i>	NOEC	10000	3981	21	Belgers et al., 2009	
					EC50	9743	3879	21	Belgers et al., 2009	
				<i>Elodea nuttallii</i>	NOEC	10000	3981	21	Belgers et al., 2009	
				<i>Hydrophilus sp.</i>	LC50	80669	3211	4	Rico et al., 2011	
				<i>Myriophyllum spicatum</i>	NOEC	10000	3981	21	Belgers et al., 2009	
				<i>Palustris laboulbenii</i>	LC50	111329	4432	4	Rico et al., 2011	
				<i>Potamogeton crispus</i>	NOEC	10000	3981	21	Belgers et al., 2009	
				Aquatic community	PNEC	0.34	0.14		Oekotoxzentrum, Centre Ecotox	
						0.57	0.23		Oekotoxzentrum, Centre Ecotox	
						0.031	1.25		Huang et al., 1996	
				683-18-1	Dibutyltin dichloride	Algae	<i>Scenedesmus acutus</i>	EC50	900	3583
		<i>Daphnia magna</i>	EC50				900	3583	1	Vighi and Calamari, 1985
		Crustacea	<i>Oncorhynchus mykiss</i>			NOEC	48.61	1935	110	De Vries et al., 1991
	NOEC		48.61			1935	110	De Vries et al., 1991		
	LOEC		243			9677	110	De Vries et al., 1991		
	LOEC		243			9677	110	De Vries et al., 1991		
<i>Oryzias latipes</i>	NOEC		1800			7659	30	Wester and Canton, 1991		
	LOEC		320			12739	30	Wester and Canton, 1991		
<i>Poecilia reticulata</i>	NOEC		1800			7659	30	Wester and Canton, 1991		
	LOEC		320			12739	30	Wester and Canton, 1991		
Mollusca	<i>Anodonta anatina</i>		LOEC			37.98	152	210	Herwig and Holwerda, 1986	
	<i>Xenopus laevis</i>		LOEC			2.68	0.27	3	Nishimura et al., 1997	
56-53-1	Diethylstilbestrol		Amphibia				LOEC	2.68	0.27	3
				LOEC	2.68	0.27	3	Nishimura et al., 1997		
		Crustacea	<i>Daphnia magna</i>	NOEC	100	100	6	Kashian and Dodson, 2004		
				NOEC	100	100	6	Kashian and Dodson, 2004		
				NOEC	100	100	21	Brennan et al., 2006		
				NOEC	200	20.00	2	Zou and Fingerman, 1997		
				NOEC	500	500	21	Baldwin et al., 1995		
				NOEC	500	500	21	Baldwin et al., 1995		
				NOEC	500	500	21	Baldwin et al., 1995		
				NOEC	500	500	21	Baldwin et al., 1995		
				NOEC	500	500	21	Baldwin et al., 1995		
				NOEC	500	500	21	Baldwin et al., 1995		
				NOEC	540	54.00	2	Baldwin et al., 1995		
			NOEC	900	90.00	2	Baldwin et al., 1995			
			LOEC	100	100	6	Kashian and Dodson, 2004			
			NOEC	200	200	21	Brennan et al., 2006			
			NOEC	500	50.00	2	Baldwin et al., 1995			
			NOEC	500	500	21	Baldwin et al., 1995			
			NOEC	500	500	21	Baldwin et al., 1995			
			NOEC	500	500	21	Baldwin et al., 1995			
			NOEC	500	500	21	Baldwin et al., 1995			
			NOEC	500	500	21	Baldwin et al., 1995			
			NOEC	500	500	21	Baldwin et al., 1995			
	NOEC	500	500	21	Baldwin et al., 1995					
	NOEC	1090	109	2	Zou and Fingerman, 1997					
	NOEC	1200	120	2	Baldwin et al., 1995					
	NOEC	1550	155	2	Baldwin et al., 1995					
	NOEC	1870	187	2	Baldwin et al., 1995					
	NOEC	2030	203	1	Baldwin et al., 1995					
	NOEC	3740	371	1	Baldwin et al., 1995					
	NOEC	30.00	30.00	18	Breitholtz and Bengtsson, 2001					
	NOEC	30.00	30.00	18	Breitholtz and Bengtsson, 2001					
	LOEC	30.00	30.00	18	Breitholtz and Bengtsson, 2001					
	LC50	290	290	4	Breitholtz and Bengtsson, 2001					
Fish	<i>Pimephales promelas</i>	NOEC	3.20	3.20	21	Panter et al., 2002				
		LOEC	3.20	3.20	21	Panter et al., 2002				
Worm	<i>Dugesia japonica</i>	LC50	500	500	4	Li, 2013				
		LC50	600	60.00	3	Li, 2013				
		LC50	700	70.00	2	Li, 2013				
		LC50	800	80.00	1	Li, 2013				
12175-5	Malathion	Algae		EC50	60000	3786	Nendza and Wenzel, 2006			
				EC50	3000	189	Nendza and Wenzel, 2006			
		Crustacea	<i>Daphnia magna</i>	EC50	100	0.063	Nendza and Wenzel, 2006			
				LC50	200	12.62	Nendza and Wenzel, 2006			
7487-94-7	Mercuric chloride	Algae	<i>Chlorella protothecoides</i>	LC50	390	778	14	Aliotta et al., 1983		
				LC50	1960	3911	14	Aliotta et al., 1983		
			LC50	260	519	14	Aliotta et al., 1983			
			LC50	3000	5986	14	Aliotta et al., 1983			
			LC50	100	200	14	Aliotta et al., 1983			
			LC50	780	1556	14	Aliotta et al., 1983			
			LC50	200	399	14	Aliotta et al., 1983			
			LC50	600	197	14	Aliotta et al., 1983			
		Crustacea	<i>Barytelphusa cucularis</i>	LC50	450	898	4	Chourpagar and Kulkarni, 2011		
				LC50	630	1257	4	Chourpagar and Kulkarni, 2011		
				LC50	860	1746	4	Chourpagar and Kulkarni, 2011		
				LC50	1040	2075	4	Chourpagar and Kulkarni, 2011		
			<i>Ceriodaphnia dubia</i>	EC50	7.00	1.40	2	Valenti et al., 2006		
				EC50	1100	2.19	2	Valenti et al., 2006		
			<i>Cypris subglobosa</i>	EC50	97.00	19.35	2	Khargarot and Das, 2009		
				EC50	369	73.63	2	Khargarot and Das, 2009		
			<i>Daphnia magna</i>	NOEC	8.00	1.60	2	Valenti et al., 2006		
				EC50	19.00	3.79	2	Valenti et al., 2006		
				NOEC	90.00	17.96	2	Valenti et al., 2006		
		Macrobrachium rosenbergii		NOEC	10.00	19.95	4	Kaoud et al., 2011		
				NOEC	50.00	99.76	4	Kaoud et al., 2011		
				NOEC	50.00	99.76	4	Kaoud et al., 2011		
				LOEC	50.00	99.76	4	Kaoud et al., 2011		
	LOEC		100	200	4	Kaoud et al., 2011				
	LOEC		100	200	4	Kaoud et al., 2011				
	LOEC		430	858	4	Kaoud et al., 2011				
<i>Ozotetelphusa senex</i>	LOEC		70.00	13.97	1	Reddy et al., 2011				
<i>Danio rerio</i>	LC50		20.00	3.99	2	Lahnsteiner, 2008				
<i>Oreochromis niloticus</i>	NOEC		80.00	160	14	Carvalho et al., 2009				
	NOEC		80.00	160	14	Carvalho et al., 2009				
	LOEC	80.00	160	14	Carvalho et al., 2009					
Fish	<i>Tinca tinca</i>	NOEC	100	19.95	1	Shah and Altindag, 2004				
		NOEC	100	19.95	4	Shah and Altindag, 2004				
		NOEC	100	19.95	4	Shah and Altindag, 2004				
		NOEC	100	200	21	Shah and Altindag, 2004				
		NOEC	250	49.88	1	Shah and Altindag, 2004				
		NOEC	250	49.88	1	Shah and Altindag, 2004				
		NOEC	250	49.88	1	Shah and Altindag, 2004				

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TECs (µg/L)	Exposure time (days)	Reference	
7487-94-7	Mercuric chloride	Fish	<i>Tinca tinca</i>	NOEC	250	49.88	1	Shah and Altindag, 2004
					250	49.88	4	Shah and Altindag, 2004
					250	499	21	Shah and Altindag, 2004
					1000	200	2	Shah and Altindag, 2004
				LOEC	100	19.95	1	Shah and Altindag, 2004
					100	19.95	4	Shah and Altindag, 2004
					100	200	21	Shah and Altindag, 2004
					100	200	21	Shah and Altindag, 2004
					250	49.88	4	Shah and Altindag, 2004
					250	49.88	4	Shah and Altindag, 2004
					250	499	21	Shah and Altindag, 2004
					250	499	21	Shah and Altindag, 2004
					1000	200	2	Shah and Altindag, 2004
					1000	200	2	Shah and Altindag, 2004
					1000	200	2	Shah and Altindag, 2004
		Insecta	<i>Chimarra sp.</i>	NOEC	100	2.00	4	Xie et al., 2009
				LOEC	100	2.00	4	Xie et al., 2009
				NOEC	100	2.00	4	Xie et al., 2009
				LOEC	100	2.00	4	Xie et al., 2009
				LOEC	100	2.00	4	Xie et al., 2009
			<i>Hydropsyche betteri</i>	NOEC	100	2.00	4	Xie et al., 2009
				LOEC	100	2.00	4	Xie et al., 2009
				NOEC	100	2.00	4	Xie et al., 2009
				LOEC	100	2.00	4	Xie et al., 2009
				LOEC	100	2.00	4	Xie et al., 2009
			<i>Isonychia sp.</i>	NOEC	100	2.00	4	Xie et al., 2009
				NOEC	100	2.00	4	Xie et al., 2009
				LOEC	100	2.00	4	Xie et al., 2009
				LOEC	100	2.00	4	Xie et al., 2009
				LOEC	100	2.00	4	Xie et al., 2009
		Mollusca	<i>Dreissena polymorpha</i>	NOEC	40.00	79.81	5	Faria et al., 2009
					40.00	79.81	5	Faria et al., 2009
					40.00	79.81	5	Faria et al., 2009
					40.00	79.81	5	Faria et al., 2009
					40.00	79.81	5	Faria et al., 2009
LOEC	40.00		79.81	5	Faria et al., 2009			
<i>Epioblasma brevidens</i>	EC50		0.73	0.15	2	Valenti et al., 2006		
	LC50		27.00	5.39	2	Valenti et al., 2006		
			47.00	9.38	2	Valenti et al., 2006		
			25.00	4.99	2	Valenti et al., 2006		
			27.00	5.39	2	Valenti et al., 2006		
<i>Epioblasma capsaeformis</i>			36.00	7.18	2	Valenti et al., 2006		
			54.00	10.77	2	Valenti et al., 2006		
			40.00	7.98	1	Valenti et al., 2006		
			40.00	7.98	2	Valenti et al., 2006		
		104	20.74	2	Faria et al., 2010			
Crustacea	<i>Unio elongatulus</i>	EC50	4.00	10.05	2	Valenti et al., 2006		
			8.00	20.10	2	Valenti et al., 2006		
			25.00	62.80	2	Valenti et al., 2006		
			30.00	75.36	2	Valenti et al., 2006		
			15.00	37.68	2	Valenti et al., 2006		
	<i>Ceriodaphnia dubia</i>	NOEC	15.00	37.68	2	Valenti et al., 2006		
		EC50	15.00	37.68	2	Valenti et al., 2006		
			8.00	45.21	2	Valenti et al., 2006		
			20.00	50.24	2	Valenti et al., 2006		
			60.00	151	2	Valenti et al., 2006		
	Fish	<i>Danio rerio</i>	LC50	11.50	28.89	4	OECD, 2012	
				13.40	33.66	4	OECD, 2012	
				14.40	36.17	4	OECD, 2012	
				20.10	50.49	2	OECD, 2012	
				21.70	54.51	2	OECD, 2012	
			22.20	55.76	2	OECD, 2012		
			29.20	73.35	4	OECD, 2012		
			30.30	76.11	4	OECD, 2012		
			30.30	76.11	4	OECD, 2012		
			39.90	100	4	OECD, 2012		
			40.60	102	4	OECD, 2012		
			42.70	107	4	OECD, 2012		
			43.10	108	2	OECD, 2012		
			43.70	110	2	OECD, 2012		
			46.20	116	2	OECD, 2012		
		60.60	152	2	OECD, 2012			
		124	311	2	Bachmann, 2002			
<i>Pimephales promelas</i>		LC50	39.00	97.96	4	Devlin, 2006		
			67.00	168	2	Valenti et al., 2006		
			120	301	2	Valenti et al., 2006		
			40.00	1005	5	Faria et al., 2009		
			40.00	1005	5	Faria et al., 2009		
Mollusca		<i>Dreissena polymorpha</i>	NOEC	40.00	1005	5	Faria et al., 2009	
				40.00	1005	5	Faria et al., 2009	
				40.00	1005	5	Faria et al., 2009	
				40.00	1005	5	Faria et al., 2009	
				40.00	1005	5	Faria et al., 2009	
		LOEC	40.00	1005	5	Faria et al., 2009		
		<i>Epioblasma brevidens</i>	EC50	14.2	3.57	2	Faria et al., 2010	
			LC50	8.00	20.10	2	Valenti et al., 2006	
			25.00	62.80	2	Valenti et al., 2006		
			8.00	20.10	2	Valenti et al., 2006		
			8.00	20.10	2	Valenti et al., 2006		
	<i>Epioblasma capsaeformis</i>	LC50	8.00	20.10	2	Valenti et al., 2006		
			21.00	52.75	2	Valenti et al., 2006		
			26.00	65.31	2	Valenti et al., 2006		
			3137	78.80	2	Faria et al., 2010		
		43.00	108	2	Valenti et al., 2006			
Amphibia	<i>Xenopus laevis</i>	NOEC	10740	214	1	Pickford and Morris, 1999		
			107	2.14	4	Hillegass et al., 2008		
			108	2.15	4	Hillegass et al., 2008		
Fish	<i>Danio rerio</i>	NOEC	16.60	0.17	3	Delebebeck et al., 2009		
			18.00	0.18	3	Delebebeck et al., 2009		
			2120	0.21	3	Delebebeck et al., 2009		
7718-54-9	Nickel (II) chloride	Algae	<i>Selenastrum capricornutum</i>	NOEC	2150	0.22	3	Delebebeck et al., 2009
					26.20	0.26	3	Delebebeck et al., 2009
					26.40	0.26	3	Delebebeck et al., 2009
					26.90	0.27	3	Delebebeck et al., 2009
					29.40	0.29	3	Delebebeck et al., 2009
					3150	0.32	3	Delebebeck et al., 2009
					3190	0.32	3	Delebebeck et al., 2009
					33.40	0.33	3	Delebebeck et al., 2009
					34.50	0.35	3	Delebebeck et al., 2009
					38.10	0.38	3	Delebebeck et al., 2009
					4190	0.42	3	Delebebeck et al., 2009
					4730	0.47	3	Delebebeck et al., 2009
					4790	0.48	3	Delebebeck et al., 2009
					48.60	0.49	3	Delebebeck et al., 2009
					52.10	0.52	3	Delebebeck et al., 2009
					52.30	0.52	3	Delebebeck et al., 2009
					54.00	0.54	3	Delebebeck et al., 2009
					55.70	0.56	3	Delebebeck et al., 2009
					65.00	0.65	3	Delebebeck et al., 2009
					65.00	0.65	3	Delebebeck et al., 2009
					67.10	0.67	3	Delebebeck et al., 2009
					68.40	0.68	3	Delebebeck et al., 2009

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference		
7718-54-9	Nickel (II) chloride	Algae	<i>Selenastrum capricornutum</i>	NOEC	73.00	0.73	3	Delebeeck et al., 2009	
					78.40	0.78	3	Delebeeck et al., 2009	
					83.00	0.83	3	Delebeeck et al., 2009	
					88.90	0.89	3	Delebeeck et al., 2009	
					89.40	0.89	3	Delebeeck et al., 2009	
					93.80	0.94	3	Delebeeck et al., 2009	
					97.70	0.98	3	Delebeeck et al., 2009	
					98.60	0.99	3	Delebeeck et al., 2009	
					104	1.04	3	Delebeeck et al., 2009	
					132	1.32	3	Delebeeck et al., 2009	
					145	1.45	3	Delebeeck et al., 2009	
					154	1.54	3	Delebeeck et al., 2009	
					181	1.81	3	Delebeeck et al., 2009	
					187	1.87	3	Delebeeck et al., 2009	
					278	2.78	3	Delebeeck et al., 2009	
					285	2.85	3	Delebeeck et al., 2009	
					292	2.92	3	Delebeeck et al., 2009	
					327	3.27	3	Delebeeck et al., 2009	
					LOEC	16.60	0.17	3	Delebeeck et al., 2009
						31.50	0.32	3	Delebeeck et al., 2009
						32.30	0.32	3	Delebeeck et al., 2009
						36.70	0.37	3	Delebeeck et al., 2009
						38.10	0.38	3	Delebeeck et al., 2009
						42.50	0.43	3	Delebeeck et al., 2009
						43.00	0.43	3	Delebeeck et al., 2009
						45.50	0.46	3	Delebeeck et al., 2009
						48.40	0.48	3	Delebeeck et al., 2009
						49.30	0.49	3	Delebeeck et al., 2009
						51.00	0.51	3	Delebeeck et al., 2009
						52.10	0.52	3	Delebeeck et al., 2009
						52.30	0.52	3	Delebeeck et al., 2009
						54.10	0.54	3	Delebeeck et al., 2009
						55.70	0.56	3	Delebeeck et al., 2009
						59.60	0.60	3	Delebeeck et al., 2009
						65.00	0.65	3	Delebeeck et al., 2009
						84.90	0.85	3	Delebeeck et al., 2009
						89.40	0.89	3	Delebeeck et al., 2009
						93.80	0.94	3	Delebeeck et al., 2009
					EC50	97.20	0.97	3	Delebeeck et al., 2009
						100	1.00	3	Delebeeck et al., 2009
						101	1.01	3	Delebeeck et al., 2009
						102	1.02	3	Delebeeck et al., 2009
						103	1.03	3	Delebeeck et al., 2009
						104	1.04	3	Delebeeck et al., 2009
						116	1.16	3	Delebeeck et al., 2009
						119	1.19	3	Delebeeck et al., 2009
						129	1.29	3	Delebeeck et al., 2009
						155	1.55	3	Delebeeck et al., 2009
						163	1.63	3	Delebeeck et al., 2009
						168	1.68	3	Delebeeck et al., 2009
						176	1.76	3	Delebeeck et al., 2009
						186	1.86	3	Delebeeck et al., 2009
						217	2.17	3	Delebeeck et al., 2009
						278	2.78	3	Delebeeck et al., 2009
						280	2.80	3	Delebeeck et al., 2009
						320	3.20	3	Delebeeck et al., 2009
						326	3.26	3	Delebeeck et al., 2009
						390	3.90	3	Delebeeck et al., 2009
					537	5.37	3	Delebeeck et al., 2009	
					570	5.70	3	Delebeeck et al., 2009	
					953	9.53	3	Delebeeck et al., 2009	
					EC50	81.50	0.81	3	Delebeeck et al., 2009
						83.10	0.83	3	Delebeeck et al., 2009
						91.80	0.92	3	Delebeeck et al., 2009
						93.70	0.94	3	Delebeeck et al., 2009
						98.30	0.98	3	Delebeeck et al., 2009
						108	1.08	3	Delebeeck et al., 2009
						114	1.14	3	Delebeeck et al., 2009
						122	1.22	3	Delebeeck et al., 2009
						124	1.24	3	Delebeeck et al., 2009
						125	1.25	3	Delebeeck et al., 2009
						136	1.36	3	Delebeeck et al., 2009
						141	1.41	3	Delebeeck et al., 2009
						144	1.44	3	Delebeeck et al., 2009
						145	1.45	3	Delebeeck et al., 2009
						172	1.72	3	Delebeeck et al., 2009
						255	2.55	3	Delebeeck et al., 2009
						321	3.21	3	Delebeeck et al., 2009
						339	3.39	3	Delebeeck et al., 2009
						345	3.45	3	Delebeeck et al., 2009
						362	3.62	3	Delebeeck et al., 2009
					395	3.95	3	Delebeeck et al., 2009	
					399	3.99	3	Delebeeck et al., 2009	
					400	4.00	3	Delebeeck et al., 2009	
					483	4.83	3	Delebeeck et al., 2009	
					506	5.06	3	Delebeeck et al., 2009	
					508	5.08	3	Delebeeck et al., 2009	
					584	5.84	3	Delebeeck et al., 2009	
					596	5.96	3	Delebeeck et al., 2009	
					601	6.01	3	Delebeeck et al., 2009	
					669	6.69	3	Delebeeck et al., 2009	
					742	7.42	3	Delebeeck et al., 2009	
					750	7.50	3	Delebeeck et al., 2009	
					812	8.12	3	Delebeeck et al., 2009	
					821	8.21	3	Delebeeck et al., 2009	
					880	8.80	3	Delebeeck et al., 2009	
					883	8.83	3	Delebeeck et al., 2009	
					914	9.14	3	Delebeeck et al., 2009	
					1040	10.40	3	Delebeeck et al., 2009	
					1120	11.20	3	Delebeeck et al., 2009	
1190	11.90	3	Delebeeck et al., 2009						
1240	12.40	3	Delebeeck et al., 2009						
1630	16.30	3	Delebeeck et al., 2009						
Amphibia	<i>Rhinella arenarum</i>		NOEC	4000	40.00	1	Szrum et al., 2011		
				10000	100	1	Szrum et al., 2011		
				LOEC	10000	100	1	Szrum et al., 2011	
				LC50	50.00	5.00	14	Szrum et al., 2011	
				150	15.00	14	Szrum et al., 2011		
				250	25.00	14	Szrum et al., 2011		

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference				
7718-54-9	Nickel (II) chloride	Amphibia	<i>Rhinella arenarum</i>	LC50	350	35.00	14	Sztrum et al., 2011			
					1000	100	14	Sztrum et al., 2011			
					2000	200	14	Sztrum et al., 2011			
					3000	300	14	Sztrum et al., 2011			
					4000	400	14	Sztrum et al., 2011			
					6000	600	14	Sztrum et al., 2011			
					8000	800	14	Sztrum et al., 2011			
					17000	1700	14	Sztrum et al., 2011			
					20000	2000	14	Sztrum et al., 2011			
				Crustacea	<i>Acartia tonsa</i>	LC50	747	7.47	3	Lussier and Cardin, 1985	
						<i>Asellus intermedius</i>	LC50	75000	7500	4	Ewell et al., 1986
								100000	10000	4	Ewell et al., 1986
		<i>Ceriodaphnia dubia</i>	NOEC		2.25	0.23	8	Puttaswamy and Liber, 2012			
					2.25	0.23	8	Puttaswamy and Liber, 2012			
					3.00	0.30	8	Puttaswamy and Liber, 2012			
					3.40	0.34	7	Keithly et al., 2004			
					3.80	0.38	7	Keithly et al., 2004			
					3.80	0.38	7	Keithly et al., 2004			
					5.30	0.53	7	Keithly et al., 2004			
					5.30	0.53	7	Keithly et al., 2004			
					5.80	0.58	7	Keithly et al., 2004			
					8.00	0.80	8	Puttaswamy and Liber, 2012			
					9.60	0.96	7	Keithly et al., 2004			
					15.30	1.53	7	Keithly et al., 2004			
			LOEC			3.80	0.38	7	Keithly et al., 2004		
						3.80	0.38	7	Keithly et al., 2004		
						5.30	0.53	7	Keithly et al., 2004		
				8.00	0.80	8	Puttaswamy and Liber, 2012				
				9.60	0.96	7	Keithly et al., 2004				
				9.90	0.99	7	Keithly et al., 2004				
				9.90	0.99	7	Keithly et al., 2004				
				17.30	1.73	7	Keithly et al., 2004				
				27.50	2.75	7	Keithly et al., 2004				
		EC50			5.00	0.50	8	Puttaswamy and Liber, 2012			
					5.90	0.59	8	Puttaswamy and Liber, 2012			
					5.90	0.59	8	Puttaswamy and Liber, 2012			
				81.00	0.81	2	Keithly et al., 2004				
				148	1.48	2	Keithly et al., 2004				
				261	2.61	2	Keithly et al., 2004				
		Cypris subglobosa	EC50		400	4.00	2	Keithly et al., 2004			
					1723	172	14	Keithly et al., 2004			
					75780	758	2	Khargarot and Das, 2009			
					86840	868	2	Khargarot and Das, 2009			
				<i>Daphnia magna</i>	NOEC	125	12.50	4	Vandenbrouck et al., 2009		
						125	12.50	4	Vandenbrouck et al., 2009		
					500	50.00	4	Vandenbrouck et al., 2009			
					500	50.00	4	Vandenbrouck et al., 2009			
			LOEC			2000	200	4	Vandenbrouck et al., 2009		
						125	12.50	4	Vandenbrouck et al., 2009		
			EC50		125	12.50	4	Vandenbrouck et al., 2009			
					125	12.50	4	Vandenbrouck et al., 2009			
				500	50.00	4	Vandenbrouck et al., 2009				
				500	50.00	4	Vandenbrouck et al., 2009				
				1000	100	4	Vandenbrouck et al., 2009				
				1000	100	4	Vandenbrouck et al., 2009				
				2400	24.00	1	Loureiro et al., 2010				
				3200	320	4	Ewell et al., 1986				
				8000	80.00	2	Loureiro et al., 2010				
		Eurytemora affinis		LC50	13182	1318	4	Lussier and Cardin, 1985			
				Gammarus fasciatus	LC50	100000	10000	4	Ewell et al., 1986		
	29.00				2.90	14	Keithly et al., 2004				
Hyalella azteca	NOEC		58.00	5.80	14	Keithly et al., 2004					
	LOEC		1500	150	14	Mwangi et al., 2012					
	1500		150	14	Mwangi et al., 2012						
Fish	<i>Capoeta fusca</i>	LC50	3045	305	4	Keithly et al., 2004					
		LC50	78000	780	4	Pourkhabbaz et al., 2011					
			86200	862	4	Pourkhabbaz et al., 2011					
			95800	958	4	Pourkhabbaz et al., 2011					
			121300	1213	4	Pourkhabbaz et al., 2011					
			127200	1272	4	Pourkhabbaz et al., 2011					
			139000	1399	4	Pourkhabbaz et al., 2011					
			165500	1655	4	Pourkhabbaz et al., 2011					
			215000	2150	4	Pourkhabbaz et al., 2011					
		Danio rerio	NOEC	5000	50.00	0.08	Kienle et al., 2009				
				5080	508	11	Kienle et al., 2009				
				15000	150	0.08	Kienle et al., 2009				
			15420	1542	11	Kienle et al., 2009					
	LOEC			7500	75.00	0.08	Kienle et al., 2009				
				10470	1047	11	Kienle et al., 2009				
	Ictalurus punctatus	LC50	289800	2898	2	Lahrsteiner, 2008					
		LOEC	15000	150	4	Kuykendall et al., 2009					
		Lepomis macrochirus	LOEC	15000	150	4	Kuykendall et al., 2009				
			LOEC	15000	150	4	Kuykendall et al., 2009				
		Pimephales promelas	LC50	40000	400	4	Ewell et al., 1986				
				100000	1000	4	Ewell et al., 1986				
		Insecta	<i>Chironomus riparius</i>	NOEC	5000	500	28	Langer-Jaeschich et al., 2010			
			<i>Planorbella trivolvis</i>	LC50	3200	320	4	Ewell et al., 1986			
		Mollusca	<i>Dugesia tigrina</i>	LC50	32000	3200	4	Ewell et al., 1986			
<i>Lumbriculus variegatus</i>			LC50	32000	3200	4	Ewell et al., 1986				
Worm			48000	4800	4	Ewell et al., 1986					
			830	83	3	Graff et al., 2003					
2310-17-0	Phosalone	Algae	<i>Selenastrum capricornutum</i>	EC50	930	147	3	Graff et al., 2003			
			Fish	<i>Channa orientalis</i>	LC50	8100	12.84	4	Verma et al., 1978b		
				<i>Danio rerio</i>	EC50	3443	546	5	Padilla et al., 2012		
		<i>Heteropneustes fossilis</i>	LC50	83.00	13.15	4	Verma et al., 1982				
			Algae	EC50	10000	19.95	4	Nendza and Wenzel, 2006			
				Bacteria	EC50	36000	7183	4	Nendza and Wenzel, 2006		
114-26-1	Propoxur	Crustacea	<i>Daphnia magna</i>	EC50	10	0.22	4	Nendza and Wenzel, 2006			
		Fish	<i>Daphnia magna</i>	LC50	8800	17.56	4	Nendza and Wenzel, 2006			
302-79-4	Retinoic acid	Crustacea	<i>Daphnia magna</i>	NOEC	100	15.85	>7	Peterson et al., 2001			
					800	127	4	Wang et al., 2005			
				Fish	<i>Danio rerio</i>	NOEC	0.60	0.10	2	Teixido et al., 2013	
						3.00	0.048	2	Teixido et al., 2013		
				LOEC		0.30	0.0048	2	Teixido et al., 2013		
						1.50	0.024	2	Teixido et al., 2013		
		EC50		3004351	47616	2	Elo et al., 2007				
				0.42	0.0067	6	Selderslaghs et al., 2012				
				1.48	0.023	3	Selderslaghs et al., 2012				
				3.58	0.057	2	Selderslaghs et al., 2012				



CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference	
302-79-4	Retinoic acid	Fish	<i>Danio rerio</i>	EC50	5.71	0.090	2	Teixido et al., 2013
					234	3.70	1	Selderslaghs et al., 2012
				LC50	15.32	0.24	2	Teixido et al., 2013
					40.26	0.64	6	Selderslaghs et al., 2012
					442	7.00	3	Selderslaghs et al., 2012
1948-33-0	tBHQ	Fish	<i>Ictalurus punctatus</i>	LC50	370	73.82	2	Cope et al., 1997
					150	29.93	2	Cope et al., 1997
					370	73.82	2	Cope et al., 1997
					1000	200	2	Cope et al., 1997
					18000	23544	2	Cope et al., 1997
56-36-0	Tributyltin acetate	Fish	<i>Oryzias latipes</i>	LC50	34.00	67.84	2	Tsuji et al., 1986
					38.00	75.82	1	Tsuji et al., 1986
					430	858	1	Kumar-Das et al., 1984
		Insecta	<i>Aedes aegypti</i>	LC50	19.00	37.91	1	Frick and Dejmenez, 1964
					31.00	61.85	1	Frick and Dejmenez, 1964
		Mollusca	<i>Biomphalaria glabrata</i>	LC50	41.00	81.81	1	Frick and Dejmenez, 1964
					74.00	148	0.25	Frick and Dejmenez, 1964
					74.00	148	1	Frick and Dejmenez, 1964
					85.00	170	1	Frick and Dejmenez, 1964
					88.00	176	0.25	Frick and Dejmenez, 1964
					94.00	188	0.25	Frick and Dejmenez, 1964
					190	379	0.25	Frick and Dejmenez, 1964
					200	399	0.25	Frick and Dejmenez, 1964
					200	399	1	Frick and Dejmenez, 1963
					230	459	1	Frick and Dejmenez, 1964
1461-22-9	Tributyltin chloride	Algae	<i>Scenedesmus acutus</i>	EC50	0.058	5.80	4	Huang et al., 1996
					1.20	120	4	Miana et al., 1993
					2.00	200	4	Miana et al., 1993
					4.00	400	4	Miana et al., 1993
					12.40	1240	4	Miana et al., 1993
		Crustacea	<i>Daphnia magna</i>	NOEC	1.00	100	21	Baer and Owens, 1999
					1.20	120	2	LeBlanc and McLachlan, 2000
					1.25	125	21	Oberdorster et al., 1998
					1.90	190	2	Miana et al., 1993
					5.50	550	1	Miana et al., 1993
					1.25	125	21	Oberdorster et al., 1998
					2.50	250	21	Oberdorster et al., 1998
					0.95	9.50	1	Kungolos et al., 2001
					3.40	340	5	Meador, 1986
					4.38	43.80	2	Bao et al., 1997
50471-44-8	Vindozolin	Algae	<i>Navicula pelliculosa</i>	EC50	1060	168	5	U.S. EPA, 2013
					2540	403	5	U.S. EPA, 2013
					1020	162	5	U.S. EPA, 2013
					870	138	5	U.S. EPA, 2013
					580	91.92	4	U.S. EPA, 2013
		Crustacea	<i>Americamysis bahia</i>	LC50	1800	285	4	U.S. EPA, 2013
					790	125	21	U.S. EPA, 2013
					790	125	21	U.S. EPA, 2013
					790	125	21	U.S. EPA, 2013
					1000	158.5	2	U.S. EPA, 2013
					1000	158	21	Haeba et al., 2008
					3000	47.55	1	Haeba et al., 2008
					3000	47.55	2	Haeba et al., 2008
					1000	158	21	Haeba et al., 2008
					1400	222	21	U.S. EPA, 2013
Fish	<i>Cyprindon variegatus</i>	EC50	3650	57.85	2	U.S. EPA, 2013		
			1100	17.43	4	U.S. EPA, 2013		
		NOEC	100	170	5	Padilla et al., 2012		
			10741	170	5			

CAS	Compound	Organism	Measured endpoint	Concentration (µg/L)	Final TEQs (µg/L)	Exposure time (days)	Reference	
50471-44-8	Vinclozolin	Fish	<i>Gasterosteus aculeatus</i>	LOEC	100	15.85	21	Jolly et al., 2009
			<i>Lepomis gibbosus</i>	NOEC	68 100	1079	4	U.S. EPA, 2013
				LC50	49800	789	4	U.S. EPA, 2013
			<i>Lepomis macrochirus</i>	LC50	47500	753	4	U.S. EPA, 2013
			<i>Oncorhynchus mykiss</i>	NOEC	1040	16.48	4	U.S. EPA, 2013
					1800	28.53	4	U.S. EPA, 2013
					3160	50.08	4	U.S. EPA, 2013
				LC50	2840	45.01	4	U.S. EPA, 2013
					13600	216	4	U.S. EPA, 2013
			<i>Oryzias latipes</i>	NOEC	2500	396	100	Kiparissis et al., 2003
				LOEC	2500	396	100	Kiparissis et al., 2003
					2500	396	100	Kiparissis et al., 2003
			<i>Pimephales promelas</i>	NOEC	50.00	7.92	175	U.S. EPA, 2013
					50.00	7.92	175	U.S. EPA, 2013
					50.00	7.92	175	U.S. EPA, 2013
					50.00	7.92	175	U.S. EPA, 2013
					450	71.32	21	Martinovic et al., 2008
					700	111	21	Makynen et al., 2000
					700	111	21	Makynen et al., 2000
					1200	190	34	Makynen et al., 2000
					1200	190	34	Makynen et al., 2000
					60.00	9.51	21	Martinovic et al., 2008
					60.00	9.51	21	Martinovic et al., 2008
					100	15.85	21	Villeneuve et al., 2007
					150	23.77	175	U.S. EPA, 2013
					150	23.77	175	U.S. EPA, 2013
					150	23.77	175	U.S. EPA, 2013
					150	23.77	175	U.S. EPA, 2013
					255	40.41	21	Martinovic et al., 2008
					255	40.41	21	Martinovic et al., 2008
					450	71.32	21	Martinovic et al., 2008
					450	71.32	21	Martinovic et al., 2008
					450	71.32	21	Martinovic et al., 2008
				700	111	21	Makynen et al., 2000	
				700	111	21	Makynen et al., 2000	
				700	111	21	Makynen et al., 2000	
				1200	190	34	Makynen et al., 2000	
		Invertebrates	<i>Brachionus calyciflorus</i>	NOEC	3120	49.45	1	Zavala-Aguirre et al., 2007
				LOEC	6250	99.06	1	Zavala-Aguirre et al., 2007
		Mollusca	<i>Crassostrea virginica</i>	LC50	30500	483	1	Zavala-Aguirre et al., 2007
				NOEC	2100	333	4	U.S. EPA, 2013
				EC50	3200	507	4	U.S. EPA, 2013
				LOEC	0.030	0.0048	140	Tillmann et al., 2001
		Plants	<i>Physa acuta</i>	LOEC	5000	792	21	Sanchez-Arguello et al., 2012
				NOEC	2400	380	14	U.S. EPA, 2013
				EC50	900	14.26	5	U.S. EPA, 2013
		Protozoa	<i>Acanthamoeba castellanii</i>	LOEC	10000	1585	6	Prescott et al., 1977

## F: PAH CALUX

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference			
1746-01-6	2,3,7,8 TCDD	Amphibia	<i>Pseudacris triseriata</i>	NOEC	30000	598579	2	Collier et al., 2008		
				LOEC	300	5986	2	Collier et al., 2008		
			<i>Xenopus laevis</i>	NOEC	30000	598579	3	Collier et al., 2008		
				LOEC	300	5986	2	Collier et al., 2008		
			Crustacea	<i>Daphnia magna</i>	NOEC	1000	199526		Wu et al., 2001	
					LOEC	0.10	19.95	8	Wu et al., 2001	
				Fish	<i>Esox lucius</i>	LOEC	0.10	2.00	4	Helder, 1980
						LOEC	0.10	2.00	4	Helder, 1980
					<i>Fundulus heteroclitus</i>	LC50	20000	399052	0.08	Toomey et al., 2001
						LOEC	0.0020	0.40	120	Wu et al., 2001
		<i>Gobiocypris rarus</i>			LOEC	0.0020	0.40	chronic	Wu et al., 2001	
					LOEC	1.00	19.95	2	Wu et al., 2001	
					LOEC	1.00	200	120	Wu et al., 2001	
					LOEC	1.00	200	chronic	Wu et al., 2001	
			<i>Oryzias latipes</i>	NOEC	3.40	67.84	until 3 dph	Kim and Cooper, 1998		
				LOEC	12.00	2394	10	Wisk and Cooper, 1992		
				LOEC	12.00	2394	10	Wisk and Cooper, 1992		
				EC50	1.20	23.94	3	Chen and Cooper, 1999		
				EC50	2.20	439	11dph	Wisk and Cooper, 1990b		
				EC50	2.80	55.87	4	Kim and Cooper, 1998		
		EC50		3.50	69.83	until 3 dph	Wisk and Cooper, 1990b			
		EC50		5.60	112	until 3 dph	Kim and Cooper, 1999			
		EC50		6.00	120	3	Wisk and Cooper, 1990a			
		EC50		10.10	202	until 3 dph	Kim and Cooper, 1999			
		<i>Pimephales promelas</i>	NOEC	12.50	249	until 3 dph	Kim and Cooper, 1999			
				14.00	279	until 3 dph	Wisk and Cooper, 1990b			
				14.00	279	until 3 dph	Wisk and Cooper, 1990b			
				15.00	299	3	Wisk and Cooper, 1990a			
				15.80	315	until 3 dph	Chen and Cooper, 1999			
				18.40	367	until 3 dph	Chen and Cooper, 1999			
				26.80	535	until 3 dph	Chen and Cooper, 1999			
				LC50	5.70	1137	17	Metcalfe et al., 1997		
				LC50	8.10	162	until 3 dph	Kim and Cooper, 1999		
				LC50	9.00	180	until 3 dph	Wisk and Cooper, 1990b		
		<i>Salvelinus namaycush</i>	NOEC	13.50	269	until 3 dph	Chen and Cooper, 1999			
				0.70	140	28	Adams et al., 1986			
				3.80	75.82	1	Olivieri and Cooper, 1997			
				3.80	75.82	1	Olivieri and Cooper, 1997			
				10.16	203	until 2 dph	Olivieri and Cooper, 1997			
				0.37	7.38	until 2 dph	Olivieri and Cooper, 1997			
				LC50	1.70	339	28	Adams et al., 1986		
				LC50	34.00	678	2	Walker et al., 1991		
				LC50	1.00	19.95	2	Spitsbergen et al., 1991		
				LC50	10.00	200	2	Spitsbergen et al., 1991		
		Insecta	<i>Aedes aegypti</i>	NOEC	55.00	1097	2	Walker et al., 1991		
				NOEC	55.00	1097	2	Walker et al., 1991		
				NOEC	226	4509	2	Walker et al., 1991		
LC50	65.00			1297	2	Walker et al., 1991				
NOEC	200			39905	17	Miller et al., 1973				
NOEC	200			39905	36	Miller et al., 1973				
NOEC	200			39905	55	Miller et al., 1973				
NOEC	200			39905	55	Miller et al., 1973				
NOEC	200			39905	55	Miller et al., 1973				
NOEC	200			39905	55	Miller et al., 1973				
Mollusca	<i>Physa sp.</i>	NOEC	200	39905	17	Miller et al., 1973				
		NOEC	200	39905	36	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
Worm	<i>Paranis sp.</i>	NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
		NOEC	200	39905	55	Miller et al., 1973				
Algae	<i>Anabaena flosaquae</i>	NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	1300000	130000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
Algae	<i>Chlamydomonas reinhardtii</i>	NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
Algae	<i>Ochromonas malhamensis</i>	NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
Algae	<i>Scenedesmus acutus</i>	NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	5000	500	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
		NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	4000000	400000	3	Schoeny et al., 1988				
Algae	<i>Selenastrum capricornutum</i>	NOEC	4000000	400000	3	Schoeny et al., 1988				
		EC50	15000	1500	3	Schoeny et al., 1988				
		NOEC	4000000	400000	1	Warshawskiy et al., 1995				
		EC50	4000000	400000	6	Warshawskiy et al., 1995				
		NOEC	4000000	400000	6	Nendza and Wenzel, 2006				
		EC50	5000	500	6	Reynaud et al., 2012				
		NOEC	500000	500000	16	Jaylet et al., 1986				
		LOEC	125000	125000	12	Mouchet et al., 2006				
		NOEC	500000	500000	6	Marquiset al., 2009				
		LOEC	500000	500000	6	Marquiset al., 2009				
Amphibia	<i>Xenopus laevis</i>	EC50	8700000	870000	4	Propst et al., 1997				
		EC50	9600000	960000	4	Propst et al., 1997				
		LC50	13400000	1340000	4	Propst et al., 1997				
		LC50	16700000	1670000	4	Propst et al., 1997				
		NOEC	25000	25000	14	Atienzar et al., 1999				
		Crustacea	<i>Daphnia magna</i>	NOEC	25000	25000	14	Atienzar et al., 1999		



G: PPARG CALUX

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEOs (ng/L)	Exposure time (days)	Reference				
53-70-3	dibenzo[a,h]anthracene	Crustacea	<i>Daphnia magna</i>	EC50	400	0.040	0.1	Newsted and Giesy, 1987			
					551	0.055	2	Lampi et al., 2005			
			Aquatic community	PNEC	1559	0.16	2	Lampi et al., 2005			
683-18-1	Dibutyltinchloride	Algae	<i>Scenedesmus acutus</i>	EC50	3144	0.99	4	Huang et al., 1996			
			<i>Daphnia magna</i>	EC50	900000	2846	1	Vighi and Calamari, 1985			
		Crustacea		NOEC	48614	1537	110	De Vries et al., 1991			
			<i>Oncorhynchus mykiss</i>	NOEC	48614	1537	110	De Vries et al., 1991			
		Fish		LOEC	243072	7687	110	De Vries et al., 1991			
				LOEC	243072	7687	110	De Vries et al., 1991			
			<i>Oryzias latipes</i>	NOEC	1800000	56921	30	Wester and Canton, 1991			
				LOEC	320000	10119	30	Wester and Canton, 1991			
			<i>Poecilia reticulata</i>	NOEC	1800000	56921	30	Wester and Canton, 1991			
				LOEC	320000	10119	30	Wester and Canton, 1991			
84371-65-3	Mifepristone	Mollusca	<i>Anodonta anatina</i>	LOEC	37980	1201	2.10	Herwig and Holwerda, 1986			
		Amphibia	<i>Xenopus laevis</i>	NOEC	10740000	33963	1	Rickford and Morris, 1999			
		Fish	<i>Danio rerio</i>	NOEC	107400	340	4	Hillegass et al., 2008			
302-79-4	Retinoic acid	Crustacea	<i>Daphnia magna</i>	NOEC	100000	31623	chronic	Peterson et al., 2001			
				NOEC	800000	252982	chronic	Wang et al., 2005			
		Fish	<i>Danio rerio</i>	NOEC	601	19.00	2	Teixido et al., 2013			
				LOEC	3004	95.01	2	Teixido et al., 2013			
				LOEC	300	9.50	2	Teixido et al., 2013			
				LOEC	1502	47.50	2	Teixido et al., 2013			
				EC50	3004351000	95005921	2	Elo et al., 2007			
					13.44	13.40	6	Selderslaghs et al., 2012			
					1475	46.65	3	Selderslaghs et al., 2012			
					3575	113	2	Selderslaghs et al., 2012			
					5708	181	2	Teixido et al., 2013			
				LC50	233739	7391	1	Selderslaghs et al., 2012			
					15322	485	2	Teixido et al., 2013			
					40258	1273	6	Selderslaghs et al., 2012			
					441640	13966	3	Selderslaghs et al., 2012			
					865253	27362	2	Selderslaghs et al., 2012			
					85000	26879	1	Selderslaghs et al., 2012			
			122320-73-4	Rosiglitazone	Fish	<i>Danio rerio</i>	LOEC	357428	35743	2	Elo et al., 2007
			56-36-0	Tributyltin acetate	Fish	<i>Oryzias latipes</i>	LC50	34000	10752	2	Tsuji et al., 1986
							38000	12017	1	Tsuji et al., 1986	
Insecta	<i>Aedes aegypti</i>	Mollusca		LC50	430000	135978	1	Kumar-Das et al., 1984			
				LC50	19000	6008	1	Frick and DeJimenez, 1964			
				LC50	31000	9803	1	Frick and DeJimenez, 1964			
					41000	12965	1	Frick and DeJimenez, 1964			
					74000	23401	0.25	Frick and DeJimenez, 1964			
					74000	23401	1	Frick and DeJimenez, 1964			
					85000	26879	1	Frick and DeJimenez, 1964			
					88000	27828	0.25	Frick and DeJimenez, 1964			
					94000	29725	0.25	Frick and DeJimenez, 1964			
					190000	60083	0.25	Frick and DeJimenez, 1964			
					200000	63246	0.25	Frick and DeJimenez, 1964			
					200000	63246	1	Frick and DeJimenez, 1963			
					230000	72732	1	Frick and DeJimenez, 1964			
					230000	72732	1	Frick and DeJimenez, 1963			
					270000	85381	0.25	Frick and DeJimenez, 1963			
					560000	177088	0.25	Frick and DeJimenez, 1964			
					560000	177088	0.25	Frick and DeJimenez, 1963			
			688-73-3	Tributyltin hydride	Fish	<i>Salmo salar</i>	NOEC	250	199	7	Greco et al., 2007
	LOEC	50.00				39.72	7	Greco et al., 2007			
Invertebrates	<i>Brachionus calyciflorus</i>	LC50			250	199	7	Greco et al., 2007			
	<i>Lymnaea stagnalis</i>	NOEC			19000	1509	1	Snell, 1991			
Mollusca		NOEC			19.20	15.25	21	Giusti et al., 2013			
		LOEC			94.20	74.83	21	Giusti et al., 2013			
		LOEC			94.20	74.83	21	Giusti et al., 2013			
		LOEC			200	159	180	Tilimann et al., 2001			
		EC50			53.00	18.3	4	Huang et al., 1996			
		NOEC			1200	3795	4	Miana et al., 1993			
1461-22-9	Tributyltinchloride	Algae	<i>Marisa cornuarietis</i>	EC50	2000	6325	4	Miana et al., 1993			
			<i>Scenedesmus acutus</i>	NOEC	4000	12649	4	Miana et al., 1993			
		Crustacea	<i>Daphnia magna</i>	EC50	12400	39212	4	Miana et al., 1993			
				NOEC	1000	3162	21	Baer and Owens, 1999			
					1200	379	2	LeBlanc and McLachlan, 2000			
					1250	3953	21	Oberdorster et al., 1998			
					1900	601	2	Miana et al., 1993			
					5500	1739	1	Miana et al., 1993			
				LOEC	1250	3953	21	Oberdorster et al., 1998			
					2500	7906	21	Oberdorster et al., 1998			
				EC50	950	300	1	Kungolos et al., 2001			
					3400	10752	5	Meador, 1986			
					4380	1385	2	Bao et al., 1997			
					5900	18657	4	Meador, 1986			
					9800	3099	2	Miana et al., 1993			
					12500	3953	1	Miana et al., 1993			
					10000	4111	1	Vighi and Calamari, 1985			
					13200	4174	1	Bao et al., 1997			
					15000	4743	1	De Coen et al., 1998			
			Fish	<i>Hyalella azteca</i>	LC50	1562	4941	28	Borgmann et al., 1996		
		6185		19557	7	Borgmann et al., 1996					
<i>Oncorhynchus mykiss</i>	NOEC	39.06		124	110	De Vries et al., 1991					
		195		618	110	De Vries et al., 1991					
	LOEC	1000		3162	110	Seinen et al., 1981					
		195		618	110	De Vries et al., 1991					
		195		618	110	De Vries et al., 1991					
		200		632	110	Seinen et al., 1981					
		977		3088	110	De Vries et al., 1991					
		1000		3162	110	Seinen et al., 1981					
Insecta	<i>Aedes aegypti</i>	Mollusca		NOEC	50000	15811	0.05	Baldwin et al., 1994			
				LC50	11200	3542	4	Douglas et al., 1986			
					20000	6325	4	Baldwin et al., 1994			
				NOEC	890	2814	9	Fent and Meier, 1992			
				LOEC	890	2814	8	Fent and Meier, 1992			
					4500	14230	9	Fent and Meier, 1992			
				NOEC	50.00	155	7	Lyssimachou et al., 2006			
				LC50	250	791	1	Lyssimachou et al., 2006			
				LOEC	250	791	1	Lyssimachou et al., 2006			
				LC50	390000	123329	7	Kumar-Das et al., 1984			
Mollusca	<i>Dreissena polymorpha</i>			EC50	1000	316	1	Clayton et al., 2000			
				EC50	360	114	2	Faria et al., 2010			
				LC50	1720000	5439118	4	Jagtap and Shejule, 2010			
					2650000	838004	3	Jagtap and Shejule, 2010			
					3390000	1072012	2	Jagtap and Shejule, 2010			
					4650000	1470459	1	Jagtap and Shejule, 2010			
				NOEC	500	1581	150	Janer et al., 2006			
				LOEC	30.00	94.87	150	Janer et al., 2006			
				LOEC	30.00	94.87	150	Janer et al., 2006			
					125	395	150	Janer et al., 2006			
Protozoa	<i>Unio elongatulus</i>			EC50	500	1581	150	Janer et al., 2006			
				EC50	13900	4396	2	Faria et al., 2010			
				EC50	18554	58672	5	Miyoshi et al., 2003			
					29296	92641	2	Miyoshi et al., 2003			
				EC50	74866	236748	5	Miyoshi et al., 2003			
			97652	308802	2	Miyoshi et al., 2003					

H: PXR CALUX

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEOs (ng/L)	Exposure time (days)	Reference				
1806-26-4	4-n-octylphenol	Amphibia	<i>Lithobates pipiens</i>	NOEC	206320	33054	7	Crump et al., 2002			
				LC50	206320	33054	10	Crump et al., 2002			
		Fish	<i>Oryzias latipes</i>	LOEC	577696	92550	7	Crump et al., 2002			
					2000	320	98	Knorr and Braunbeck, 2002			
15972-60-8	Alachlor	Algae	<i>Chlorella vulgaris</i>	NOEC	100000	10108	4	Garten and Frank, 1984			
				LOEC	1000000	101083	4	Garten and Frank, 1984			
			<i>Selenastrum capricornutum</i>	NOEC	350	35.38	5	U.S. EPA, 2013			
				LOEC	10000	1011	4	Garten and Frank, 1984			
			Algae	EC50	1640	166	5	U.S. EPA, 2013			
				NOEC	5350	541	28	Carder et al., 1998			
		Crustacea	<i>Daphnia magna</i>		78550	7940	28	Carder et al., 1998			
				LOEC	78550	7940	28	Carder et al., 1998			
			NOEC	5600000	56607	2	U.S. EPA, 2013				
				12000000	121300	2	U.S. EPA, 2013				
				14000000	141516	2	U.S. EPA, 2013				
				18000000	181950	2	U.S. EPA, 2013				
		Fish	<i>Cyprinodon variegatus</i>	LOEC	230000	23249	21	U.S. EPA, 2013			
					430000	43466	21	U.S. EPA, 2013			
					1700000	171841	21	U.S. EPA, 2013			
				NOEC	2600000	26282	4	U.S. EPA, 2013			
				LC50	3900000	39422	4	U.S. EPA, 2013			
				LC50	6500000	65704	4	U.S. EPA, 2013			
				<i>Lepomis macrochirus</i>	NOEC	1800000	18195	4	U.S. EPA, 2013		
						3700000	37401	4	U.S. EPA, 2013		
						4200000	42455	4	U.S. EPA, 2013		
				Crustacea	<i>Daphnia magna</i>	LC50	5600000	56607	4	U.S. EPA, 2013	
							2800000	28303	4	U.S. EPA, 2013	
							6200000	62672	4	U.S. EPA, 2013	
							6400000	64693	4	U.S. EPA, 2013	
							7600000	76823	4	U.S. EPA, 2013	
							12400000	125343	4	U.S. EPA, 2013	
						<i>Oncorhynchus mykiss</i>	NOEC	1000000	10108	4	U.S. EPA, 2013
								1800000	18195	4	U.S. EPA, 2013
								2400000	24260	4	U.S. EPA, 2013
			2400000				24260	4	U.S. EPA, 2013		
		LOEC	388000				39220	96	U.S. EPA, 2013		
			390000				39422	96	U.S. EPA, 2013		
		LC50	240000				2426	4	U.S. EPA, 2013		
			1800000				18195	4	U.S. EPA, 2013		
			3600000				36390	4	U.S. EPA, 2013		
120-12-7	Anthracene	Algae	<i>Chlorella fusca</i>	EC50	499044	3183	1	Attenburger et al., 2004			
			<i>Scenedesmus subspicatus</i>	EC50	1040000	66330	7	Djomo et al., 2004			
			<i>Selenastrum capricornutum</i>	EC50	3300	2105	1	Gala and Giesy, 1992			
		Algae		3900	24.87	0.9	Gala and Giesy, 1992				
				12100	77.17	0.9	Gala and Giesy, 1992				
				37400	239	0.9	Gala and Giesy, 1992				
			Amphibia	<i>Lithobates pipiens</i>	LC50	25000	159	0.2	Kagan et al., 1984		
					65000	415	0.02	Kagan et al., 1984			
			Crustacea	<i>Daphnia magna</i>	EC50	10000	702	1	Kagan et al., 1987		
					95000	606	2	Munoz and Tarazona, 1993.			
					211000	1346	1	Munoz and Tarazona, 1993.			
		Fish	<i>Lepomis macrochirus</i>	LC50	5600	357	10	Hatch, 1999			
					3360	2143	2	McCloskey and Oris, 1991			
					3740	23.85	4	McCloskey and Oris, 1991			
					5100	32.53	2	McCloskey and Oris, 1991			
					7470	47.64	4	McCloskey and Oris, 1991			
					8270	52.75	4	McCloskey and Oris, 1991			
					9690	61.80	2	McCloskey and Oris, 1991			
					10050	64.10	2	McCloskey and Oris, 1991			
					11920	76.02	4	Oris et al., 1984			
					6000	383	10	Hatch, 1999			
		Insecta	<i>Chironomus tentans</i>	LC50	6000	383	10	Hatch, 1999			
			<i>Utterbackia imbecillilis</i>	LC50	1930	12.31	1	Weinstein and Polk, 2001			
		Mollusca	<i>Utterbackia imbecillilis</i>		2010	12.82	0.7	Weinstein and Polk, 2001			
					2840	18.11	0.3	Weinstein and Polk, 2001			
					300000	19134	7	Mallakin et al., 1999			
		Plants	<i>Lemma gibba</i>	EC50	1300000	82913	7	Mallakin et al., 1999			
					100	6.38		EU Risk Assessment Report, 2008			
		25057-89-0	bentazon	Algae	<i>Anabaena flosaquae</i>	NOEC	3040000	3869	5	U.S. EPA, 2013	
						EC50	10130000	12891	5	U.S. EPA, 2013	
					<i>Selenastrum capricornutum</i>	NOEC	880000	1120	5	U.S. EPA, 2013	
						EC50	4500000	5727	5	U.S. EPA, 2013	
					<i>Skeletonema costatum</i>	NOEC	3720000	4734	5	U.S. EPA, 2013	
						EC50	10100000	12853	5	U.S. EPA, 2013	
				NOEC		5670	7.22	7	De la Broise and Stachowski-Haberkorn, 2012		
				Algae		5670	7.22	7	De la Broise and Stachowski-Haberkorn, 2012		
	5670				7.22	7	De la Broise and Stachowski-Haberkorn, 2012				
	28490				36.26	7	De la Broise and Stachowski-Haberkorn, 2012				
	28490				36.26	7	De la Broise and Stachowski-Haberkorn, 2012				
Crustacea	<i>Americanysis bahia</i>				NOEC	132500000	168614	4	U.S. EPA, 2013		
					LC50	132500000	168614	4	U.S. EPA, 2013		
Fish	<i>Carassius auratus</i>			NOEC	10000000	1273	0.01	Saglio et al., 2001			
					10000000	1273	0.01	Saglio et al., 2001			
				<i>Oncorhynchus mykiss</i>	NOEC	50000000	6363	4	U.S. EPA, 2013		
						100000000	12726	4	U.S. EPA, 2013		
				Mollusca	<i>Crassostrea virginica</i>	LC50	100000000	12726	4	U.S. EPA, 2013	
						NOEC	100000000	12726	4	U.S. EPA, 2013	
Plants	<i>Lemma gibba</i>			EC50	109000000	138709	4	U.S. EPA, 2013			
				NOEC	1530000	1847	14	U.S. EPA, 2013			
				EC50	5350000	6808	14	U.S. EPA, 2013			
71-43-2	Benzene			Crustacea	<i>Daphnia magna</i>	NOEC	98000000	9906154	21	LeBlanc and Surprenant, 1980	
						98000000	9906154	21	LeBlanc and Surprenant, 1980		
				Fish	<i>Oncorhynchus mykiss</i>	LC50	21639240	218736	4	Hodson et al., 1984	
					Aquatic community	PNEC	8000	809		OSPAR Agreement, 2014-05	
207-08-9	Benzo(k)fluoranthene			Crustacea	<i>Daphnia magna</i>	EC50	46000	4650		U.S. EPA, 1996	
						1400	28.24	0.5	Newsted and Giesy, 1987		





CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference		
2921-88-2	Chlorpyrifos-ethyl	Crustacea	<i>Daphnia magna</i>	LOEC	100	20.17	21	Li and Tan, 2011	
					130	26.22	21	Liu et al., 2012	
					400	8.07	2	Loureiro et al., 2010	
					500	101	21	Li and Tan, 2011	
					500	101	21	Li and Tan, 2011	
					1170	236	21	Liu et al., 2012	
				EC50	32.40	0.65	2	Antunes et al., 2010	
					106	21.38	21	Palma et al., 2009	
					170	34.29	4	Rubach et al., 2011	
					190	38.32	21	Palma et al., 2009	
				250	5.04	2	Brooke, 1995		
				250	50.42	4	Rubach et al., 2011		
				400	8.07	2	Brooke, 1995		
				480	96.81	4	Rubach et al., 2011		
				900	18.15	2	Matsumoto et al., 2009		
				953	19.22	1	Ha and Choi, 2009		
				1400	28.24	2	Matsumoto et al., 2009		
				1720	34.69	2	U.S. EPA, 2013		
				6910	1394	4	Rubach et al., 2011		
				7120	144	2	Liu et al., 2012		
				450000	9076	1	Loureiro et al., 2010		
				580000	1698	2	Loureiro et al., 2010		
				LC50	820	165	4	Rubach et al., 2011	
					4370	881	4	Rubach et al., 2011	
					27430	5532	4	Rubach et al., 2011	
					889000	179300	4	Rubach et al., 2011	
				<i>Gammarus pulex</i>	NOEC	850	17.14	1	Maltby and Hills, 2008
					850	171	6	Maltby and Hills, 2008	
					1280	25.82	1	Maltby and Hills, 2008	
					1280	258	6	Maltby and Hills, 2008	
		LOEC	3700		74.62	1	Maltby and Hills, 2008		
			3700		746	6	Maltby and Hills, 2008		
			5550		112	1	Maltby and Hills, 2008		
			5550		1119	6	Maltby and Hills, 2008		
			35000		7059	105	Van Wijngaarden et al., 1995		
		EC50	230		46.39	4	Rubach et al., 2011		
			240	48.41	4	Rubach et al., 2011			
			380	76.64	4	Rubach et al., 2011			
			3100	625	4	Rubach et al., 2011			
		LC50	230	46.39	4	Rubach et al., 2011			
			230	46.39	4	Rubach et al., 2011			
			430	86.73	4	Rubach et al., 2011			
			3100	625	4	Rubach et al., 2011			
			3400	68.57	2	Ashauer et al., 2007			
			<i>Hyalella azteca</i>	NOEC	14.00	2.82	10	Deanovic et al., 2013	
				66.00	13.31	10	Deanovic et al., 2013		
				66.00	13.31	10	Deanovic et al., 2013		
				66.00	13.31	10	Deanovic et al., 2013		
				66.00	13.31	10	Deanovic et al., 2013		
				66.00	13.31	10	Deanovic et al., 2013		
		LOEC		14.00	2.82	10	Deanovic et al., 2013		
				66.00	13.31	10	Deanovic et al., 2013		
				128	25.82	10	Deanovic et al., 2013		
				133	26.82	10	Deanovic et al., 2013		
		LC50	51.00	10.29	4	Ding et al., 2012			
			103	20.77	10	Deanovic et al., 2013			
			105	21.18	10	Deanovic et al., 2013			
			<i>Hyalella curvispina</i>	LOEC	65.00	1.31	3	Mugni et al., 2010	
				120	2.42	1	Mugni et al., 2010		
			LC50	60.00	1.21	2	Mugni et al., 2012		
			170	3.43	2	Mugni et al., 2012			
			<i>Macrobrachium rosenbergii</i>	NOEC	313	6.30	2	Satapornvanit et al., 2009	
				LOEC	625	12.61	2	Satapornvanit et al., 2009	
				EC50	293	5.91	2	Satapornvanit et al., 2009	
				LC50	300	6.05	2	Satapornvanit et al., 2009	
			<i>Neocaridina denticulata</i>	EC50	171000	34489	4	Rubach et al., 2011	
				237000	47800	4	Rubach et al., 2011		
				327000	65952	4	Rubach et al., 2011		
				410000	82692	4	Rubach et al., 2011		
		LC50		457000	92171	4	Rubach et al., 2011		
				477000	96205	4	Rubach et al., 2011		
				660000	133114	4	Rubach et al., 2011		
				1103000	222461	4	Rubach et al., 2011		
				408000	82289	15	Wang et al., 2012		
				25500	5143	15	Wang et al., 2012		
			Fish	<i>Carassius auratus</i>	NOEC	68000	13715	35	Ali et al., 2009
					15000000	302531	4	Malla et al., 2009a	
				<i>Channa punctata</i>	NOEC	25000000	504219	4	Malla et al., 2009b
					LOEC	68000	13715	35	Ali et al., 2009
					15000000	302531	4	Malla et al., 2009a	
					25000000	504219	4	Malla et al., 2009b	
				<i>Clarias gariepinus</i>	LC50	811980	16377	4	Ali et al., 2009
					NOEC	9200	186	4	Ogueji, 2008
				<i>Cnesterodon decemmaculatus</i>	LC50	920000	18555	4	Ogueji, 2008
					NOEC	1000	20.17	4	Mugni et al., 2012
			<i>Cyprinodon variegatus</i>	<i>Cyprinus carpio</i>	NOEC	5000	101	4	Mugni et al., 2012
					LC50	1000000	20169	2	Mayer, 1987
					1160	234	40	Xing et al., 2012a	
					1160	234	40	Xing et al., 2012a	
					116000	23396	40	Xing et al., 2012b	
					120000	2420	4	Halappa and David, 2009a	
					200000	4034	4	Halappa and David, 2009b	
				LOEC	11600	2340	40	Xing et al., 2012a	
					11600	2340	40	Xing et al., 2012a	
				LC50	160000	3227	4	Halappa and David, 2009a	
			<i>Danio rerio</i>	NOEC	6000	121	0.08	Langer-Jaeschich et al., 2010	
					48000	968	1	Tilton et al., 2011b	
					6500	133	1	Tilton et al., 2011a	
					100000	2017	1	Sledge et al., 2011	
					100000	2017	1	Sledge et al., 2011	
		100000		2017	2	Sledge et al., 2011			
		100000		2017	2	Sledge et al., 2011			
		100000		2017	3	Sledge et al., 2011			
		100000		2017	3	Sledge et al., 2011			
		100000		20169	11	Kienle et al., 2009			
		110000	2219	1	Tilton et al., 2011b				
		126450	2550	1	Tilton et al., 2011a				
		500000	100844	11	Kienle et al., 2009				
		1000000	20169	0.08	Kienle et al., 2009				



CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TECs (ng/L)	Exposure time (days)	Reference			
2921-88-2	Chlorpyrifos-ethyl	Insecta	<i>Culex pipiens</i>	LC50	173000	3489	1	Emtithal and Thanaa, 2012		
					240000	4841	1	Wood et al., 1984		
					262000	5284	1	Emtithal and Thanaa, 2012		
					41000	8289	1	Emtithal and Thanaa, 2012		
					742000	14965	1	Emtithal and Thanaa, 2012		
					1450000	29245	1	Emtithal and Thanaa, 2012		
					1793000	36163	1	Emtithal and Thanaa, 2012		
					7100	143	1	U.S. EPA, 2013		
					1100000	22186	1	Cheng et al., 2009a		
					1300000	26219	1	Cheng et al., 2009b		
					NOEC	1000000	20169	1	Beehler et al., 1991	
					EC50	1630	329	4	Rubach et al., 2011	
						1660	335	4	Rubach et al., 2011	
						3130	631	4	Rubach et al., 2011	
						3430	692	4	Rubach et al., 2011	
						5160	1041	4	Rubach et al., 2011	
						5270	1063	4	Rubach et al., 2011	
						6160	1242	4	Rubach et al., 2011	
						7000	1412	4	Rubach et al., 2011	
						1980	399	4	Rubach et al., 2011	
						LC50	2350	474	4	Rubach et al., 2011
							3290	664	4	Rubach et al., 2011
							6930	1398	4	Rubach et al., 2011
							7000	1412	4	Rubach et al., 2011
							7640	1541	4	Rubach et al., 2011
							8580	1730	4	Rubach et al., 2011
						LC50	4700000	94793	1	Chang et al., 2009
						EC50	180	36.30	4	Rubach et al., 2011
							320	64.54	4	Rubach et al., 2011
							440	88.74	4	Rubach et al., 2011
							860	173	4	Rubach et al., 2011
						LC50	300	60.51	4	Rubach et al., 2011
							610	123	4	Rubach et al., 2011
							1130	228	4	Rubach et al., 2011
							2470	498	4	Rubach et al., 2011
						NOEC	265	53.45	96	LeBlanc et al., 2012
						LOEC	796	161	96	LeBlanc et al., 2012
						EC50	250	50.42	4	Ding et al., 2012
						LC50	290	58.49	4	Ding et al., 2012
							634	128	96	LeBlanc et al., 2012
						NOEC	10.00	2.02	28	Agra and Soares, 2009
							10.00	2.02	28	Agra and Soares, 2009
							50.00	10.08	28	Agra and Soares, 2009
							100	20.17	28	Agra and Soares, 2009
							100	20.17	28	Agra and Soares, 2009
							100	20.17	28	Agra and Soares, 2009
							1000	202	28	Langer-Jaesrich et al., 2010
							200000	4034	1	Park et al., 2012
							1000000	20169	0.08	Kienle et al., 2009
							1000000	20169	0.08	Kienle et al., 2009
							1000000	20169	1	Park et al., 2012
						LOEC	50.00	10.08	28	Agra and Soares, 2009
							50.00	10.08	28	Agra and Soares, 2009
							100	20.17	28	Agra and Soares, 2009
							200000	4034	1	Park et al., 2012
							200000	4034	1	Park et al., 2012
							1000000	20169	1	Park et al., 2012
							2000000	40338	1	Park et al., 2012
						EC50	100	2.02	2	Perez et al., 2013b
							120	2.42	2	Perez et al., 2013b
							130	2.62	2	Perez et al., 2013b
							150	3.03	2	Perez et al., 2013b
							150	3.03	2	Perez et al., 2013b
							160	3.23	2	Perez et al., 2013b
							160	32.27	4	Belden and Lydy, 2006
							170	3.43	2	Perez et al., 2013b
						EC50	310	62.52	4	Rubach et al., 2011
							410	82.69	4	Rubach et al., 2011
							760	153	4	Rubach et al., 2011
							880	177	4	Rubach et al., 2011
						LC50	360	72.61	4	Rubach et al., 2011
							580	117	4	Rubach et al., 2011
							810	163	4	Rubach et al., 2011
							110	224	4	Rubach et al., 2011
						LC50	910	18.35	1	Wirth, 1998
							1600	32.27	1	Chandre et al., 1997
							6000	121	1	Liu et al., 2004
							200000	4034	1	Liu et al., 2004
							280000	5647	1	Khayrandish and Wood, 1993
							450000	9076	1	Khayrandish and Wood, 1993
							620000	12505	1	Khayrandish and Wood, 1993
							900000	18152	1	Liu et al., 2004
							1020000	20572	1	Khayrandish and Wood, 1993
							1060000	21379	1	Khayrandish and Wood, 1993
							1600000	32270	1	Khayrandish and Wood, 1993
							4300000	86726	1	Liu et al., 2004
							11700000	235974	1	Khayrandish and Wood, 1993
						EC50	1860	375	4	Rubach et al., 2011
							1860	375	4	Rubach et al., 2011
						LC50	34200	6898	4	Rubach et al., 2011
							34200	6898	4	Rubach et al., 2011
						EC50	2780	561	4	Rubach et al., 2011
							6060	1222	4	Rubach et al., 2011
							9070	1829	4	Rubach et al., 2011
							19500	3933	4	Rubach et al., 2011
						LC50	7970	1607	4	Rubach et al., 2011
							11600	2340	4	Rubach et al., 2011
							16000	3227	4	Rubach et al., 2011
							23900	4820	4	Rubach et al., 2011
						EC50	2860	577	4	Rubach et al., 2011
			2940	593	4	Rubach et al., 2011				
			3870	781	4	Rubach et al., 2011				
			5880	1186	4	Rubach et al., 2011				
		LC50	27200	5486	4	Rubach et al., 2011				
			29400	5930	4	Rubach et al., 2011				
			31600	6373	4	Rubach et al., 2011				
			55100	1113	4	Rubach et al., 2011				
		NOEC	100000	2017	3	Ke et al., 2009				
		LOEC	1000000	20169	3	Ke et al., 2009				
		Invertebrates	<i>Brachionus calyciflorus</i>	NOEC	100000	2017	3	Ke et al., 2009		
				LOEC	1000000	20169	3	Ke et al., 2009		

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference					
2921-88-2	Chlorpyrifos-ethyl	Mollusca	<i>Coretus corneus</i>	EC50	5000	101	2	Cacciatore et al., 2013				
					6000	121	2	Cacciatore et al., 2013				
					7000	141	2	Cacciatore et al., 2013				
				<i>Lamellidens marginalis</i>	LOEC	5000000	1008438	30	Amanullah et al., 2010			
			<i>Potamopyrgus antipodarum</i>		LOEC	35000	7059	105	Van Wijngaarden et al., 1995			
					NOEC	1543	3111	1	Lundqvist et al., 2012			
				<i>Theodoxus fluviatilis</i>		1543	3111	1	Lundqvist et al., 2012			
						1543	3111	1	Lundqvist et al., 2012			
			Plants		<i>Lemna minor</i>	NOEC	500000	100844	7	Prasertsup and Ariyakanon, 2011		
						500000	100844	7	Prasertsup and Ariyakanon, 2011			
				LOEC		1000000	201688	7	Prasertsup and Ariyakanon, 2011			
				Polyp	<i>Hydra vulgaris</i>	LC50	1000000	201688	7	Prasertsup and Ariyakanon, 2011		
						1000000	201688	7	Prasertsup and Ariyakanon, 2011			
	1500000	302531	4			Demetrio et al., 2012						
115-32-2	Dicofol	Crustacea	<i>Daphnia magna</i>	NOEC	100000	1273	21	Haeba et al., 2008				
				LOEC	100000	1273	21	Haeba et al., 2008				
				EC50	200000	2545	2	Haeba et al., 2008				
					380000	4836	1	Haeba et al., 2008				
					3827265	4870	5	Padilla et al., 2012				
206-44-0	Fluoranthene	Fish	<i>Danio rerio</i>	EC50	41700	335	4	Brooke, 1993				
					41700	335	4	Brooke, 1993				
		Algae	<i>Selenastrum capricornutum</i>	NOEC	17000	136	21	Brooke, 1993				
				EC50	17000	136	21	Brooke, 1993				
		Crustacea	<i>Daphnia magna</i>	NOEC	35300	283	21	Brooke, 1993				
					70500	56.61	2	Brooke, 1993				
				LOEC	35300	283	21	Brooke, 1993				
					73200	588	21	Brooke, 1993				
					170000	93.94	2	Brooke, 1993				
					108000	867	4	Brooke, 1993				
					43800	352	4	Brooke, 1993				
					43500	34.93	4	Brooke, 1993				
					17000	93.94	4	Brooke, 1993				
					90500	72.67	4	Brooke, 1993				
					10400	83.50	32	Brooke, 1993				
					10400	83.50	32	Brooke, 1993				
					21700	174	32	Brooke, 1993				
			21700	174	32	Brooke, 1993						
			212000	170	4	Brooke, 1993						
		Insecta	<i>Aedes aegypti</i>	NOEC	50000	40.15	1	Tetreau et al., 2014				
				LOEC	500	0.40	1	Tetreau et al., 2014				
					500	0.40	1	Tetreau et al., 2014				
		Plants	<i>Chironomus tentans</i>	EC50	2000	161	2	Cho, 2005				
				NOEC	166000	133	4	Brooke, 1993				
				EC50	166000	133	4	Brooke, 1993				
		Polyp	<i>Hydra americana</i>	LC50	70100	563	4	Brooke, 1993				
				NOEC	178000	1429	4	Brooke, 1993				
		Worm	<i>Lumbriculus variegatus</i>	LC50	178000	1429	4	Brooke, 1993				
					178000	1429	4	Brooke, 1993				
			Aquatic community	PNEC	6.30	0.051		OSPAR Agreement, 2014-05				
					10.00	0.080		EU Risk Assessment Report, 2008				
		91-20-3	Naphtalene	Crustacea	<i>Daphnia magna</i>	NOEC	480000	9681	2	U.S. EPA, 2013		
						EC50	1600000	32270	2	U.S. EPA, 2013		
						LC50	4438000	89509	2	Schirmer and Knoebel, 2012		
							4903000	98887	2	Schirmer and Knoebel, 2012		
							4903000	98887	3	Schirmer and Knoebel, 2012		
							7499000	15245	2	Schirmer and Knoebel, 2012		
							7499000	15245	4	Schirmer and Knoebel, 2012		
							8464000	170708	4	Schirmer and Knoebel, 2012		
							8804000	177566	2	Schirmer and Knoebel, 2012		
							8920000	179905	2	Scholz S., 2012		
							11430000	230529	3	Schirmer and Knoebel, 2012		
							11430000	230529	4	Schirmer and Knoebel, 2012		
							11930000	240613	2	Schirmer and Knoebel, 2012		
							12220000	246462	2	Schirmer and Knoebel, 2012		
							12220000	246462	3	Schirmer and Knoebel, 2012		
							12330000	248681	2	Schirmer and Knoebel, 2012		
	14020000					282766	2	Schirmer and Knoebel, 2012				
	14020000					282766	3	Schirmer and Knoebel, 2012				
	1400000					28236	4	U.S. EPA, 2013				
	3200000					64540	4	U.S. EPA, 2013				
	4000000					8068	4	U.S. EPA, 2013				
	670000					135131	40	U.S. EPA, 2013				
	670000					135131	40	U.S. EPA, 2013				
	860000					17345	4	U.S. EPA, 2013				
	10000000					2016875	4	U.S. EPA, 2013				
	2000000					40338	4	U.S. EPA, 2013				
	10000000					2016875	4	U.S. EPA, 2013				
	Aquatic community					PNEC	2000	403		OSPAR Agreement, 2014-05		
							2000	403		EU Risk Assessment Report, 2008		
84852-15-3	Nonylphenol technical mixture					Fish	<i>Danio rerio</i>	NOEC	2500	79.91	3	Jin et al., 2010
									10000	320	2	Jin et al., 2009
									12500	400	3	Jin et al., 2010
									100000	3197	2	Jin et al., 2009
			1000000	31965	2			Kammann et al., 2009				
			1000000	31965	2			Kammann et al., 2009				
			1200000	38358	2			Kammann et al., 2009				
			1400000	44751	2			Kammann et al., 2009				
			1500000	47948	2			Kammann et al., 2009				
			2000000	63931	2			Kammann et al., 2009				
			10000	320	2			Jin et al., 2009				
			12500	400	3			Jin et al., 2010				
			100000	3197	2			Jin et al., 2009				
			1000000	63931	2			Kammann et al., 2009				
			2100000	67127	2			Kammann et al., 2009				
			4400000	140647	2			Kammann et al., 2009				
			<i>Oncorhynchus mykiss</i>	NOEC	2300			73.52	4	Shelley et al., 2012		
				18000	575			4	Shelley et al., 2012			
				18000	575			4	Shelley et al., 2012			
				18000	575			4	Shelley et al., 2012			
				18000	575			4	Shelley et al., 2012			
				18000	575			4	Shelley et al., 2012			
				18000	575			4	Shelley et al., 2012			
			LOEC	18000	575			4	Shelley et al., 2012			
				18000	575			4	Shelley et al., 2012			
		122-34-9	Simazine	Algae	<i>Anabaena flosaquae</i>			EC50	36000	7261	5	U.S. EPA, 2013
								EC50	82000	16538	4	Ma et al., 2002b
NOEC	30000					6051	5	U.S. EPA, 2013				
EC50	90000					18152	5	U.S. EPA, 2013				
NOEC	30000					6051	5	U.S. EPA, 2013				
	32000					645	3	Perez et al., 2011b				
LOEC	100000					2017	3	Perez et al., 2011b				
EC50	100000					20169	5	U.S. EPA, 2013				
		241000	4861	3	Perez et al., 2011b							

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference				
122-34-9	Simazine	Algae	<i>Selenastrum capricornutum</i>	EC50	252000	5083	3	Perez et al., 2011b			
					392000	7906	3	Perez et al., 2011b			
					748500	150963	4	Ma et al., 2006			
					250000	50422	5	U.S. EPA, 2013			
					600000	121013	5	U.S. EPA, 2013			
					50000	10084	84	Vervliet-Scheebaum et al., 1993			
					500000	100844	84	Vervliet-Scheebaum et al., 1993			
					2500000	504219	21	U.S. EPA, 2013			
					10000000	2016875	2	U.S. EPA, 2013			
					2500000	504219	365	U.S. EPA, 2013			
			Crustacea	<i>Daphnia magna</i>	LOEC	2500000	504219	21	U.S. EPA, 2013		
					10000000	2016875	2	U.S. EPA, 2013			
					2500000	504219	365	U.S. EPA, 2013			
					32000000	645400	4	U.S. EPA, 2013			
					4300000	86726	4	U.S. EPA, 2013			
					4300000	86726	4	U.S. EPA, 2013			
					60.00	12.10	36	Velisek et al., 2012b			
					60.00	12.10	60	Stara et al., 2012			
					60.00	12.10	60	Stara et al., 2012			
					60.00	12.10	90	Velisek et al., 2012a			
			60.00	12.10	90	Velisek et al., 2012a					
			2000	403	90	Velisek et al., 2012a					
			4000	807	90	Velisek et al., 2012a					
			4000	807	90	Velisek et al., 2012a					
			4000	807	90	Velisek et al., 2012a					
			4000	807	90	Velisek et al., 2012a					
			4000	807	90	Velisek et al., 2012a					
			4000	807	90	Velisek et al., 2012a					
			4000	807	90	Velisek et al., 2012a					
			4000	807	90	Velisek et al., 2012a					
			60000	12101	36	Velisek et al., 2012b					
			600000	121013	36	Velisek et al., 2012b					
			2000000	403375	60	Stara et al., 2012					
			2000000	403375	60	Stara et al., 2012					
			3000000	605063	36	Velisek et al., 2012b					
			3000000	605063	36	Velisek et al., 2012b					
			4000000	806750	60	Stara et al., 2012					
			4000000	806750	60	Stara et al., 2012					
			4000000	806750	60	Stara et al., 2012					
			LOEC	60.00	12.10	60	Stara et al., 2012				
				60.00	12.10	60	Stara et al., 2012				
				60.00	12.10	90	Velisek et al., 2012a				
				1000	202	90	Velisek et al., 2012a				
				1000	202	90	Velisek et al., 2012a				
				4000	807	90	Velisek et al., 2012a				
				4000	807	90	Velisek et al., 2012a				
				60000	12101	36	Velisek et al., 2012b				
				600000	121013	36	Velisek et al., 2012b				
				2000000	403375	60	Stara et al., 2012				
				2000000	403375	60	Stara et al., 2012				
				3000000	605063	36	Velisek et al., 2012b				
				4000000	806750	60	Stara et al., 2012				
				4000000	806750	60	Stara et al., 2012				
			Fish	<i>Carassius auratus</i>	LC50	32000000	645400	4	U.S. EPA, 2013		
					NOEC	4300000	86726	4	U.S. EPA, 2013		
					LC50	4300000	86726	4	U.S. EPA, 2013		
					<i>Cyprinodon variegatus</i>	NOEC	4300000	86726	4	U.S. EPA, 2013	
						LC50	4300000	86726	4	U.S. EPA, 2013	
						<i>Cyprinus carpio</i>	NOEC	60.00	12.10	36	Velisek et al., 2012b
								60.00	12.10	60	Stara et al., 2012
								60.00	12.10	60	Stara et al., 2012
								60.00	12.10	90	Velisek et al., 2012a
								60.00	12.10	90	Velisek et al., 2012a
				2000			403	90	Velisek et al., 2012a		
				4000			807	90	Velisek et al., 2012a		
				4000			807	90	Velisek et al., 2012a		
				4000	807		90	Velisek et al., 2012a			
				4000	807		90	Velisek et al., 2012a			
				60000	12101	36	Velisek et al., 2012b				
				600000	121013	36	Velisek et al., 2012b				
				2000000	403375	60	Stara et al., 2012				
				2000000	403375	60	Stara et al., 2012				
				3000000	605063	36	Velisek et al., 2012b				
				3000000	605063	36	Velisek et al., 2012b				
				4000000	806750	60	Stara et al., 2012				
				4000000	806750	60	Stara et al., 2012				
				LOEC	60.00	12.10	60	Stara et al., 2012			
					60.00	12.10	60	Stara et al., 2012			
					60.00	12.10	90	Velisek et al., 2012a			
					1000	202	90	Velisek et al., 2012a			
					1000	202	90	Velisek et al., 2012a			
					4000	807	90	Velisek et al., 2012a			
					4000	807	90	Velisek et al., 2012a			
					60000	12101	36	Velisek et al., 2012b			
					600000	121013	36	Velisek et al., 2012b			
					2000000	403375	60	Stara et al., 2012			
					2000000	403375	60	Stara et al., 2012			
					3000000	605063	36	Velisek et al., 2012b			
					4000000	806750	60	Stara et al., 2012			
					4000000	806750	60	Stara et al., 2012			
			Mollusca	<i>Danio rerio</i>	NOEC	6000	1210	28	Pihalova et al., 2011		
					60000	12101	28	Pihalova et al., 2011			
					10000000	201688	4	Sun et al., 2010b			
					LOEC	60000	12101	28	Pihalova et al., 2011		
					EC50	800000	16135	4	Sun et al., 2010b		
					NOEC	2500000	504219	365	U.S. EPA, 2013		
						5600000	112945	4	U.S. EPA, 2013		
					<i>Morone saxatilis</i>	NOEC	1000000	20169	4	U.S. EPA, 2013	
						LC50	3000000	60506	4	U.S. EPA, 2013	
					<i>Oncorhynchus mykiss</i>	NOEC	6000000	121013	4	U.S. EPA, 2013	
		10000000	201688	4		U.S. EPA, 2013					
		22000000	443713	4		U.S. EPA, 2013					
		28600000	576826	4		U.S. EPA, 2013					
		34300000	691788	4		U.S. EPA, 2013					
	LC50	2500000	504219	28		U.S. EPA, 2013					
		10000000	201688	4		U.S. EPA, 2013					
		40500000	816834	4		U.S. EPA, 2013					
		44600000	899526	4		U.S. EPA, 2013					
		60000000	1210125	4		U.S. EPA, 2013					
		70500000	1421897	4	U.S. EPA, 2013						
		82000000	1653838	4	U.S. EPA, 2013						
	<i>Pimephales promelas</i>	NOEC	2500000	50422	4	U.S. EPA, 2013					
		LOEC	2500000	504219	120	U.S. EPA, 2013					
		LC50	6400000	129080	4	U.S. EPA, 2013					
	Plants	<i>Crassostrea virginica</i>	NOEC	1000000	201688	7	U.S. EPA, 2013				
				3700000	746244	4	U.S. EPA, 2013				
			EC50	1000000	201688	7	U.S. EPA, 2013				
	<i>Elodea canadensis</i>	Plants	NOEC	50000	10084	14	U.S. EPA, 2013				
			EC50	140000	28236	14	U.S. EPA, 2013				
				83000	16740	84	Vervliet-Scheebaum et al., 1993				
				83000	16740	84	Vervliet-Scheebaum et al., 1993				
				110000	223873	84	Vervliet-Scheebaum et al., 1993				
				110000	223873	84	Vervliet-Scheebaum et al., 1993				
				110000	223873	84	Vervliet-Scheebaum et al., 1993				
				8470000	1708293	84	Vervliet-Scheebaum et al., 1993				
				8470000	1708293	84	Vervliet-Scheebaum et al., 1993				
				83000	16740	84	Vervliet-Scheebaum et al., 1993				
	<i>Myriophyllum spicatum</i>	Plants	NOEC	50000	10084	84	Vervliet-Scheebaum et al., 1993				
				83000	16740	84	Vervliet-Scheebaum et al., 1993				
				83000	16740	84	Vervliet-Scheebaum et al., 1993				
				110000	223873	84	Vervliet-Scheebaum et al., 1993				
				110000	223873	84	Vervliet-Scheebaum et al., 1993				
				110000	223873	84	Vervliet-Scheebaum et al., 1993				
				8470000	1708293	84	Vervliet-Scheebaum et al., 1993				
				8470000	1708293	84	Vervliet-Scheebaum et al., 1993				
				500000	100844	84	Vervliet-Scheebaum et al., 1993				
				1100000	223873	84	Vervliet-Scheebaum et al., 1993				
		1100000	223873	84	Vervliet-Scheebaum et al., 1993						
		8470000	1708293	84	Vervliet-Scheebaum et al., 1993						
		8470000	1708293	84	Vervliet-Scheebaum et al., 1993						
		LOEC	500000	100844	84	Vervliet-Scheebaum et al., 1993					
			1100000	223873	84	Vervliet-Scheebaum et al., 1993					
			1100000	223873	84	Vervliet-Scheebaum et al., 1993					
			8470000	1708293	84	Vervliet-Scheebaum et al., 1993					
			8470000	1708293	84	Vervliet-Scheebaum et al., 1993					

## ANTIBIOTIC ACTIVITY

### I.1 AMINOGLYCOSIDES

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TECs (ng/L)	Exposure time (days)	Reference	
1263-89-4	Aminosidine	Crustacea	<i>Artemia franciscana</i>	LC50	846500000	846500000	3	Migliore et al., 1996
				LC50	2220000000	2220000000	2	Migliore et al., 1996
128-46-1	Dihydrostreptomycin	Algae	<i>Selenastrum capricornutum</i>	NOEC	39000	7800		Eguchi et al., 2004
				EC50	107000	21400		Eguchi et al., 2004
1404-04-2	Neomycin	Bacteria	<i>Vibrio fischeri</i>	EC50	1000000000	1000000000	0.01	Park and Choi, 2008
				EC50	1000000000	1000000000	0.03	Park and Choi, 2008
				NOEC	30000	30000	21	Park and Choi, 2008
		Crustacea	<i>Daphnia magna</i>	LOEC	10000	10000	21	Park and Choi, 2008
				EC50	90000	90000	21	Park and Choi, 2008
				EC50	42100000	42100000	2	Park and Choi, 2008
				EC50	116600000	116600000	1	Park and Choi, 2008
				NOEC	500000	500000	8	Park and Choi, 2008
				LOEC	1600000	1600000	8	Park and Choi, 2008
		Fish	<i>Oryzias latipes</i>	EC50	740000	740000	8	Park and Choi, 2008
				EC50	34100000	34100000	2	Park and Choi, 2008
				EC50	61900000	61900000	1	Park and Choi, 2008
				LC50	808000000	808000000	4	Park and Choi, 2008
				LC50	1388000000	1388000000	2	Park and Choi, 2008
				PNEC	300	300		Park and Choi, 2008
57-92-1	Streptomycin	Algae	Aquatic community	EC50	20080000	160640000	4	Qian et al., 2010
				EC50	7000	56000	7	Halling-Sorensen, 2000
				EC50	34000	27200	1	Grinten et al., 2010
		<i>Chlorella vulgaris</i>	EC50	280000	2240000	4	Qian et al., 2010	
			EC50	133000	1064000	3	Halling-Sorensen, 2000	
			EC50	1500000	1200000	1	Grinten et al., 2010	
		Bacteria	<i>Selenastrum capricornutum</i>	EC50	2900000	23200000	1	Grinten et al., 2010
				EC50	2900000	23200000	1	Grinten et al., 2010
				EC50	2900000	23200000	1	Grinten et al., 2010
				EC50	400000	3200000	0.02	Grinten et al., 2010
				EC50	8210000	65680000	1	Backhaus and Grimme, 1999
				EC50	2900000	23200000	1	Grinten et al., 2010
		Crustacea	<i>Daphnia magna</i>	EC50	487000000	389600000	2	Wollenberger et al., 2000
				EC50	947000000	757600000	1	Wollenberger et al., 2000

### I.2 MACROLIDES & B-LACTAMS

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TECs (ng/L)	Exposure time (days)	Reference	
26787-78-0	Amoxicillin	Algae	<i>Microcystis aeruginosa</i>	EC50	3700	2220	7	Holten Lützhof et al., 1999
				EC50	3108000	1864800	7	Holten Lützhof et al., 1999
				NOEC	250000000	150000000	7	Holten Lützhof et al., 1999
		Bacteria	<i>Selenastrum capricornutum</i>	EC50	1320000000	792000000	0.01	Park and Choi, 2008
				EC50	3597000000	2158200000	0.01	Park and Choi, 2008
		Crustacea	<i>Vibrio fischeri</i>	EC50	1000000000	600000000	1	Park and Choi, 2008
				EC50	1000000000	600000000	2	Park and Choi, 2008
				EC50	1000000000	600000000	1	Park and Choi, 2008
		Fish	<i>Daphnia magna</i>	EC50	1000000000	600000000	2	Park and Choi, 2008
				EC50	1000000000	600000000	2	Park and Choi, 2008
				EC50	1000000000	600000000	4	Park and Choi, 2008
		Plants	<i>Moina macrocopa</i>	LOEC	1000000	60000		Park and Choi, 2008
				EC50	3700	222		Park and Choi, 2008
				PNEC	3.70	2.22		Jones et al., 2002
		69-53-4	Ampicillin	Algae	Aquatic community	EC50	250000	150000
NOEC	1000000000					600000000		Eguchi et al., 2004
EC50	1000000000					600000000		Eguchi et al., 2004
Bacteria	<i>Chlorella vulgaris</i>			NOEC	1000000000	600000000		Eguchi et al., 2004
				EC50	1000000000	600000000		Eguchi et al., 2004
				EC50	1000000000	600000000		Eguchi et al., 2004
Crustacea	<i>Selenastrum capricornutum</i>			EC50	1630000000	978000000	1	Backhaus and Grimme, 1999
				EC50	1056000000	633600000	0.003	Park and Choi, 2008
				EC50	2627000000	1576200000	0.01	Park and Choi, 2008
				EC50	1000000000	600000000	1	Park and Choi, 2008
				EC50	1000000000	600000000	2	Park and Choi, 2008
				EC50	1000000000	600000000	1	Park and Choi, 2008
Fish	<i>Daphnia magna</i>			EC50	1000000000	600000000	2	Park and Choi, 2008
				EC50	1000000000	600000000	2	Park and Choi, 2008
				EC50	1000000000	600000000	4	Park and Choi, 2008
1405-87-4	Bacitracin	Crustacea	Aquatic community	PNEC	163000	97800		Park and Choi, 2008
				LC50	21820000	81825	2	Ferreira et al., 2007
				EC50	21820000	81825	2	Migliore et al., 1996
61-33-6	Benzylpenicillin	Algae	<i>Artemia nauplii</i>	EC50	340600000	127725	1	Migliore et al., 1996
				EC50	6000	3600	7	Halling-Sorensen, 2000
				NOEC	1000000000	600000000	3	Halling-Sorensen, 2000
114-07-8	Erythromycin	Algae	<i>Selenastrum capricornutum</i>	NOEC	12500000	375000		Eguchi et al., 2004
				EC50	33800000	1040000		Eguchi et al., 2004
				NOEC	10300	309		Eguchi et al., 2004
EC50	20000	600	3	Isidori et al., 2005				

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference	
				36600	1098		Eguchi et al., 2004	
		Bacteria	<i>Vibrio fischeri</i>	EC50	100000000	3000000	0.02	Isidori et al., 2005
		Crustacea	<i>Ceriodaphnia dubia</i>	EC50	220000	66000	7	Isidori et al., 2005
					10230000	306900	2	Isidori et al., 2005
			<i>Daphnia magna Straus</i>	EC50	22450000	673500	1	Isidori et al., 2005
			<i>Thamnocephalus platyurus</i>	LC50	17680000	530400	1	Isidori et al., 2005
		Fish	<i>Danio rerio</i>	LC50	1000000000	30000000	4	Isidori et al., 2005
		Invertebrates	<i>Brachionus calyciflorus</i>	EC50	940000	28200	2	Isidori et al., 2005
				LC50	27530000	825900	1	Isidori et al., 2005
		Plants	<i>Lemna gibba</i>	LOEC	1000000	300000	7	Brain et al., 2004
			<i>Lemna minor</i>	EC50	5620000	168600	7	Pomati et al., 2004
			Aquatic community	PNEC	103	3.09	3	Yang et al., 2011
					22700	6810		Jones et al., 2002
154-21-2	Lincomycin	Algae	<i>Selenastrum capricornutum</i>	EC50	70000	525	3	Isidori et al., 2005
		Bacteria	<i>Vibrio fischeri</i>	EC50	100000000	750000	0.02	Isidori et al., 2005
		Crustacea	<i>Artemia franciscana</i>	LC50	283100000	2123250	3	Ferreira et al., 2007
			<i>Artemia nauplii</i>	EC50	283100000	2123250	3	Migliore et al., 1996
			<i>Ceriodaphnia dubia</i>	EC50	7200000	540000	7	Isidori et al., 2005
					13980000	104850	2	Isidori et al., 2005
			<i>Daphnia magna Straus</i>	EC50	23180000	173850	1	Isidori et al., 2005
			<i>Thamnocephalus platyurus</i>	LC50	30000000	225000	1	Isidori et al., 2005
		Fish	<i>Danio rerio</i>	LC50	1000000000	7500000	4	Isidori et al., 2005
		Invertebrates	<i>Brachionus calyciflorus</i>	EC50	680000	5100	2	Isidori et al., 2005
				LC50	24940000	187050	1	Isidori et al., 2005
		Plants	<i>Lemna gibba</i>	LOEC	100000	7500	7	Brain et al., 2004
					100000	7500	7	Brain et al., 2004
					100000	7500	7	Brain et al., 2004
					300000	22500	7	Brain et al., 2004
					1000000	75000	7	Brain et al., 2004
303-81-1	Novobiocin	Polyp	<i>Hydra attenuata</i>	NOEC	50000000	937500	4	Quinn et al., 2007
				LOEC	100000000	1875000	4	Quinn et al., 2007
				EC50	13530000	253688	4	Quinn et al., 2007
				LC50	100000000	1875000	4	Quinn et al., 2007
87-08-1	Phenoxymethylpenicillin		Aquatic community	PNEC	177000	53100		Jones et al., 2002
8025-81-8	Spiramycin	Algae	<i>Microcystis aeruginosa</i>	EC50	5000	250	7	Halling-Sorensen, 2000
			<i>Selenastrum capricornutum</i>	EC50	23000000	11500	3	Halling-Sorensen, 2000
55297-95-5	Tiamulin	Algae	<i>Microcystis aeruginosa</i>	EC50	3000	18.00	7	Halling-Sorensen, 2000
			<i>Selenastrum capricornutum</i>	EC50	165000	99	3	Halling-Sorensen, 2000
		Crustacea	<i>Daphnia magna</i>	EC50	5400000	32400	21	Wollenberger et al., 2000
					40000000	24000	2	Wollenberger et al., 2000
					81000000	48600	1	Wollenberger et al., 2000
1401-69-0	Tylosin	Algae	<i>Microcystis aeruginosa</i>	EC50	34000	1275	7	Halling-Sorensen, 2000
					290000	1088	1	Grinten et al., 2010
			<i>Selenastrum capricornutum</i>	NOEC	64000	240	3	Yang et al., 2008
				LOEC	206000	773		Eguchi et al., 2004
				EC50	64000	240	3	Yang et al., 2008
				EC50	8900	33.38	1	Grinten et al., 2010
					210000	788	3	Yang et al., 2008
					411000	1541		Eguchi et al., 2004
					950000	3563		Yang et al., 2008
					1380000	5175	3	Halling-Sorensen, 2000
					1380000	5175		Yang et al., 2008
		Bacteria	<i>Bacillus cereus</i>	EC50	3100000	116250	1	Grinten et al., 2010
			<i>Bacillus pumilus</i>	EC50	1090000	40875	1	Grinten et al., 2010
			<i>Micrococcus luteus</i>	EC50	570000	21375	1	Grinten et al., 2010
			<i>Vibrio fischeri</i>	EC50	1800000	6750	0.02	Grinten et al., 2010
			<i>Yersinia ruckeri</i>	EC50	3100000	116250	1	Grinten et al., 2010
		Crustacea	<i>Daphnia magna</i>	LOEC	700000000	2625000	1	Wollenberger et al., 2000
				EC50	680000000	2550000	2	Wollenberger et al., 2000
		Plants	<i>Lemna gibba</i>	LOEC	300000	11250	7	Brain et al., 2004
					1000000	37500	7	Brain et al., 2004
					1000000	37500	7	Brain et al., 2004



### I.3 SULFONAMIDES

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEOs (ng/L)	Exposure time (days)	Reference		
80-32-0	Sulfachlorpyridazine	Algae	<i>Chlorella vulgaris</i>	EC50	200000000	100000000	2	Pro et al., 2003	
			Bacteria	<i>Vibrio fischeri</i>	EC50	264000000	132000000	0.01	Kim et al., 2007
			Crustacea	<i>Daphnia magna</i>	LC50	233500000	116750000	4	Kim et al., 2007
		Fish	<i>Oryzias latipes</i>	LC50	375300000	187650000	2	Kim et al., 2007	
				LC50	535700000	267850000	4	Kim et al., 2007	
				LC50	589300000	294650000	2	Kim et al., 2007	
				EC50	2330000	1165000	7	Pro et al., 2003	
				EC50	1226000	61300	2	Baran et al., 2006	
				EC50	135000	67500	7	Holten Lützhof et al., 1999	
				EC50	403000000	201500000	2	Holten Lützhof et al., 1999	
68-35-9	Sulfadiazine	Algae	<i>Microcystis aeruginosa</i>	EC50	1000000	500000	1	Eguchi et al., 2004	
			<i>Rhodomonas salina</i>	EC50	403000000	201500000	2	Holten Lützhof et al., 1999	
			<i>Selenastrum capricornutum</i>	NOEC	1000000	500000	1	Eguchi et al., 2004	
			EC50	2490000	1095000	2	Eguchi et al., 2004		
			EC50	7800000	3900000	2	Holten Lützhof et al., 1999		
		Crustacea	<i>Daphnia magna</i>	LOEC	150000000	75000000	1	Wollenberger et al., 2000	
				EC50	137000000	68500000	21	Wollenberger et al., 2000	
				EC50	212000000	106000000	2	Liguoro et al., 2009	
				EC50	221000000	110500000	2	Wollenberger et al., 2000	
				NOEC	1000000	500000	2	Lin et al., Manuscript Draft	
Fish	<i>Danio rerio</i>	LOEC	1000000	500000	0.3	Lin et al., Manuscript Draft			
		LOEC	1000000	500000	1	Lin et al., Manuscript Draft			
		LOEC	1000000	500000	4	Lin et al., Manuscript Draft			
		LOEC	1000000	500000	2	Lin et al., Manuscript Draft			
		LOEC	1000000	500000	2	Lin et al., Manuscript Draft			
122-11-2	Sulfadimethoxine	Algae	Aquatic community	PNEC	1200000	1060000	2	Yang et al., 2011	
			<i>Chlorella vulgaris</i>	NOEC	203000000	203000000	2	Eguchi et al., 2004	
			<i>Selenastrum capricornutum</i>	EC50	112000000	112000000	2	Eguchi et al., 2004	
			NOEC	5290000	5290000	2	Eguchi et al., 2004		
			EC50	230000000	230000000	2	Eguchi et al., 2004		
		Bacteria	<i>Vibrio fischeri</i>	EC50	500000000	500000000	0.01	Kim et al., 2007	
				EC50	204500000	204500000	4	Kim et al., 2007	
				EC50	248000000	248000000	2	Kim et al., 2007	
				EC50	270000000	270000000	2	Liguoro et al., 2009	
				EC50	639800000	639800000	1	Park and Choi, 2008	
Crustacea	<i>Moina macrocopa</i>	EC50	183900000	183900000	2	Park and Choi, 2008			
		EC50	296600000	296600000	1	Park and Choi, 2008			
		LC50	100000000	100000000	2	Kim et al., 2007			
		LC50	100000000	100000000	4	Kim et al., 2007			
		LC50	500000000	500000000	2	Park and Choi, 2008			
Plants	<i>Lenna gibba</i>	LOEC	3000000	3000000	4	Park and Choi, 2008			
		LOEC	3000000	3000000	7	Brain et al., 2004			
		LOEC	3000000	3000000	7	Brain et al., 2004			
		LOEC	10000000	10000000	7	Brain et al., 2004			
		LOEC	10000000	10000000	7	Brain et al., 2004			
		LOEC	10000000	10000000	7	Brain et al., 2004			
		LOEC	10000000	10000000	7	Brain et al., 2004			
		LOEC	10000000	10000000	7	Brain et al., 2004			
		EC50	2480000	2480000	7	Brain et al., 2004			
		EC50	4450000	4450000	7	Brain et al., 2004			
57-68-1	Sulfamethazine	Algae	Aquatic community	PNEC	248	248	7	Brain et al., 2004	
			<i>Selenastrum capricornutum</i>	NOEC	1000000	500000	3	Yang et al., 2008	
			NOEC	8000000	4000000	3	Yang et al., 2008		
			NOEC	8700000	4350000	3	Yang et al., 2008		
			EC50	344700000	172350000	0.01	Kim et al., 2007		
		Bacteria	<i>Vibrio fischeri</i>	NOEC	1563000	781500	21	Liguoro et al., 2009	
				NOEC	3125000	1562500	21	Liguoro et al., 2009	
				NOEC	4250000	2125000	21	Liguoro et al., 2009	
				NOEC	14750000	7375000	4	Jung et al., 2008	
				NOEC	18530000	9265000	2	Jung et al., 2008	
Crustacea	<i>Daphnia magna</i>	LOEC	202000000	101000000	2	Liguoro et al., 2009			
		LOEC	215900000	107950000	2	Park and Choi, 2008			
		LOEC	506300000	253150000	1	Park and Choi, 2008			
		LC50	158800000	79400000	4	Kim et al., 2007			
		LC50	174400000	87200000	2	Kim et al., 2007			
723-46-6	Sulfamethoxazole	Algae	<i>Moina macrocopa</i>	EC50	110700000	55350000	2	Park and Choi, 2008	
			<i>Oryzias latipes</i>	LC50	310900000	155450000	1	Park and Choi, 2008	
			LC50	100000000	50000000	2	Kim et al., 2007		
			LC50	100000000	50000000	4	Kim et al., 2007		
			LC50	500000000	250000000	2	Park and Choi, 2008		
		Plants	<i>Lenna gibba</i>	LOEC	500000000	250000000	4	Park and Choi, 2008	
				LOEC	10000000	5000000	7	Brain et al., 2004	
				LOEC	1277000	638500	7	Brain et al., 2004	
				PNEC	1277	639	2	Park and Choi, 2008	
				PNEC	2020000	1010000	2	Yang et al., 2011	
723-46-6	Sulfamethoxazole	Algae	<i>Chlorella vulgaris</i>	EC50	1570000	1570000	2	Baran et al., 2006	
			<i>Cyclotella meneghiniana</i>	NOEC	1250000	1250000	4	Ferrari et al., 2004	
			<i>Microcystis aeruginosa</i>	EC50	2400000	2400000	4	Ferrari et al., 2004	
			<i>Selenastrum capricornutum</i>	EC50	550000	550000	1	Grinten et al., 2010	
			NOEC	900000	900000	4	Ferrari et al., 2004		
		Bacteria	<i>Vibrio fischeri</i>	NOEC	5000000	5000000	3	Yang et al., 2008	
				NOEC	6140000	6140000	3	Eguchi et al., 2004	
				LOEC	8000000	8000000	3	Yang et al., 2008	
				EC50	1460000	1460000	4	Ferrari et al., 2004	
				EC50	1460000	1460000	4	Yang et al., 2008	
Amphibia	<i>Xenopus laevis</i>	Bacteria	<i>Bacillus cereus</i>	EC50	520000	520000	3	Isidori et al., 2005	
			<i>Bacillus pumilus</i>	EC50	520000	520000	3	Yang et al., 2008	
			<i>Micrococcus luteus</i>	EC50	1500000	1500000	1	Yang et al., 2008	
			<i>Vibrio fischeri</i>	EC50	1500000	1500000	0.02	Grinten et al., 2010	
			EC50	233000000	233000000	0.02	Isidori et al., 2005		
		Synechococcus	<i>Synechococcus leopoliensis</i>	NOEC	78100000	78100000	0.01	Kim et al., 2007	
				NOEC	5900	5900	4	Ferrari et al., 2004	
				EC50	26800	26800	4	Ferrari et al., 2004	
				NOEC	100000000	100000000	4	Richards and Cole, 2006	
				NOEC	100000000	100000000	4	Richards and Cole, 2006	

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference
				84000000	8400000	0.02	Ferrari et al., 2004
		Crustacea	<i>Yersinia ruckeri</i>	EC50	1500000	1	Grinten et al., 2010
			<i>Ceriodaphnia dubia</i>	NOEC	250000	7	Ferrari et al., 2004
				EC50	210000	7	Isidori et al., 2005
					15510000	2	Isidori et al., 2005
					100000000	2	Ferrari et al., 2004
			<i>Daphnia magna</i>	EC50	25200000	1	Isidori et al., 2005
					100000000	2	Ferrari et al., 2004
					123100000	2	Park and Choi, 2008
					177600000	4	Jung et al., 2008
					200000000	1	Park and Choi, 2008
					205200000	2	Jung et al., 2008
					177300000	4	Kim et al., 2007
					189200000	2	Kim et al., 2007
			<i>Moina macrocopa</i>	EC50	70400000	2	Park and Choi, 2008
					84900000	1	Park and Choi, 2008
		Fish	<i>Thamnocephalus platyurus</i>	LC50	35360000	1	Isidori et al., 2005
			<i>Danio rerio</i>	NOEC	1000	1	Lin et al., Manuscript Draft
				LOEC	8000000	10	Ferrari et al., 2004
					1000	0.3	Lin et al., Manuscript Draft
					1000	2	Lin et al., Manuscript Draft
					100000	1	Lin et al., Manuscript Draft
					10000000	4	Lin et al., Manuscript Draft
					1000000000	4	Isidori et al., 2005
			<i>Oryzias latipes</i>	LC50	562500000	4	Kim et al., 2007
					750000000	2	Kim et al., 2007
		Invertebrates	<i>Brachionus calyciflorus</i>	NOEC	25000000	2	Ferrari et al., 2004
				LC50	9630000	2	Isidori et al., 2005
					26270000	1	Isidori et al., 2005
		Plants	<i>Lemna gibba</i>	LOEC	30000	7	Brain et al., 2004
					100000	7	Brain et al., 2004
					100000	7	Brain et al., 2004
					100000	7	Brain et al., 2004
					300000	7	Brain et al., 2004
				EC50	81000	7	Brain et al., 2004
					249000	7	Brain et al., 2004
					682000	7	Brain et al., 2004
					985000	7	Brain et al., 2004
					4963000	7	Brain et al., 2004
		Polyp	<i>Hydra attenuata</i>	NOEC	5000000	4	Quinn et al., 2007
				LOEC	10000000	4	Quinn et al., 2007
				LC50	100000000	4	Quinn et al., 2007
			Aquatic community	PNEC	30.00		Park and Choi, 2008
					146	4	Kim et al., 2007
					189000	2	Yang et al., 2011
144-83-2	Sulfapyridine	Polyp	<i>Hydra attenuata</i>	NOEC	1000000	4	Quinn et al., 2007
				LOEC	5000000	4	Quinn et al., 2007
				EC50	21610000	4	Quinn et al., 2007
				LC50	100000000	4	Quinn et al., 2007
			Aquatic community	PNEC	10000	4	Yang et al., 2011
72-14-0	Sulfathiazole	Algae	<i>Chlorella vulgaris</i>	EC50	16340000	2	Baran et al., 2006
		Bacteria	<i>Vibrio fischeri</i>	EC50	1000000000	0.01	Kim et al., 2007
		Crustacea	<i>Daphnia magna</i>	NOEC	11000000	21	Park and Choi, 2008
				LOEC	35000000	21	Park and Choi, 2008
				EC50	78900000	4	Jung et al., 2008
					135700000	2	Jung et al., 2008
					616700000	1	Park and Choi, 2008
				LC50	854000000	4	Kim et al., 2007
					1493000000	2	Kim et al., 2007
			<i>Moina macrocopa</i>	EC50	391100000	2	Park and Choi, 2008
					430100000	1	Park and Choi, 2008
		Fish	<i>Oryzias latipes</i>	LC50	500000000	2	Kim et al., 2007
					5000000000	4	Kim et al., 2007
738-70-5	Trimethoprim	Algae	Aquatic community	PNEC	100		Park and Choi, 2008
			<i>Microcystis aeruginosa</i>	EC50	6900000	1	Grinten et al., 2010
					112000000	7	Holten Lützhof et al., 1999
					120000000		Halling-Sorensen et al., 2000
			<i>Rhodomonas salina</i>	EC50	16000000		Holten Lützhof et al., 1999
			<i>Selenastrum capricornutum</i>	NOEC	16000000	3	Yang et al., 2008
					25500000		Eguchi et al., 2004
				LOEC	40000000	3	Yang et al., 2008
				EC50	9000000	1	Grinten et al., 2010
					40000000	3	Yang et al., 2008
					80300000		Eguchi et al., 2004
					80300000		Yang et al., 2008
					100000000		Halling-Sorensen et al., 2000
					130000000		Holten Lützhof et al., 1999
					130000000		Yang et al., 2008
		Amphibia	<i>Xenopus laevis</i>	NOEC	100000000	4	Richards and Cole, 2006
				LOEC	1000000000	4	Richards and Cole, 2006
		Bacteria	<i>Bacillus cereus</i>	EC50	350000	1	Grinten et al., 2010
			<i>Bacillus pumilus</i>	EC50	28000	1	Grinten et al., 2010
			<i>Micrococcus luteus</i>	EC50	350000	1	Grinten et al., 2010
			<i>Vibrio fischeri</i>	EC50	280000	0.02	Grinten et al., 2010
					176700000	0.01	Kim et al., 2007
			<i>Yersinia ruckeri</i>	EC50	350000	1	Grinten et al., 2010
					17800000		Halling-Sorensen et al., 2000
		Bacteria		EC50	123000000		Park and Choi, 2008
		Crustacea	<i>Ceriodaphnia dubia</i>	NOEC	6000000	21	Park and Choi, 2008
			<i>Daphnia magna</i>	LOEC	20000000	21	Park and Choi, 2008
				EC50	92000000	2	Park and Choi, 2008
					120700000	4	Kim et al., 2007
					123000000	2	Halling-Sorensen et al., 2000
					149000000	2	Liguoro et al., 2009
					149000000	2	Yang et al., 2011
					155600000	1	Park and Choi, 2008
					167400000	2	Kim et al., 2007
			<i>Moina macrocopa</i>	EC50	54800000	2	Park and Choi, 2008
					144800000	1	Park and Choi, 2008
		Fish	<i>Brachydanio rerio</i>	NOEC	100000000	3	Halling-Sorensen et al., 2000
			<i>Oryzias latipes</i>	LC50	100000000	2	Kim et al., 2007
					100000000	4	Kim et al., 2007
					100000000	4	Quinn et al., 2007
					100000000	4	Quinn et al., 2007
					100000000	4	Quinn et al., 2007
			Aquatic community	PNEC	60000		Park and Choi, 2008
					120700	4	Kim et al., 2007
					149000	2	Yang et al., 2011
					178000		Halling-Sorensen et al., 2000

## I.4 TETRACYCLINES

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference			
56-75-7	Chloramphenicol	Algae	<i>Tetraselmis suecica</i>	NOEC	2500000	25000	1	Seoane et al., 2014		
				LOEC	2500000	25000	1	Seoane et al., 2014		
				EC50	11160000	1116000	4	Seoane et al., 2014		
57-62-5	Chlortetracycline	Bacteria	<i>Vibrio fisheri</i>	EC50	64300	6430	1	Backhaus and Grimme, 1999		
		Algae	<i>Microcystis aeruginosa</i>	EC50	50000	200000	7	Halling-Sorensen, 2000		
				NOEC	500000	200000	3	Yang et al., 2008		
				LOEC	1000000	400000	3	Yang et al., 2008		
				EC50	1800000	720000	3	Yang et al., 2008		
					3100000	1240000	3	Halling-Sorensen, 2000		
			3100000	1240000		Yang et al., 2008				
		Amphibia	<i>Xenopus laevis</i>	NOEC	100000000	40000000	4	Richards and Cole, 2006		
				LOEC	100000000	40000000	4	Richards and Cole, 2006		
		Bacteria	<i>Vibrio fisheri</i>	EC50	13000000	5200000	0.01	Park and Choi, 2008		
					20000000	8000000	0.003	Park and Choi, 2008		
		Crustacea	<i>Daphnia magna</i>	EC50	225000000	90000000	2	Park and Choi, 2008		
					380100000	152040000	1	Park and Choi, 2008		
				EC50	272000000	108800000	2	Park and Choi, 2008		
		Crustacea	<i>Moina macrocopa</i>		515000000	206000000	1	Park and Choi, 2008		
				EC50	789000000	315600000	4	Park and Choi, 2008		
		Fish	<i>Oryzias latipes</i>	LC50	88400000	35360000	2	Park and Choi, 2008		
					100000	400000	7	Brain et al., 2004		
		Plants	<i>Lemna gibba</i>	LOEC	300000	1200000	7	Brain et al., 2004		
					300000	1200000	7	Brain et al., 2004		
	300000			1200000	7	Brain et al., 2004				
	300000			1200000	7	Brain et al., 2004				
	300000			1200000	7	Brain et al., 2004				
EC50	2190000			876000	7	Brain et al., 2004				
	3180000			1272000	7	Brain et al., 2004				
	6300000			2520000	7	Brain et al., 2004				
	6500000			2600000	7	Brain et al., 2004				
	16200000			6480000	7	Brain et al., 2004				
564-25-0	Doxycycline	Plants	<i>Lemna gibba</i>	Aquatic community	PNEC	50	200		Park and Choi, 2008	
				LOEC	300000	1200000	7	Brain et al., 2004		
					300000	1200000	7	Brain et al., 2004		
					300000	1200000	7	Brain et al., 2004		
					1000000	4000000	7	Brain et al., 2004		
				EC50	316000	1264000	7	Brain et al., 2004		
					4730000	1892000	7	Brain et al., 2004		
					1844000	7376000	7	Brain et al., 2004		
					2616000	10464000	7	Brain et al., 2004		
				79-57-2	Oxytetracycline	Algae	<i>Chlorella vulgaris</i>	NOEC	3580000	358000
EC50	6400000	640000	2					Kolodziejska et al., 2013		
	6400000	640000	2					Pro et al., 2003		
	7050000	705000	3					Kolodziejska et al., 2013		
	7050000	705000						Eguchi et al., 2004		
EC50	6430000	6430000	4					Seoane et al., 2014		
EC50	207000	207000	7					Holten Lützhof et al., 1999		
	5400000	540000	1					Grinten et al., 2010		
EC50	1730000	1730000	4					Seoane et al., 2014		
EC50	1600000	160000						Holten Lützhof et al., 1999		
EC50	40400000	4040000	1					Kolodziejska et al., 2013		
NOEC	183000	183000						Eguchi et al., 2004		
EC50	170000	17000	3					Isidori et al., 2005		
	170000	17000	3					Kolodziejska et al., 2013		
	342000	34200	3					Kolodziejska et al., 2013		
	342000	34200						Eguchi et al., 2004		
	470000	47000	2					Kolodziejska et al., 2013		
	600000	60000	1					Grinten et al., 2010		
	3100000	3100000	4					Kolodziejska et al., 2013		
	4500000	450000	3					Kolodziejska et al., 2013		
	4500000	450000						Holten Lützhof et al., 1999		
<i>Tetraselmis chuii</i>	NOEC	3600000	360000					3	Ferreira et al., 2007	
	LOEC	5300000	530000					3	Ferreira et al., 2007	
	EC50	1180000	1180000					4	Ferreira et al., 2007	
		1180000	1180000					4	Kolodziejska et al., 2013	
		13160000	1316000					3	Ferreira et al., 2007	
		13160000	1316000					3	Kolodziejska et al., 2013	
	<i>Tetraselmis suecica</i>	NOEC	7500000					750000	1	Seoane et al., 2014
		LOEC	5000000					500000	1	Seoane et al., 2014
			5000000					500000	1	Seoane et al., 2014
		EC50	17250000					17250000	4	Seoane et al., 2014
EC50		81000	81000					1	Grinten et al., 2010	
EC50		150000	150000					1	Grinten et al., 2010	
EC50		150000	150000					1	Grinten et al., 2010	
EC50		100000	10000					0.02	Grinten et al., 2010	
		21000000	2100000					0.02	Kolodziejska et al., 2013	
		64500000	6450000					0.02	Isidori et al., 2005	
	64500000	6450000	0.02					Kolodziejska et al., 2013		
	66000000	6600000	0.01					Kolodziejska et al., 2013		
	87000000	8700000	0.01					Kolodziejska et al., 2013		
	87000000	8700000	0.01	Park and Choi, 2008						
	108000000	10800000	0.02	Kolodziejska et al., 2013						
	132300000	13230000	0.02	Kolodziejska et al., 2013						
	235400000	23540000	0.003	Kolodziejska et al., 2013						
	235400000	23540000	0.003	Park and Choi, 2008						
<i>Yersinia ruckeri</i>	EC50	150000	150000	1	Grinten et al., 2010					

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference								
60-54-8	Tetracycline	Crustacea	<i>Artemia parthenogenetica</i>	NOEC	637000000	637000000	2	Ferreira et al., 2007							
			LOEC	828000000	828000000	2	Ferreira et al., 2007								
					EC50	805990000	805990000	2	Kolodziejska et al., 2013						
					870470000	870470000	1	Kolodziejska et al., 2013							
					LC50	805990000	805990000	2	Ferreira et al., 2007						
					870470000	870470000	1	Ferreira et al., 2007							
						EC50	180000	180000	7	Isidori et al., 2005					
							180000	180000	7	Kolodziejska et al., 2013					
							18650000	18650000	2	Isidori et al., 2005					
							18650000	18650000	2	Kolodziejska et al., 2013					
						LOEC	100000000	100000000	2	Kolodziejska et al., 2013					
							100000000	100000000	2	Wollenberger et al., 2000					
							22640000	22640000	1	Isidori et al., 2005					
							22640000	22640000	1	Kolodziejska et al., 2013					
							46200000	46200000	21	Kolodziejska et al., 2013					
							46200000	46200000	21	Wollenberger et al., 2000					
							86000000	86000000	2	Kolodziejska et al., 2013					
							114000000	114000000	2	Kolodziejska et al., 2013					
							621200000	621200000	2	Kolodziejska et al., 2013					
							621200000	621200000	2	Park and Choi, 2008					
						EC50	831600000	831600000	1	Kolodziejska et al., 2013					
							831600000	831600000	1	Park and Choi, 2008					
							300000000	300000000	2	Rico et al., 2014b					
							126700000	126700000	2	Kolodziejska et al., 2013					
							126700000	126700000	2	Park and Choi, 2008					
							137100000	137100000	1	Kolodziejska et al., 2013					
							137100000	137100000	1	Park and Choi, 2008					
											LC50	250000000	250000000	1	Isidori et al., 2005
												1000000000	1000000000	4	Isidori et al., 2005
												125000000	125000000	2	Carraschi et al., 2011
		110100000	110100000	4	Park and Choi, 2008										
						LC50	215400000	215400000	2	Park and Choi, 2008					
							76000000	76000000	2	Carraschi et al., 2011					
							200000000	200000000	1	Carraschi et al., 2011					
							647000000	647000000	2	Rico et al., 2014b					
						LC50	18700000	18700000	2	Isidori et al., 2005					
							34210000	34210000	1	Isidori et al., 2005					
						EC50	958000000	958000000	2	Rico et al., 2014b					
							791000000	791000000	2	Rico et al., 2014b					
						LOEC	100000	100000	7	Brain et al., 2004					
							1000000	1000000	7	Brain et al., 2004					
							1000000	1000000	7	Brain et al., 2004					
							1000000	1000000	7	Brain et al., 2004					
							1010000	1010000	7	Brain et al., 2004					
							1010000	1010000	7	Kolodziejska et al., 2013					
							1152000	1152000	7	Brain et al., 2004					
							1179000	1179000	7	Brain et al., 2004					
							1401000	1401000	7	Brain et al., 2004					
							1401000	1401000	7	Brain et al., 2004					
						EC50	2100000	2100000	7	Kolodziejska et al., 2013					
3260000	3260000						7	Kolodziejska et al., 2013							
49200000	49200000						7	Kolodziejska et al., 2013							
49200000	49200000						7	Pro et al., 2003							
				NOEC	50000000	50000000	4	Quinn et al., 2007							
					100000000	100000000	4	Quinn et al., 2007							
					40130000	40130000	4	Quinn et al., 2007							
					100000000	100000000	4	Quinn et al., 2007							
				EC50	217000000	217000000	2	Rico et al., 2014b							
					170	170		Park and Choi, 2008							
					230	230		Jones et al., 2002							
					2180	2180		Rico et al., 2014b							
					4500	4500		Jones et al., 2002							
					117000	117000		Rico et al., 2014b							
				EC50	7000000	7000000		Carraschi et al., 2011							
					90000	90000	7	Halling-Sorensen, 2000							
					500000	500000	3	Yang et al., 2008							
					1000000	1000000	3	Yang et al., 2008							
					1000000	1000000	3	Yang et al., 2008							
					2200000	2200000	3	Halling-Sorensen, 2000							
					2200000	2200000	3	Yang et al., 2008							
									EC50	25100	25100	1	Backhaus and Grimme, 1999		
										340000000	340000000	2	Wollenberger et al., 2000		
									LOEC	448000000	448000000	21	Wollenberger et al., 2000		
100000	100000	7	Brain et al., 2004												
3000000	3000000	7	Brain et al., 2004												
1000000	1000000	7	Brain et al., 2004												
1000000	1000000	7	Brain et al., 2004												
723000	723000	7	Brain et al., 2004												
1114000	1114000	7	Brain et al., 2004												
2592000	2592000	7	Brain et al., 2004												
4569000	4569000	7	Brain et al., 2004												
1060000	1060000	7	Pomati et al., 2004												
				EC50	198000000	495000		Eguchi et al., 2004							
					522000000	1305000		Eguchi et al., 2004							
					4060000	10150		Eguchi et al., 2004							
					8860000	22150		Eguchi et al., 2004							

## I.5 QUINOLONES

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference				
8572-133-1	Ciprofloxacin	Algae	<i>Anabaena flosaquae</i>	EC50	10200	5100	Ebert et al., 2011				
					363000	18150	Ebert et al., 2011				
					23000000	11500000	3	Andrieu et al., 2005			
				<i>Chlorella vulgaris</i>	EC50	23000000	11500000	3	Andrieu et al., 2005		
				<i>Desmodesmus subspicatus</i>	NOEC	8042000	4021000		Ebert et al., 2011		
				<i>Microcystis aeruginosa</i>	EC50	5000	2500		Halling-Sorensen et al., 2000		
					17000	85000	5	Robinson et al., 2005			
					5000000	2500000	3	Yang et al., 2008			
				<i>Selenastrum capricornutum</i>	NOEC	5000000	2500000	3	Yang et al., 2008		
					LOEC	5000000	2500000	3	Yang et al., 2008		
					EC50	1100000	550000		Yang et al., 2008		
						2970000	1485000		Halling-Sorensen et al., 2000		
						2970000	1485000		Yang et al., 2008		
					6700000	3350000	3	Yang et al., 2008			
					18700000	9350000	3	Robinson et al., 2005			
			Amphibia	<i>Xenopus laevis</i>	NOEC	100000000	50000000	4	Richards and Cole, 2006		
			LOEC		100000000	50000000	4	Richards and Cole, 2006			
			Bacteria	<i>Bacteria</i>	EC50	610000	305000		Halling-Sorensen et al., 2000		
			Crustacea	<i>Daphnia magna</i>	NOEC	60000000	30000000	2	Halling-Sorensen et al., 2000		
					EC50	1200000	600000		Andrieu et al., 2005		
				<i>Moina macrocopa</i>	EC50	71000000	35500000	2	Andrieu et al., 2005		
			Fish	<i>Brachydanio rerio</i>	NOEC	100000000	50000000	3	Halling-Sorensen et al., 2000		
				<i>Gambusia holbrooki</i>	EC50	60000000	30000000		Andrieu et al., 2005		
			Plants	<i>Lemna gibba</i>	LOEC	300000	1500000	7	Brain et al., 2004		
						300000	1500000	7	Brain et al., 2004		
						1000000	5000000	7	Brain et al., 2004		
						1000000	5000000	7	Brain et al., 2004		
						1000000	5000000	7	Brain et al., 2004		
						1000000	5000000	7	Brain et al., 2004		
						EC50	1000000	5000000	7	Brain et al., 2004	
							697000	3485000	7	Brain et al., 2004	
							698000	3490000	7	Brain et al., 2004	
							992000	4960000	7	Brain et al., 2004	
						1279000	6395000	7	Brain et al., 2004		
						1762000	8810000	7	Brain et al., 2004		
					<i>Lemna minor</i>	NOEC	10000	5000		Ebert et al., 2011	
						EC50	62500	31250		Ebert et al., 2011	
						203000	1015000	7	Robinson et al., 2005		
					<i>Myriophyllum spicatum</i>	NOEC	980000	490000		Ebert et al., 2011	
					Aquatic community	PNEC	50.00	250		Halling-Sorensen et al., 2000	
							1200	6000		Andrieu et al., 2005	
						60000	300000		Andrieu et al., 2005		
		93106-60-6		Enrofloxacin	Algae	<i>Anabaena flosaquae</i>	NOEC	19100	7640	Ebert et al., 2011	
							173000	69200	Ebert et al., 2011		
							465000	186000	Ebert et al., 2011		
						<i>Chlorella vulgaris</i>	EC50	11000000	44400000	3	Andrieu et al., 2005
						<i>Desmodesmus subspicatus</i>	NOEC	500000	200000		Ebert et al., 2011
						1140000	456000		Ebert et al., 2011		
			EC50			28369	11348		Ebert et al., 2011		
						5568000	2227200		Ebert et al., 2011		
			<i>Microcystis aeruginosa</i>			EC50	49000	196000	5	Robinson et al., 2005	
			<i>Selenastrum capricornutum</i>			EC50	3100000	1240000	3	Robinson et al., 2005	
	Archea		<i>Archaeal amoA gene</i>			NOEC	1000	4000	7	Rico et al., 2014	
	Bacteria		<i>Bacterial amoA gene</i>			NOEC	10000	40000	7	Rico et al., 2014	
			<i>Bacterial OTUs</i>			NOEC	100000	400000	7	Rico et al., 2014	
			<i>nifH gene</i>		NOEC	100000	400000	7	Rico et al., 2014		
			<i>Vibrio fischeri</i>		EC50	326800000	130720000	0.01	Park and Choi, 2008		
						425000000	170000000	0.003	Park and Choi, 2008		
	Crustacea		<i>Alonella sp.</i>		NOEC	100000	400000	7	Rico et al., 2014		
					NOEC	1000	4000	7	Rico et al., 2014		
			<i>Ceriodaphnia reticulata</i>		NOEC	5000000	20000000	21	Park and Choi, 2008		
					LOEC	15000000	60000000	21	Park and Choi, 2008		
			<i>Daphnia magna</i>		EC50	11470000	45880000	21	Park and Choi, 2008		
						53300000	21320000		Andrieu et al., 2005		
						56700000	22680000	2	Park and Choi, 2008		
						131700000	52680000	1	Park and Choi, 2008		
					<i>Macrobrachium lancesteri</i>	LC50	202000000	80800000	2	Rico et al., 2014b	
					<i>Moina macrocopa</i>	EC50	69000000	27600000	2	Andrieu et al., 2005	
					200000000	80000000	2	Park and Choi, 2008			
					285700000	114280000	1	Park and Choi, 2008			
	Fish		<i>Lepomis macrochirus</i>		EC50	79500000	31800000		Andrieu et al., 2005		
			<i>Oryzias latipes</i>		LC50	100000000	40000000	2	Park and Choi, 2008		
						100000000	40000000	4	Park and Choi, 2008		
	Insecta		<i>Micronectinae sp.</i>		LC50	408000000	163200000	2	Rico et al., 2014b		
	Mollusca		<i>Melanooides tuberculata</i>		EC50	520000000	208000000	2	Rico et al., 2014b		
			<i>Physa acuta</i>		EC50	281000000	112400000	2	Rico et al., 2014b		
	Plants		<i>Lemna minor</i>		NOEC	30000	12000		Ebert et al., 2011		
					EC50	107000	42800		Ebert et al., 2011		
					114000	456000	7	Robinson et al., 2005			
			<i>Myriophyllum spicatum</i>		NOEC	11650000	4660000		Ebert et al., 2011		
	Worm		<i>Limnodrilus hoffmeisteri</i>		EC50	360000000	144000000	2	Rico et al., 2014b		
			Aquatic community		PNEC	49.00	196		Park and Choi, 2008		
						490	1960		Rico et al., 2014b		

CAS	Compound	Organism	Measured endpoint	Concentration (ng/L)	Final TEQs (ng/L)	Exposure time (days)	Reference			
93106-60-6	Enrofloxacin	Aquatic community	PNEC	53300	213200		Andrieu et al., 2005			
				57500	230000		Rico et al., 2014b			
				79500	318000		Andrieu et al., 2005			
42835-25-6	Flumequine	Algae	<i>Microcystis aeruginosa</i>	EC50	159000	159000	7	Holten Lützhof et al., 1999		
					1960000	1960000	5	Robinson et al., 2005		
					8800000	880000	1	Grinten et al., 2010		
					<i>Rhodomonas salina</i>	EC50	18000000	18000000		Holten Lützhof et al., 1999
					<i>Selenastrum capricornutum</i>	EC50	5000000	500000	3	Robinson et al., 2005
							5000000	5000000		Holten Lützhof et al., 1999
							16000000	1600000	1	Grinten et al., 2010
			Bacteria	<i>Bacillus cereus</i>	EC50	1200000	1200000	1	Grinten et al., 2010	
				<i>Bacillus pumilus</i>	EC50	1200000	1200000	1	Grinten et al., 2010	
				<i>Micrococcus luteus</i>	EC50	1200000	1200000	1	Grinten et al., 2010	
				<i>Vibrio fisheri</i>	EC50	800000	80000	0.02	Grinten et al., 2010	
				<i>Yersinia ruckeri</i>	EC50	200000	200000	1	Grinten et al., 2010	
			Crustacea	<i>Artemia franciscana</i>	LC50	307700000	307700000	2	Ferreira et al., 2007	
				<i>Artemia nauplii</i>	EC50	96350000	9635000	3	Migliore et al., 1996	
						307700000	307700000	2	Migliore et al., 1996	
				476800000	476800000	1	Migliore et al., 1996			
70458-96-7	Norfloxacin	Plants	<i>Lemna minor</i>	EC50	2470000	2470000	7	Robinson et al., 2005		
		Algae	<i>Chlorella vulgaris</i>	NOEC	4020000	804000		Eguchi et al., 2004		
				EC50	10400000	2080000		Eguchi et al., 2004		
					<i>Scenedesmus obliquus</i>	EC50	38590000	7718000	2	Nie et al., 2009
					<i>Selenastrum capricornutum</i>	NOEC	2000000	400000	3	Yang et al., 2008
							4010000	802000		Eguchi et al., 2004
						LOEC	16000000	3200000	3	Yang et al., 2008
						EC50	1800000	360000	3	Yang et al., 2008
							16600000	3320000		Eguchi et al., 2004
							16600000	3320000		Yang et al., 2008
			Bacteria	<i>Vibrio fisheri</i>	EC50	11500	23000	1	Backhaus and Grimme, 1999	
						23300	46600	1	Backhaus and Grimme, 1999	
			Plants	<i>Lemna gibba</i>	LOEC	1000000	2000000	7	Brain et al., 2004	
						1000000	2000000	7	Brain et al., 2004	
						1000000	2000000	7	Brain et al., 2004	
				1000000	2000000	7	Brain et al., 2004			
				1000000	2000000	7	Brain et al., 2004			
			EC50	913000	1826000	7	Brain et al., 2004			
				1049000	2098000	7	Brain et al., 2004			
				1072000	2144000	7	Brain et al., 2004			
				1130000	2260000	7	Brain et al., 2004			
				1146000	2292000	7	Brain et al., 2004			
14698-29-4	Oxolinic acid	Algae	Aquatic community	PNEC	20000	40000	3	Yang et al., 2011		
			<i>Microcystis aeruginosa</i>	EC50	180000	360000	7	Holten Lützhof et al., 1999		
			<i>Rhodomonas salina</i>	EC50	10000000	20000000		Holten Lützhof et al., 1999		
			<i>Selenastrum capricornutum</i>	EC50	16000000	32000000		Holten Lützhof et al., 1999		
		Crustacea	<i>Daphnia magna</i>	EC50	4600000	920000	2	Wollenberger et al., 2000		
98105-99-8	Sarafloxacin	Algae	<i>Microcystis aeruginosa</i>	EC50	15000	30000	7	Holten Lützhof et al., 1999		
			<i>Rhodomonas salina</i>	EC50	24000000	48000000		Holten Lützhof et al., 1999		
			<i>Selenastrum capricornutum</i>	EC50	16000000	32000000		Holten Lützhof et al., 1999		
3380-34-5	Triclosan	Algae	<i>Selenastrum capricornutum</i>	NOEC	200	2.00	3	Yang et al., 2008		
				LOEC	400	4.00	3	Yang et al., 2008		
				EC50	530	5.30	3	Yang et al., 2008		
				1400	14.00		Yang et al., 2008			
		Crustacea	<i>Daphnia magna</i>	EC50	390000	3900	2	Ishibashi et al., 2004		
		Fish	<i>Lepomis macrochirus</i>	LC50	370000	3700	4	Ishibashi et al., 2004		
			<i>Oryzias latipes</i>	LC50	399000	39900	14	Ishibashi et al., 2004		
					602000	6020	4	Ishibashi et al., 2004		
			<i>Pimephales promelas</i>	LC50	260000	2600	4	Ishibashi et al., 2004		

# APPENDIX SI-III

## Lowest toxicity values found for TWO REP groups

REP1: >0.1

Activity	Bioassay	Endpoint	CAS	Compound	Organism	Original data	TEQs	Reference		
Estrogenic activity	ERA CALUX	PNEC	57-63-6	17a-Ethinyl estradiol		0.04	ng/L	0.05	ng EEQ/L	James et al., 2014
		NOEC	50-28-2	17a-Ethinyl estradiol	<i>Rutilus rutilus</i> (Roach)	0.04	ng/L	0.06	ng EEQ/L	Hogan et al., 2008
		LOEC	57-63-6	17a-Ethinyl estradiol	<i>Danio rerio</i> (Zebrafish)	0.5	ng/L	0.02	ng EEQ/L	Colman et al., 2009
		EC50	57-63-6	17a-Ethinyl estradiol	<i>Danio rerio</i> (Zebrafish)	1.1	ng/L	0.17	ng EEQ/L	Wenzel et al., 2001
		LC50	57-63-6	17a-Ethinyl estradiol	<i>Danio rerio</i> (Zebrafish)	100	ng/L	1.6	ng EEQ/L	Wenzel et al., 2001
Anti-androgenic	antiAR CALUX	PNEC	50-32-8	Benzo [a] pyrene		0.00017	µg/L	0.00017	µg FEQ/L	OSPAR Agreement, 2014-05
		NOEC	115-29-7	Endosulfan	<i>Mesocyclops longisetus</i> (Copepod)	0.004	µg/L	0.001	µg FEQ/L	Gutierrez et al., 2013
		LOEC	115-29-7	Endosulfan	<i>Mesocyclops longisetus</i> (Copepod)	0.004	µg/L	0.0003	µg FEQ/L	Gutierrez et al., 2013
		EC50	298-00-0	Parathion-methyl	<i>Daphnia magna</i> (Water flea)	0.14	µg/L	0.0014	µg FEQ/L	Nendza and Wenzel, 2006
		LC50	115-29-7	Endosulfan	<i>Mesocyclops longisetus</i> (Copepod)	0.016	µg/L	0.0005	µg FEQ/L	Gutierrez et al., 2013
Dioxin and dioxin-like	DR CALUX	PNEC					pg/L		pg TEQ/L	
		NOEC	1746-01-6	2,3,7,8-TCDD	<i>Oryzias latipes</i> (Japanese medaka)	3400	pg/L	340	pg TEQ/L	Kim and Cooper, 1998
		LOEC	1746-01-6	2,3,7,8-TCDD	<i>Gobiocypris rarus</i> (Rare minnow)	2	pg/L	0.4	pg TEQ/L	Wu et al., 2001
		EC50	1746-01-6	2,3,7,8-TCDD	<i>Oryzias latipes</i> (Japanese medaka)	1200	pg/L	12	pg TEQ/L	Chen and Cooper, 1999
		LC50	1746-01-6	2,3,7,8-TCDD	<i>Oryzias latipes</i> (Japanese medaka)	8100	pg/L	8.1	pg TEQ/L	Kim and Cooper, 1999
Glucocorticoid	GR CALUX	PNEC	50-24-8	Prednisolone		139000	ng/L	27800	ng DEQ/L	Escher et al., 2011
		NOEC	50-02-2	Dexamethasone	<i>Danio rerio</i> (Zebrafish)	39246	ng/L	3925	ng DEQ/L	Gustafson et al., 2013
		LOEC	50-02-2	Dexamethasone	<i>Pimephales promelas</i> (Fathead Minnow)	100	ng/L	20	ng DEQ/L	Lalone et al., 2012
		EC50	50-24-8	Prednisolone	<i>Ceriodaphnia dubia</i> (Water Flea)	230000	ng/L	4600	ng DEQ/L	DellaGreca et al., 2004
		LC50	50-02-2	Dexamethasone	<i>Pimephales promelas</i> (Fathead Minnow)	254000	ng/L	2540	ng DEQ/L	Overturf et al., 2012
PPARγ receptor	PPARγ CALUX	PNEC					ng/L		ng REQ/L	
		NOEC	688-73-3	Tributyltin hydride	<i>Lymnaea stagnalis</i> (Great Pond Snail)	19	ng/L	15	ng REQ/L	Giusti et al., 2013
		LOEC	302-79-4	Retinoic acid	<i>Danio rerio</i> (Zebrafish)	300	ng/L	1.9	ng REQ/L	Teixido et al., 2013
		EC50	302-79-4	Retinoic acid	<i>Danio rerio</i> (Zebrafish)	424	ng/L	1.3	ng REQ/L	Selderslaghs et al., 2012
		LC50	302-79-4	Retinoic acid	<i>Danio rerio</i> (Zebrafish)	15322	ng/L	4.8	ng REQ/L	Teixido et al., 2013
Toxic PAHs	PAH CALUX	PNEC	50-32-8	Benzo [a] pyrene		0.17	ng/L	0.17	ng BEQ/L	OSPAR Agreement, 2014-05
		NOEC	57465-28-8	PCB126	<i>Oryzias latipes</i> (Japanese medaka)	150	ng/L	47	ng BEQ/L	Kim and Cooper, 1998
		LOEC	50-32-8	Benzo [a] pyrene	<i>Daphnia magna</i> (Water flea)	20	ng/L	0.4	ng BEQ/L	Ha and Choi, 2009
		EC50	57465-28-8	PCB126	<i>Oryzias latipes</i> (Japanese medaka)	170	ng/L	5.4	ng BEQ/L	Kim and Cooper, 1999
		LC50	57465-28-8	PCB126	<i>Oryzias latipes</i> (Japanese medaka)	250	ng/L	0.8	ng BEQ/L	Kim and Cooper, 1999
Oxidative stress	Nrf2 CALUX	PNEC	10605-21-7	Carbendazim		0.57	µg/L	0.02	µg CEQ/L	Oekotoxzentrum, Centre Ecotox
		NOEC	302-79-4	Retinoic acid	<i>Danio rerio</i> (Zebrafish)	0.6	µg/L	0.01	µg CEQ/L	Teixido et al., 2013
		LOEC	50471-44-8	Vinclozolin	<i>Marisa cornuarietis</i> (Giant ramshorn snail)	0.03	µg/L	0.001	µg CEQ/L	Tillmann et al., 2001
		EC50	302-79-4	Retinoic acid	<i>Danio rerio</i> (Zebrafish)	0.42	µg/L	0.0007	µg CEQ/L	Selderslaghs et al., 2012
		LC50	10108-64-2	Cadmium chloride	<i>Oncorhynchus mykiss</i> (Rainbow Trout)	0.84	µg/L	0.001	µg CEQ/L	Mebane et al., 2008
Pregnane X receptor	PXR CALUX	PNEC	207-08-9	Benzo[k]fluoranthene		0.17	ng/L	0.03	ng NEO/L	OSPAR Agreement, 2014-05
		NOEC	2921-88-2	Chlorpyrifos-ethyl	<i>Daphnia magna</i> (Water flea)	24	ng/L	0.48	ng NEO/L	U.S. EPA, 2013
		LOEC	2921-88-2	Chlorpyrifos-ethyl	<i>Daphnia magna</i> (Water flea)	1	ng/L	0.004	ng NEO/L	Ha and Choi, 2009
		EC50	2921-88-2	Chlorpyrifos-ethyl	<i>Daphnia magna</i> (Water flea)	32	ng/L	0.07	ng NEO/L	Antunes et al., 2010
		LC50	2921-88-2	Chlorpyrifos-ethyl	<i>Hyalella curvispina</i> (Scud)	60	ng/L	0.01	ng NEO/L	Mugni et al., 2012
Aminoglycosides		PNEC	1404-04-2	Neomycin	<i>Daphnia magna</i> (Water flea)	300	ng/L	300	ng NEO/L	Park and Choi, 2008
		NOEC	128-46-1	Dihydrostreptomycin	<i>Selenastrum capricornutum</i> (Green algae)	39000	ng/L	7800	ng NEO/L	Eguchi et al., 2004
		LOEC	1404-04-2	Neomycin	<i>Daphnia magna</i> (Water flea)	100000	ng/L	20000	ng NEO/L	Park and Choi, 2008
		EC50	128-46-1	Dihydrostreptomycin	<i>Selenastrum capricornutum</i> (Green algae)	107000	ng/L	2140	ng NEO/L	Eguchi et al., 2004
		LC50	1404-04-2	Neomycin	<i>Oryzias latipes</i> (Japanese medaka)	8080000	ng/L	80800	ng NEO/L	Park and Choi, 2008
Macrolides & β-Lactam		PNEC	26787-78-0	Amoxicillin	<i>Microcystis aeruginosa</i>	3.7	ng/L	2.22	ng PEQ/L	Jones et al., 2002
		NOEC	114-07-8	Erythromycin	<i>Selenastrum capricornutum</i> (Green algae)	10300	ng/L	309	ng PEQ/L	Eguchi et al., 2004
		LOEC	26787-78-0	Amoxicillin	<i>Lemna gibba</i> (Inflated Duckweed)	1000000	ng/L	12000	ng PEQ/L	Park and Choi, 2008
		EC50	26787-78-0	Amoxicillin	<i>Microcystis aeruginosa</i>	3700	ng/L	22.2	ng PEQ/L	Park and Choi, 2008
		LC50	114-07-8	Erythromycin	<i>Brachionus calyciflorus</i> (Rotifer)	940000	ng/L	282	ng PEQ/L	Isidori et al., 2005
Antibiotic activity	Sulphonamides	PNEC	72-14-0	Sulfathiazole	<i>Lemna gibba</i> (Inflated Duckweed)	100	ng/L	25	ng SEQ/L	Park and Choi, 2008
		NOEC	723-46-6	Sulfamethoxazole	<i>Danio rerio</i> (Zebrafish)	1000	ng/L	100	ng SEQ/L	Lin et al., Manuscript Draft
		LOEC	68-35-9	Sulfadiazine	<i>Danio rerio</i> (Zebrafish)	1000	ng/L	10	ng SEQ/L	Lin et al., Manuscript Draft
		EC50	723-46-6	Sulfamethoxazole	<i>Synechococcus leopoliensis</i>	26800	ng/L	2680	ng SEQ/L	Ferrari et al., 2004
		LC50	723-46-6	Sulfamethoxazole	<i>Brachionus calyciflorus</i> (Rotifer)	9630000	ng/L	9630	ng SEQ/L	Isidori et al., 2005
Tetracyclines		PNEC	79-57-2	Oxytetracycline	<i>Selenastrum capricornutum</i> (Green algae)	170	ng/L	170	ng OEQ/L	Park and Choi, 2008
		NOEC	79-57-2	Oxytetracycline	<i>Selenastrum capricornutum</i> (Green algae)	183000	ng/L	18300	ng OEQ/L	Eguchi et al., 2004
		LOEC	56-75-7	Chloramphenicol	<i>Tetraselmis suecica</i>	2500000	ng/L	5000	ng OEQ/L	Seoane et al., 2014
		EC50	56-75-7	Chloramphenicol	<i>Vibrio fischeri</i>	64300	ng/L	643	ng OEQ/L	Backhaus and Grimme, 1999
		LC50	79-57-2	Oxytetracycline	<i>Brachionus calyciflorus</i> (Rotifer)	1870000	ng/L	1870	ng OEQ/L	Isidori et al., 2005
Quinolones		PNEC	93106-60-6	Enrofloxacin	<i>Microcystis aeruginosa</i>	49	ng/L	196	ng FEQ/L	Park and Choi, 2008
		NOEC	3380-34-5	Tridosan	<i>Selenastrum capricornutum</i> (Green algae)	200	ng/L	2	ng FEQ/L	Yang et al., 2008
		LOEC	3380-34-5	Tridosan	<i>Selenastrum capricornutum</i> (Green algae)	400	ng/L	0.8	ng FEQ/L	Yang et al., 2008
		EC50	3380-34-5	Tridosan	<i>Selenastrum capricornutum</i> (Green algae)	530	ng/L	0.53	ng FEQ/L	Yang et al., 2008
		LC50	3380-34-5	Tridosan	<i>Pimephales promelas</i> (Fathead Minnow)	260000	ng/L	2.6	ng FEQ/L	Ishibashi et al., 2004



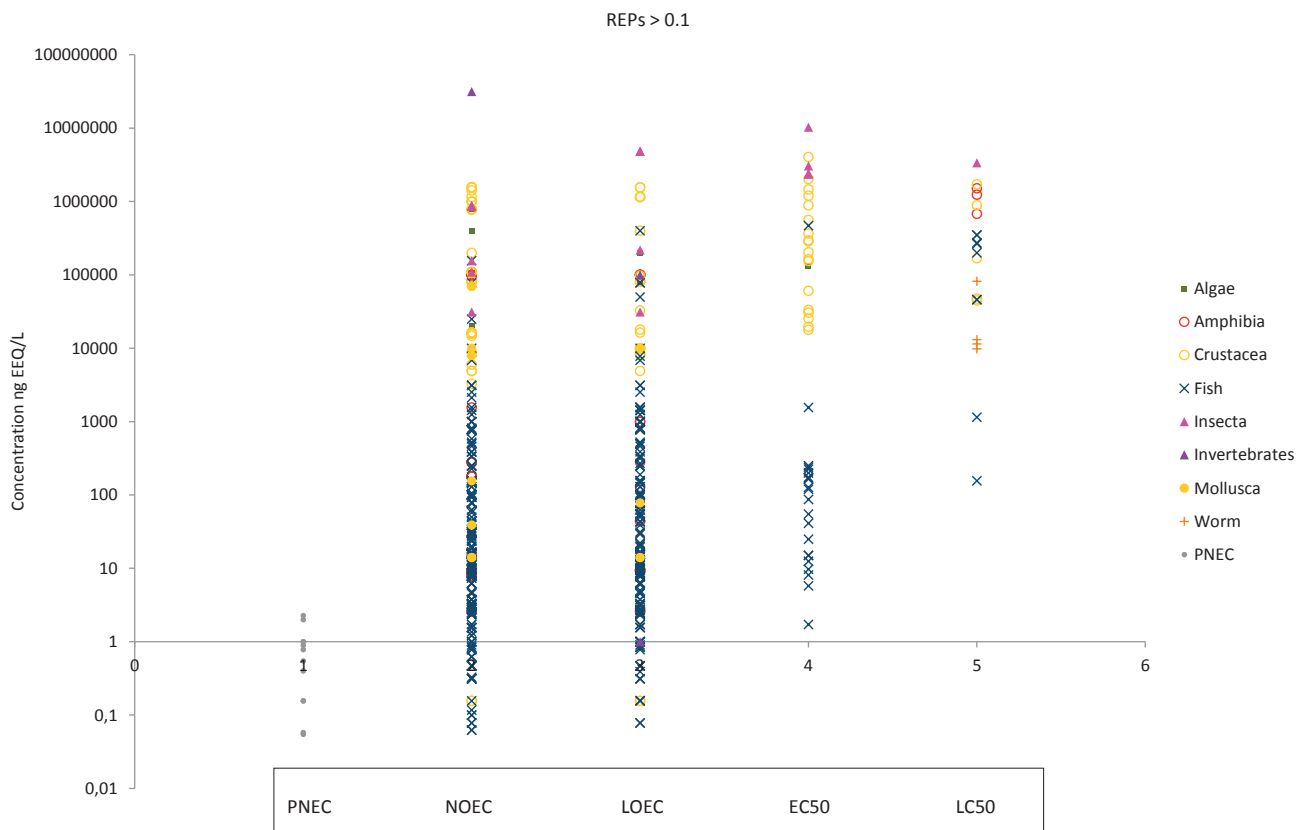
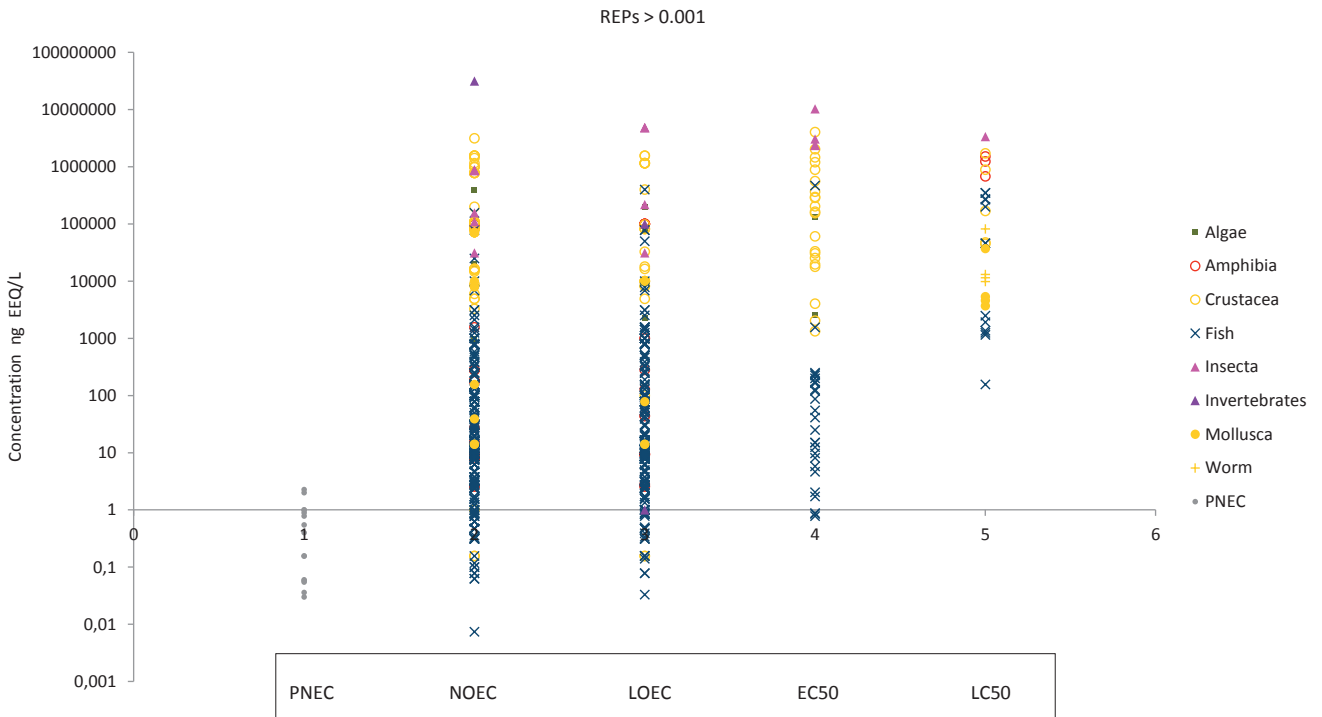
REP2: >0.001

Activity	Bioassay	Endpoint	CAS	Compound	Organism	Original data	TEOs	Reference
Estrogenic activity	ErA CALUX	PNEC	53-16-7	Estrone		3 ng/L	0.03 ng EEQ/L	Johnson et al., 2007
		NOEC	53-16-7	Estrone	<i>Oncorhynchus mykiss</i> (Rainbow Trout)	0.74 ng/L	0.007 ng EEQ/L	Thorpe et al., 2003
		LOEC	53-16-7	Estrone	<i>Oncorhynchus mykiss</i> (Rainbow Trout)	3.3 ng/L	0.007 ng EEQ/L	Thorpe et al., 2003
		EC50	53-16-7	Estrone	<i>Danio rerio</i> (Zebrafish)	78 ng/L	0.08 ng EEQ/L	Holbech et al., 2006
		LC50	57-63-6	17 $\alpha$ -Ethinyl estradiol	<i>Danio rerio</i> (Zebrafish)	100 ng/L	1.6 ng EEQ/L	Wenzel et al., 2001
Anti-androgenic	antiAR CALLUX	PNEC	50-32-8	Benzo [a] pyrene		0.00017 $\mu$ g/L	0.00017 $\mu$ g FEQ/L	OSPAR Agreement, 2014-05
		NOEC	115-29-7	Endosulfan	<i>Mesocyclops longisetus</i> (Copepod)	0.004 $\mu$ g/L	0.001 $\mu$ g FEQ/L	Gutierrez et al., 2013
		LOEC	115-29-7	Endosulfan	<i>Mesocyclops longisetus</i> (Copepod)	0.004 $\mu$ g/L	0.0003 $\mu$ g FEQ/L	Gutierrez et al., 2013
		EC50	10605-21-7	Carbendazim	<i>Daphnia magna</i> (Water Flea)	3.5 $\mu$ g/L	0.00007 $\mu$ g FEQ/L	Ferreira et al., 2008
		LC50	115-29-7	Endosulfan	<i>Mesocyclops longisetus</i> (Copepod)	0.016 $\mu$ g/L	0.00005 $\mu$ g FEQ/L	Gutierrez et al., 2013
Dioxin and dioxin-like	DR CALUX	PNEC	207-08-9	Benzo(k)fluoranthene		170 pg/L	0.85 pg TEQ/L	OSPAR Agreement, 2014-05
		NOEC	1746-01-6	2,3,7,8-TCDD	<i>Oryzias latipes</i> (Japanese medaka)	3400 pg/L	340 pg TEQ/L	Kim and Cooper, 1998
		LOEC	1746-01-6	2,3,7,8-TCDD	<i>Gobiocypris rarus</i> (Rare minnow)	2 pg/L	0.4 pg TEQ/L	Wu et al., 2001
		EC50	1746-01-6	2,3,7,8-TCDD	<i>Oryzias latipes</i> (Japanese medaka)	1200 pg/L	12 pg TEQ/L	Chen and Cooper, 1999
		LC50	1746-01-6	2,3,7,8-TCDD	<i>Oryzias latipes</i> (Japanese medaka)	8100 pg/L	8.1 pg TEQ/L	Kim and Cooper, 1999
Glucocorticoid	GR CALUX	PNEC	50-24-8	Prednisolone		139000 ng/L	27800 ng DEQ/L	Escher et al., 2011
		NOEC	52-39-1	Aldosterone	<i>Xenopus laevis</i> (African Clawed Frog)	36044 ng/L	288 ng DEQ/L	Lorenz et al., 2009
		LOEC	50-02-2	Dexamethasone	<i>Pimephales promelas</i> (Fathead Minnow)	100 ng/L	20 ng DEQ/L	Lalone et al., 2012
		EC50	50-24-8	Prednisolone	<i>Ceriodaphnia dubia</i> (Water Flea)	230000 ng/L	4600 ng DEQ/L	DellaGreca et al., 2004
		LC50	50-02-2	Dexamethasone	<i>Pimephales promelas</i> (Fathead Minnow)	254000 ng/L	2540 ng DEQ/L	Overturf et al., 2012
PPAR $\gamma$ receptor	PPAR $\gamma$ CALUX	PNEC	53-70-3	dibenzo[a,h]anthracene		0.14 ng/L	0.00014 ng REQ/L	OSPAR Agreement, 2014-05
		NOEC	688-73-3	Tributyltin hydride	<i>Lymnaea stagnalis</i> (Great Pond Snail)	19 ng/L	15 ng REQ/L	Giusti et al., 2013
		LOEC	302-79-4	Retinoic acid	<i>Danio rerio</i> (Zebrafish)	300 ng/L	1.9 ng REQ/L	Teixido et al., 2013
		EC50	53-70-3	dibenzo[a,h]anthracene	<i>Daphnia magna</i> (Water Flea)	400 ng/L	0.004 ng REQ/L	Newsted and Giesy, 1987
		LC50	302-79-4	Retinoic acid	<i>Danio rerio</i> (Zebrafish)	15322 ng/L	4.8 ng REQ/L	Teixido et al., 2013
Toxic PAHs	PAH CALUX	PNEC	50-32-8	Benzo[a]pyrene		0.17 ng/L	0.17 ng BEQ/L	OSPAR Agreement, 2014-05
		NOEC	57465-28-8	PCB126	<i>Oryzias latipes</i> (Japanese medaka)	150 ng/L	47 ng BEQ/L	Kim and Cooper, 1998
		LOEC	1746-01-6	2,3,7,8-TCDD	<i>Gobiocypris rarus</i> (Rare minnow)	0.002 ng/L	0.08 ng BEQ/L	Wu et al., 2001
		EC50	1746-01-6	2,3,7,8-TCDD	<i>Oryzias latipes</i> (Japanese medaka)	1.2 ng/L	2.4 ng BEQ/L	Chen and Cooper, 1999
		LC50	57465-28-8	PCB126	<i>Oryzias latipes</i> (Japanese medaka)	250 ng/L	0.8 ng BEQ/L	Kim and Cooper, 1999
Oxidative stress	Nrf2 CALUX	PNEC	50-28-2	17 $\beta$ -estradiol		0.0004 $\mu$ g/L	0.00003 $\mu$ g CEQ/L	Oekotoxzentrum, Centre Ecotox
		NOEC	50-28-2	17 $\beta$ -estradiol	<i>Oryzias latipes</i> (Japanese medaka)	0.001 $\mu$ g/L	0.00006 $\mu$ g CEQ/L	Lee et al., 2012
		LOEC	50-28-2	17 $\beta$ -estradiol	<i>Oryzias latipes</i> (Japanese medaka)	0.01 $\mu$ g/L	0.00001 $\mu$ g CEQ/L	Lee et al., 2012
		EC50	50-28-2	17 $\beta$ -estradiol	<i>Oncorhynchus mykiss</i> (Rainbow Trout)	0.015 $\mu$ g/L	0.00009 $\mu$ g CEQ/L	Thorpe et al., 2000
		LC50	50-28-2	17 $\beta$ -estradiol	<i>Pimephales promelas</i> (Fathead Minnow)	1.2 $\mu$ g/L	0.0007 $\mu$ g CEQ/L	Kramer et al., 1998
Pregnane X receptor	PXR CALUX	PNEC	207-08-9	Benzo(k)fluoranthene		0.17 ng/L	0.03 ng NEQ/L	OSPAR Agreement, 2014-05
		NOEC	2921-88-2	Chlorpyrifos-ethyl	<i>Daphnia magna</i> (Water Flea)	24 ng/L	0.48 ng NEQ/L	U.S. EPA, 2013
		LOEC	2921-88-2	Chlorpyrifos-ethyl	<i>Daphnia magna</i> (Water Flea)	1 ng/L	0.004 ng NEQ/L	Ha and Choi, 2009
		EC50	2921-88-2	Chlorpyrifos-ethyl	<i>Daphnia magna</i> (Water Flea)	32 ng/L	0.07 ng NEQ/L	Antunes et al., 2010
		LC50	2921-88-2	Chlorpyrifos-ethyl	<i>Hyalella curvispina</i> (Scud)	60 ng/L	0.01 ng NEQ/L	Mugni et al., 2012
Aminoglycosides		PNEC	1404-04-2	Neomycin	<i>Daphnia magna</i> (Water Flea)	300 ng/L	300 ng NEQ/L	Park and Choi, 2008
		NOEC	128-46-1	Dihydrostreptomycin	<i>Selenastrum capricornutum</i> (Green algae)	39000 ng/L	7800 ng NEQ/L	Eguchi et al., 2004
		LOEC	1404-04-2	Neomycin	<i>Daphnia magna</i> (Water Flea)	100000 ng/L	20000 ng NEQ/L	Park and Choi, 2008
		EC50	128-46-1	Dihydrostreptomycin	<i>Selenastrum capricornutum</i> (Green algae)	107000 ng/L	2140 ng NEQ/L	Eguchi et al., 2004
		LC50	1404-04-2	Neomycin	<i>Oryzias latipes</i> (Japanese medaka)	80800000 ng/L	80800 ng NEQ/L	Park and Choi, 2008
Macrolides & $\beta$ -Lactam		PNEC	26787-78-0	Amoxicillin	<i>Microcystis aeruginosa</i>	3.7 ng/L	2.22 ng PEQ/L	Jones et al., 2002
		NOEC	1401-69-0	Tylosin	<i>Selenastrum capricornutum</i>	64000 ng/L	240 ng PEQ/L	Yang et al., 2008
		LOEC	1401-69-0	Tylosin	<i>Selenastrum capricornutum</i>	64000 ng/L	240 ng PEQ/L	Yang et al., 2008
		EC50	55297-95-5	Tiamulin	<i>Microcystis aeruginosa</i>	3000 ng/L	18 ng PEQ/L	Halling-Sorensen, 2000
		LC50	1405-87-4	Bacitracin	<i>Artemia franciscana</i>	21820000 ng/L	81825 ng PEQ/L	Ferreira et al., 2007
Antibiotic activity	Sulphonamides	PNEC	72-14-0	Sulfathiazole	<i>Lemna gibba</i> (Inflated Duckweed)	100 ng/L	25 ng SEQ/L	Park and Choi, 2008
		NOEC	723-46-6	Sulfamethoxazole	<i>Danio rerio</i> (Zebrafish)	1000 ng/L	100 ng SEQ/L	Lin et al., Manuscript Draft
		LOEC	68-35-9	Sulfadiazine	<i>Danio rerio</i> (Zebrafish)	1000 ng/L	10 ng SEQ/L	Lin et al., Manuscript Draft
		EC50	723-46-6	Sulfamethoxazole	<i>Synechococcus leopoliensis</i>	26800 ng/L	2680 ng SEQ/L	Ferrari et al., 2004
		LC50	723-46-6	Sulfamethoxazole	<i>Brachionus calyciflorus</i> (Rotifer)	9630000 ng/L	9630 ng SEQ/L	Isidori et al., 2005
	Tetracyclines	PNEC	79-57-2	Oxytetracycline	<i>Selenastrum capricornutum</i> (Green algae)	170 ng/L	170 ng OEQ/L	Park and Choi, 2008
		NOEC	15318-45-3	Thiamphenicol	<i>Selenastrum capricornutum</i> (Green algae)	4060000 ng/L	10150 ng OEQ/L	Eguchi et al., 2004
		LOEC	56-75-7	Chloramphenicol	<i>Tetraselmis suecica</i>	2500000 ng/L	5000 ng OEQ/L	Seoane et al., 2014
		EC50	56-75-7	Chloramphenicol	<i>Vibrio fisheri</i>	64300 ng/L	643 ng OEQ/L	Backhaus and Grimme, 1999
		LC50	79-57-2	Oxytetracycline	<i>Brachionus calyciflorus</i> (Rotifer)	1870000 ng/L	1870 ng OEQ/L	Isidori et al., 2005
Quinolones		PNEC	93106-60-6	Enrofloxacin	<i>Microcystis aeruginosa</i>	49 ng/L	196 ng FEQ/L	Park and Choi, 2008
		NOEC	3380-34-5	Triclosan	<i>Selenastrum capricornutum</i> (Green algae)	200 ng/L	2 ng FEQ/L	Yang et al., 2008
		LOEC	3380-34-5	Triclosan	<i>Selenastrum capricornutum</i> (Green algae)	400 ng/L	0.8 ng FEQ/L	Yang et al., 2008
		EC50	3380-34-5	Triclosan	<i>Selenastrum capricornutum</i> (Green algae)	530 ng/L	0.53 ng FEQ/L	Yang et al., 2008
		LC50	3380-34-5	Triclosan	<i>Pimephales promelas</i> (Fathead Minnow)	260000 ng/L	2.6 ng FEQ/L	Ishibashi et al., 2004

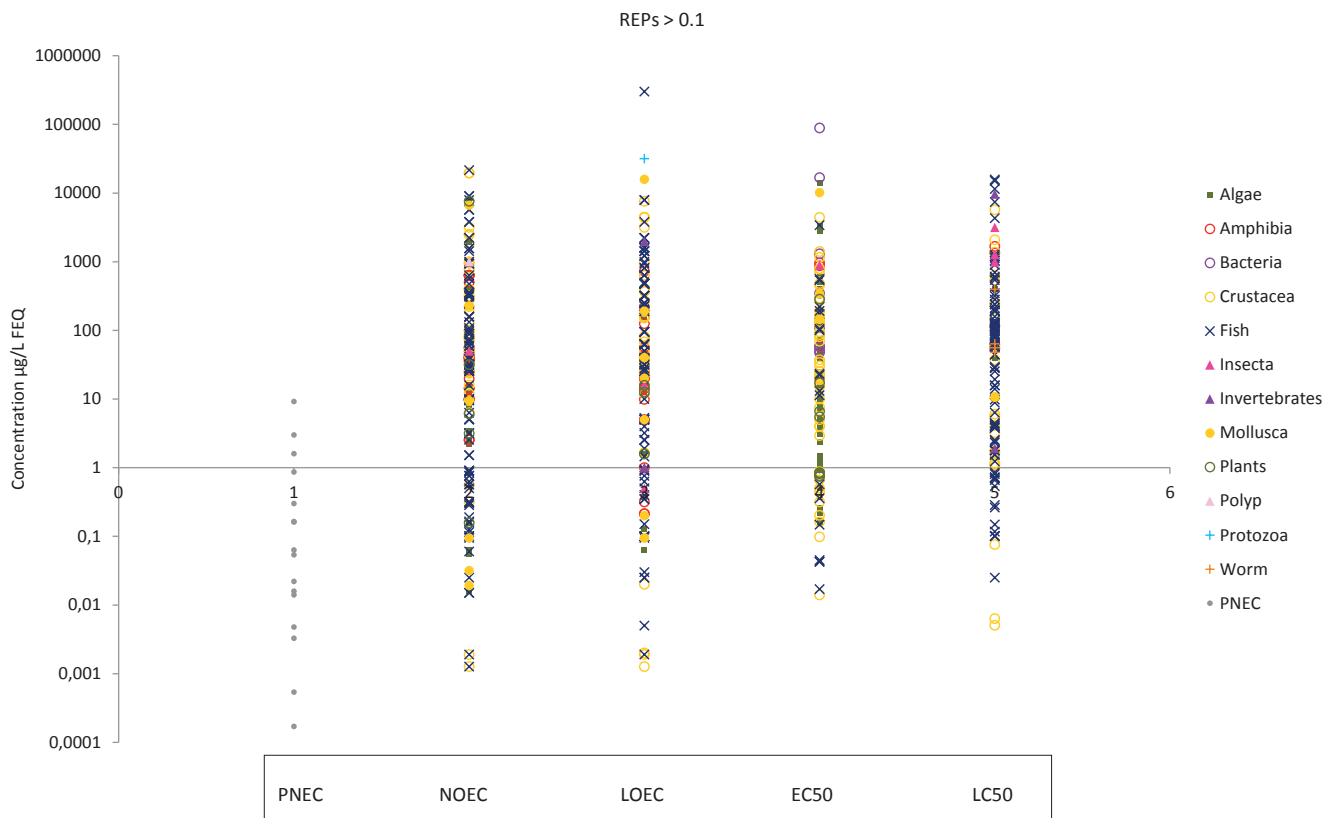
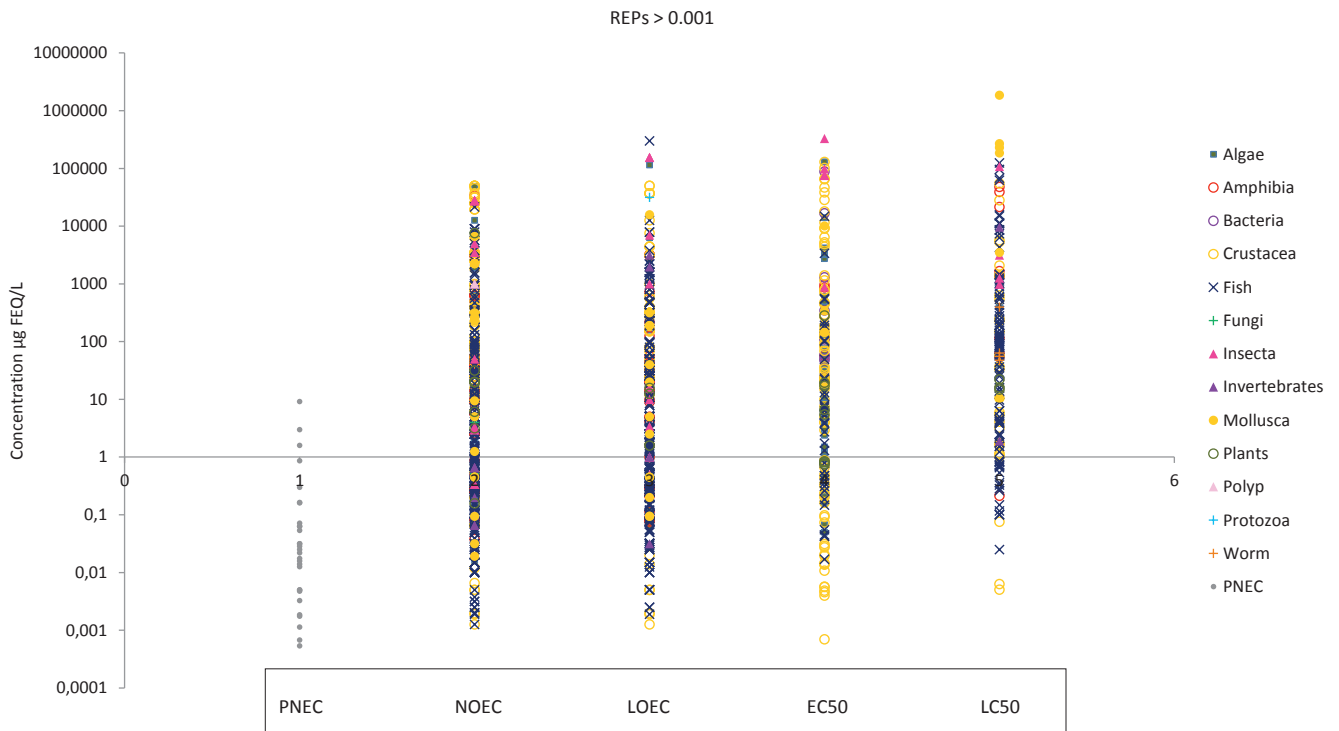
# APPENDIX SI-IV

Graphic representations of the toxicity data collected for each bioassay for REP1 and REP2 groups

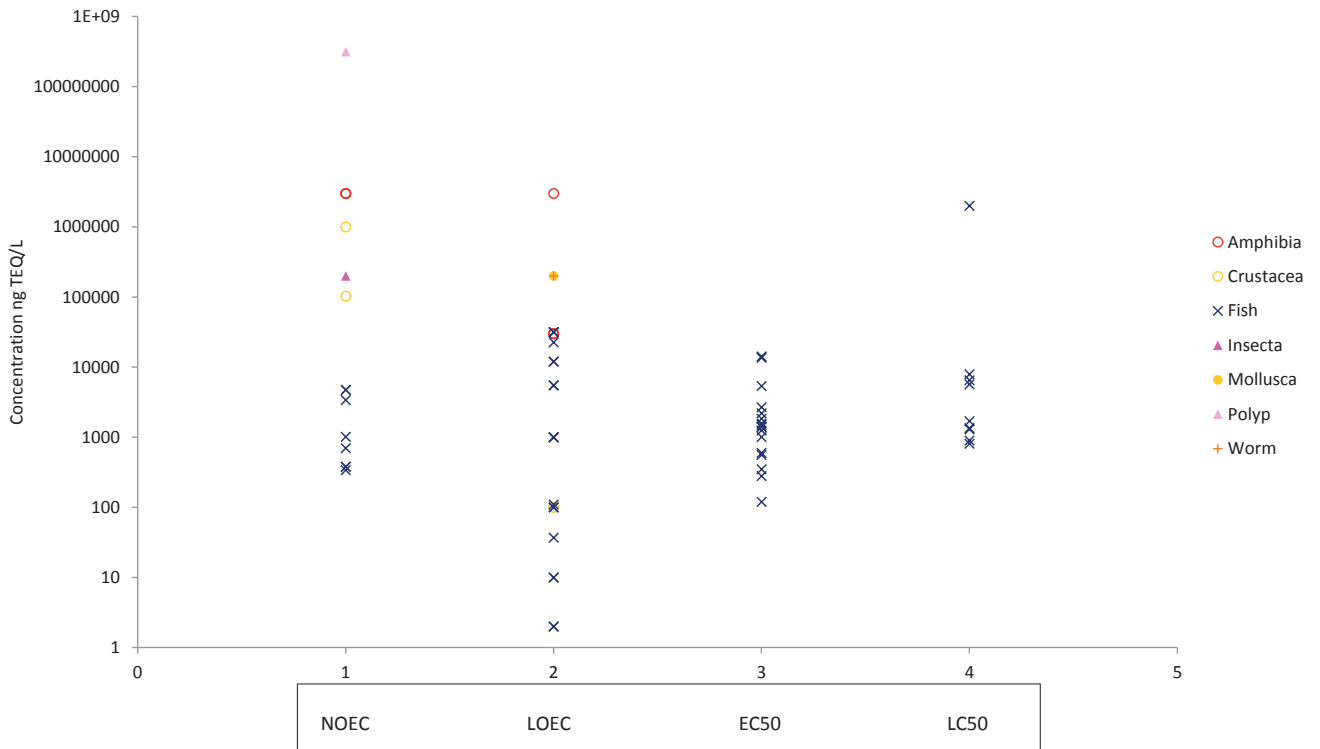
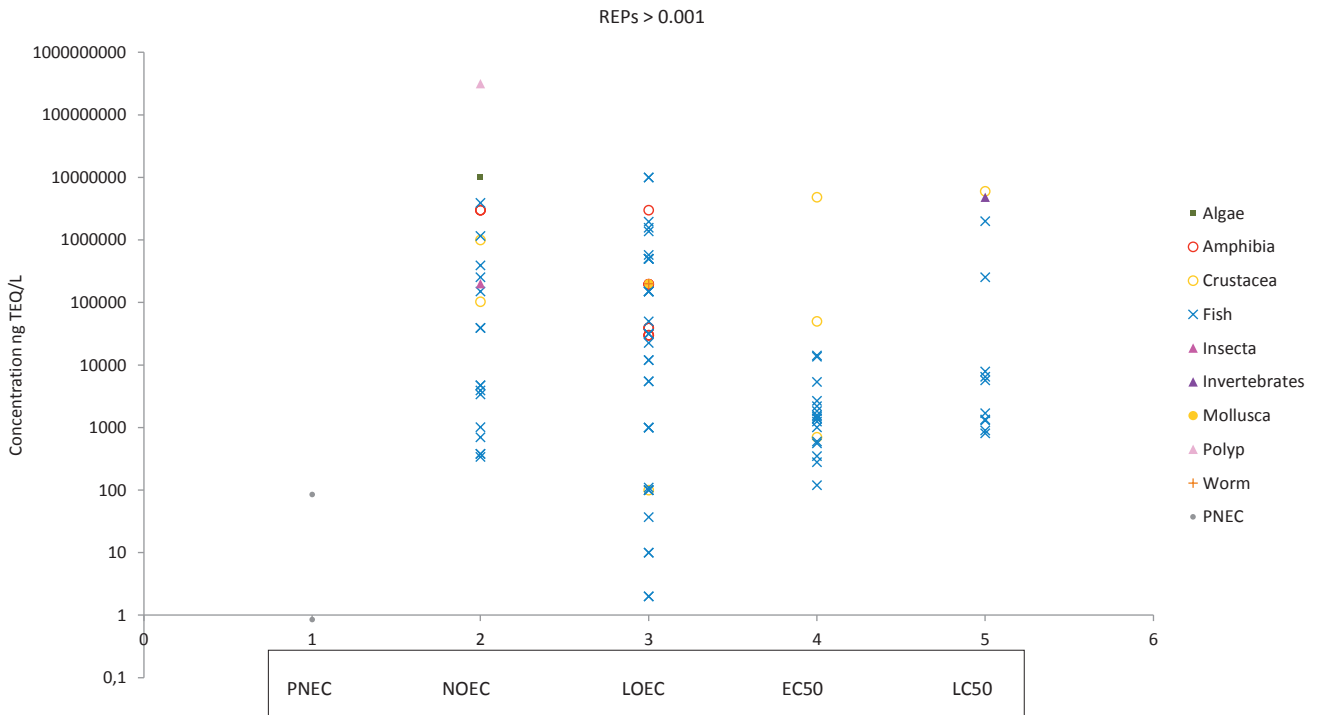
## A: ER CALUX

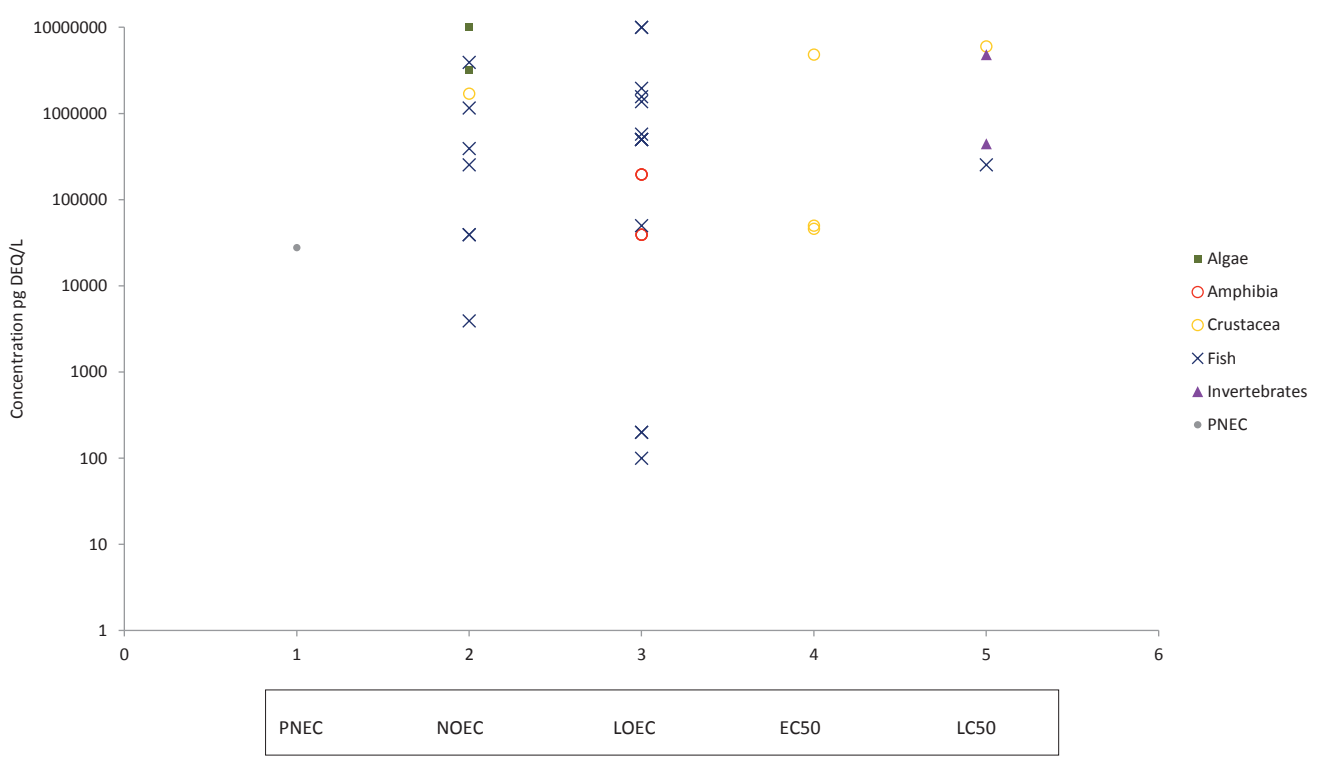
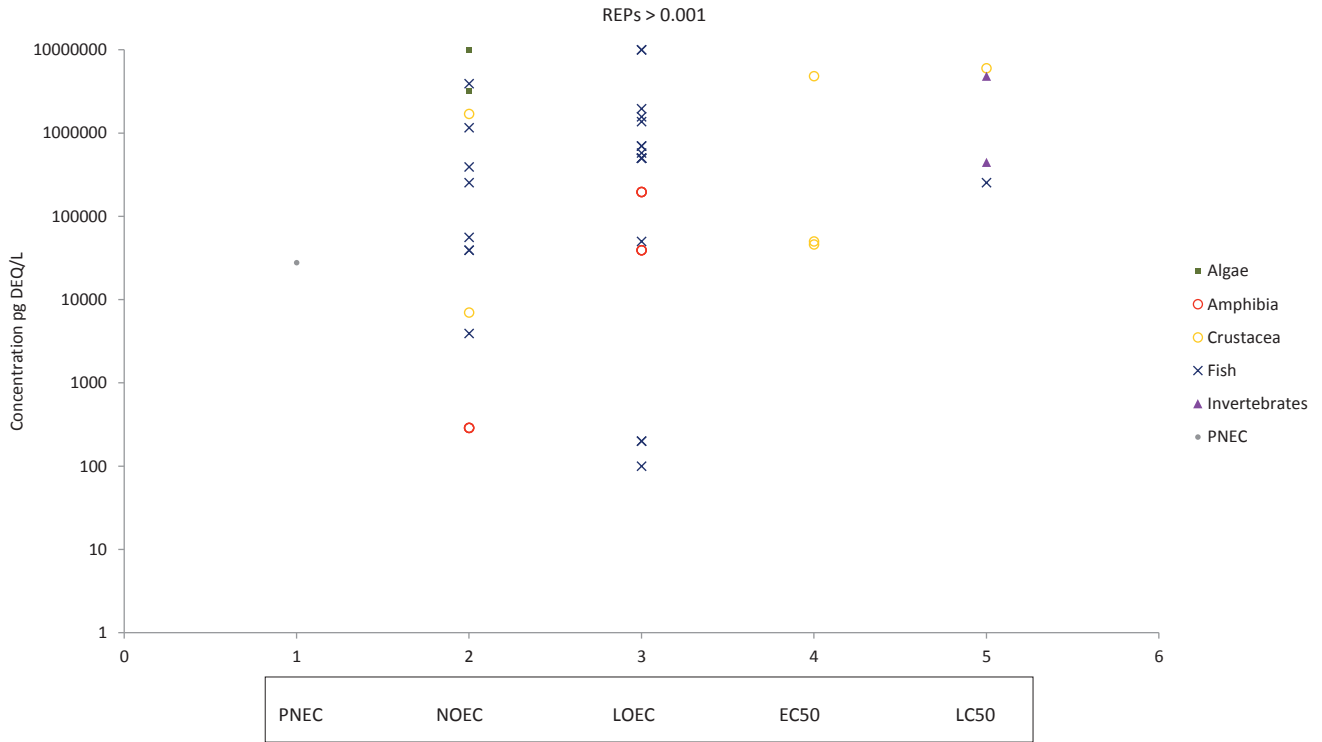


## B: ANTI-AR CALUX



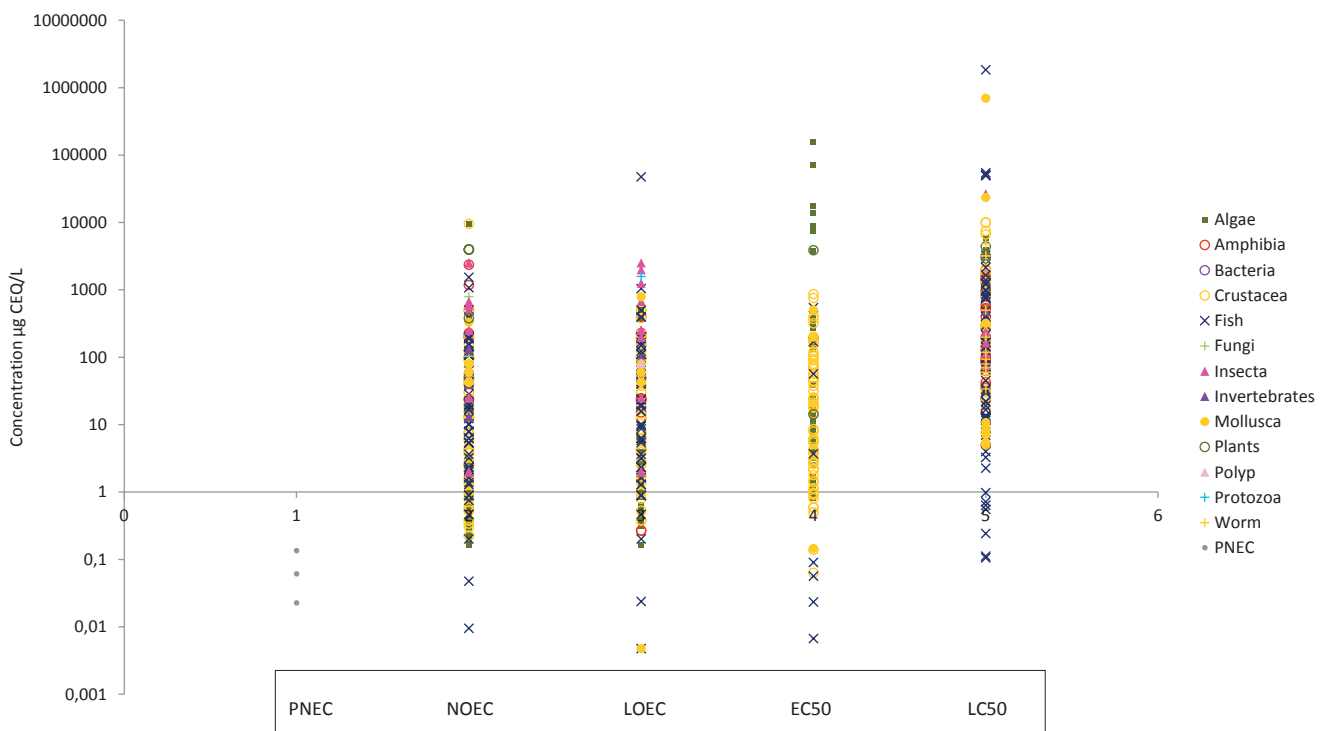
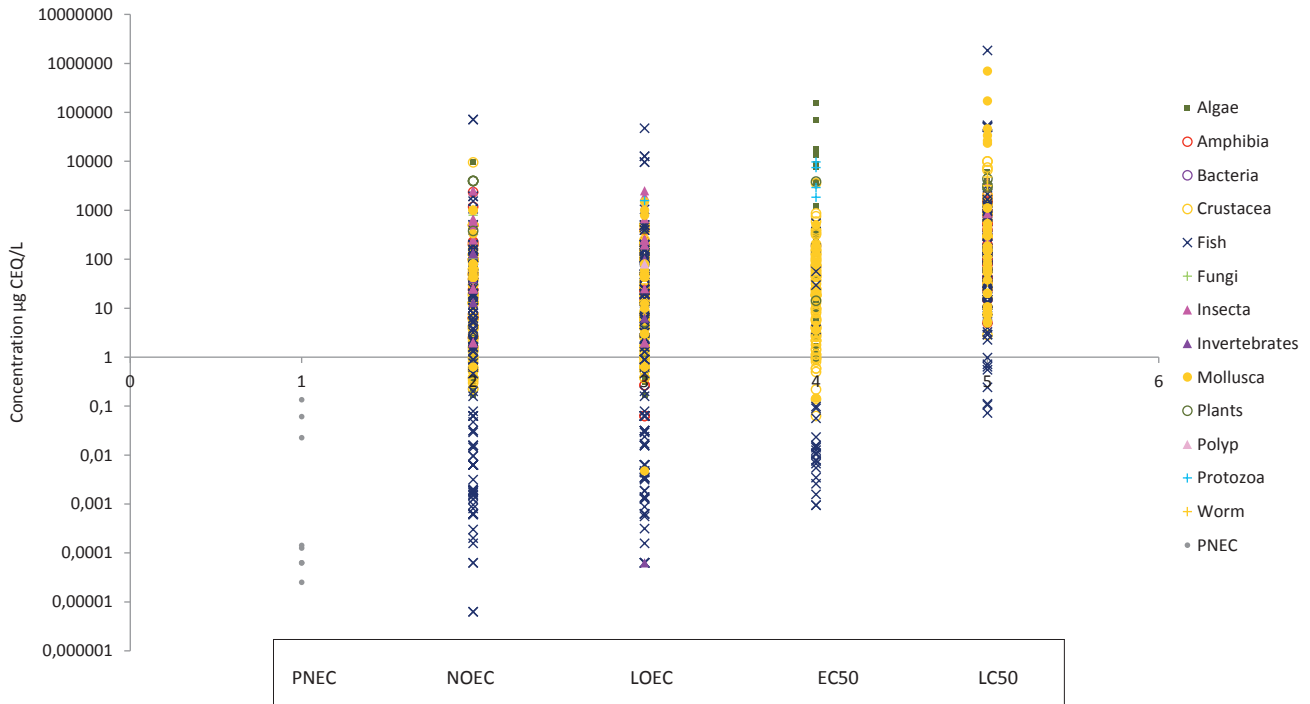
### C: DR CALUX



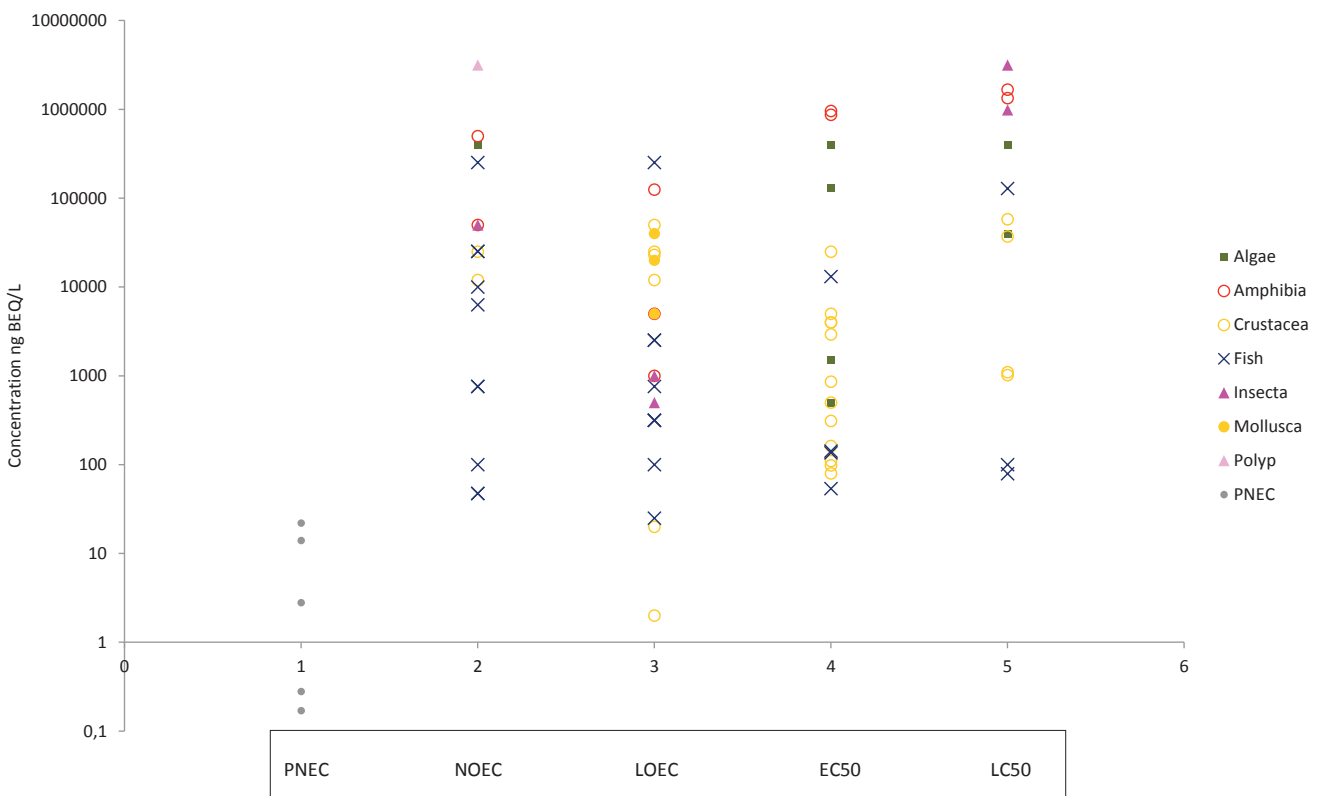
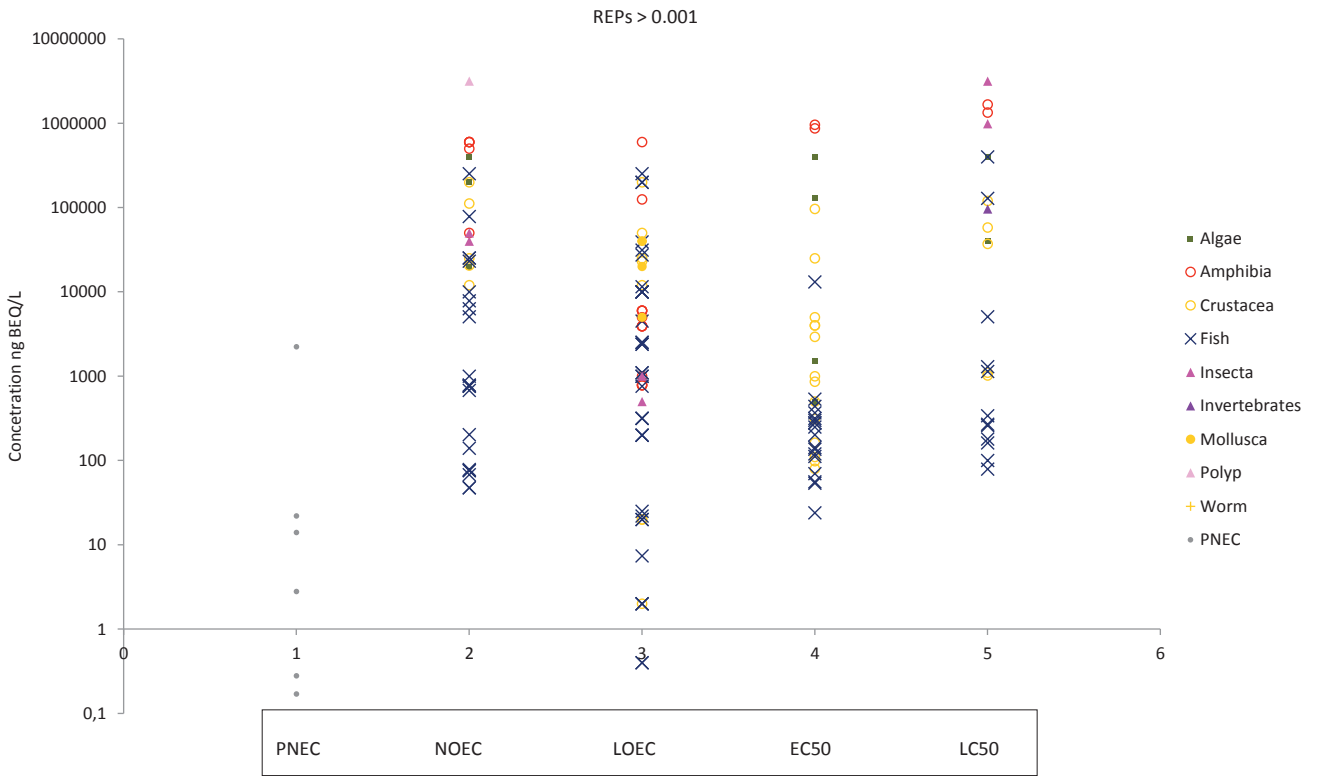


## D: GR CALUX

REPs > 0.001

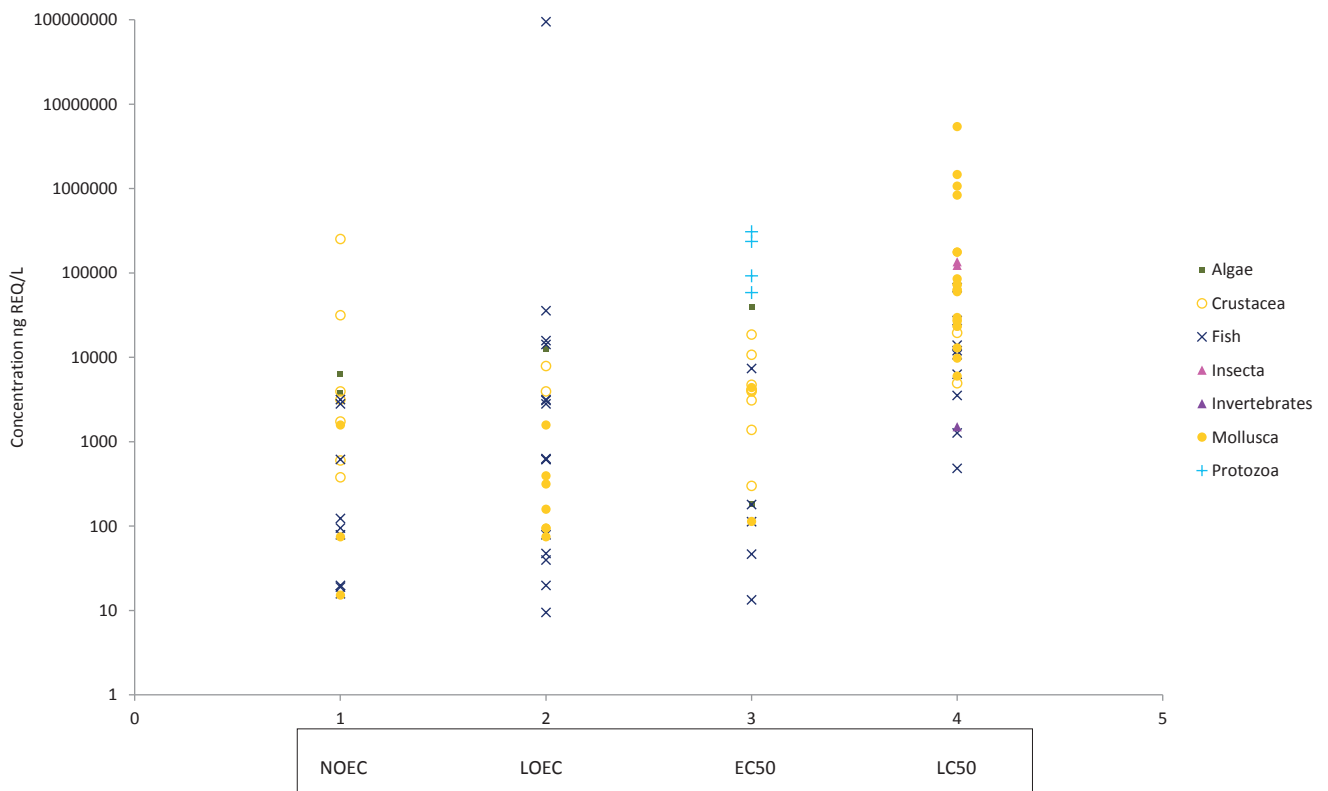
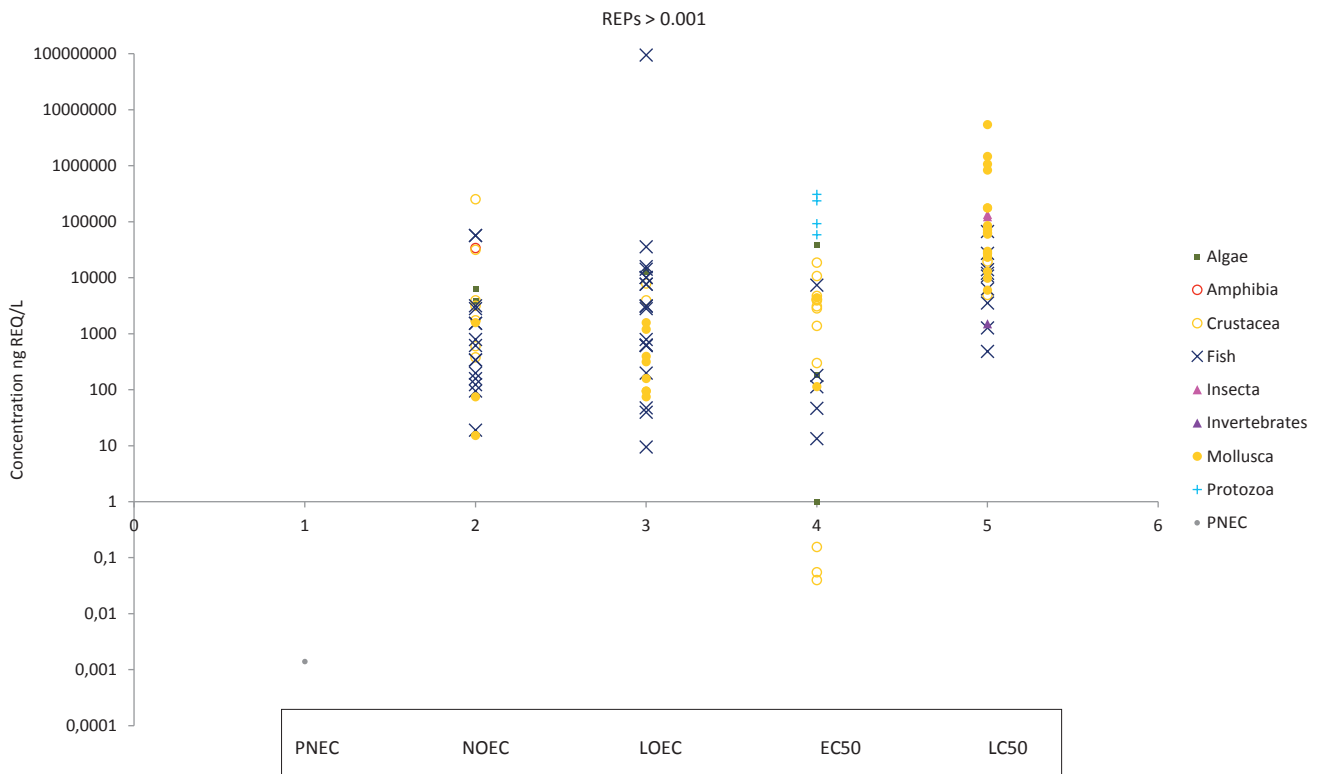


### E: NRF2 CALUX

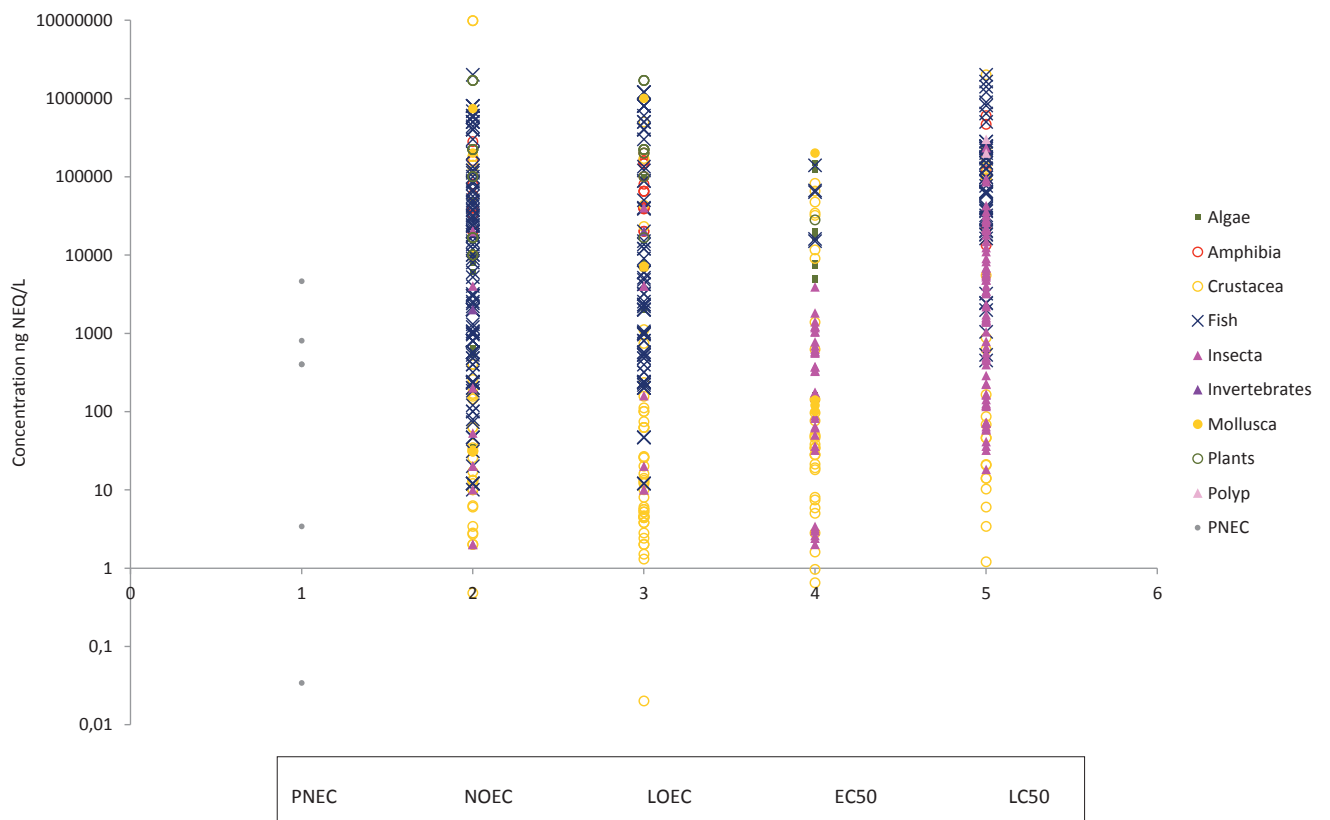
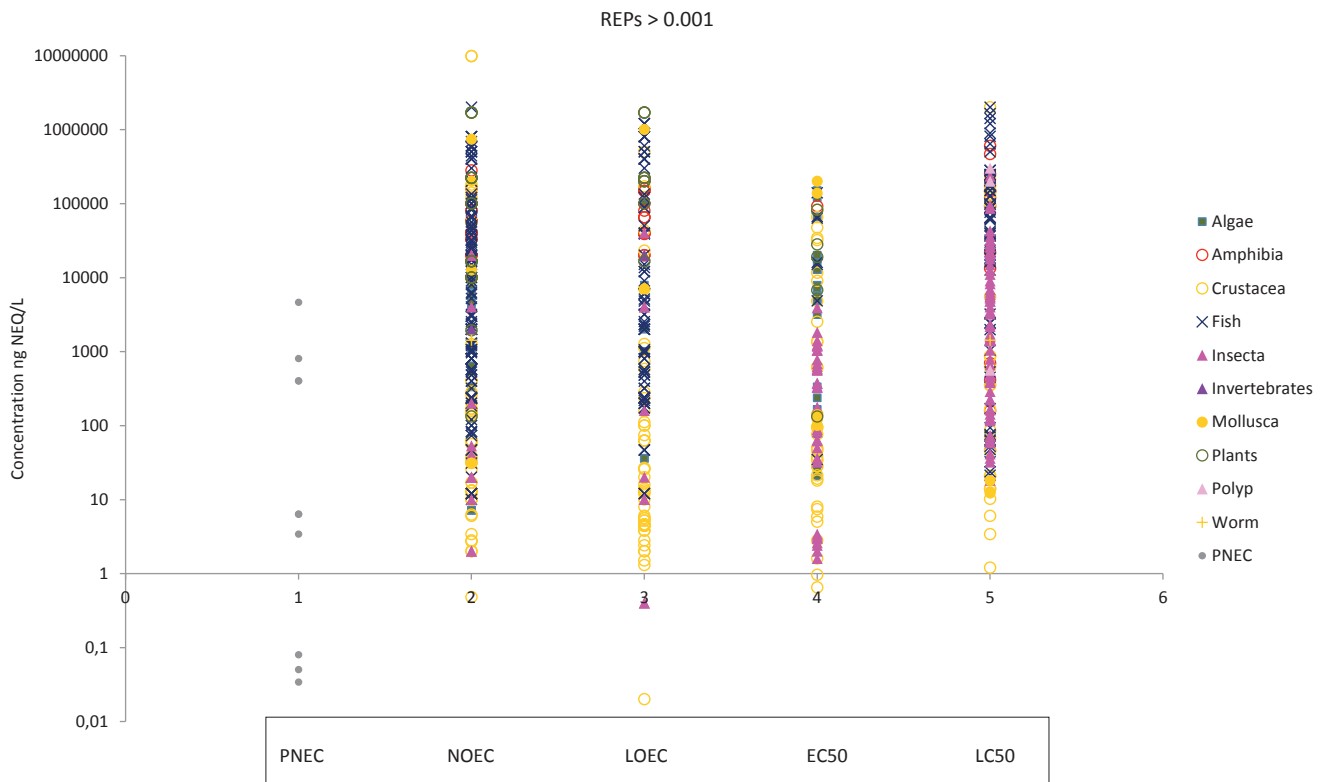




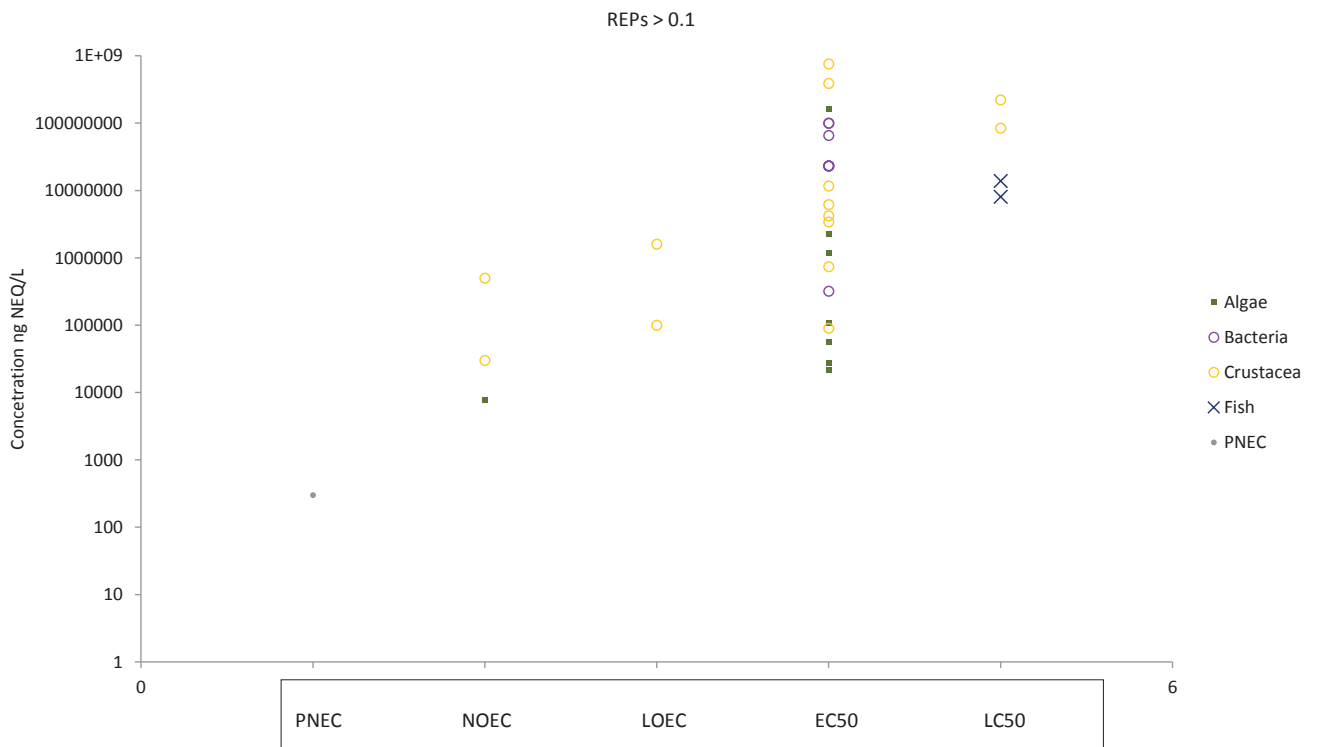
## F: PAH CALUX



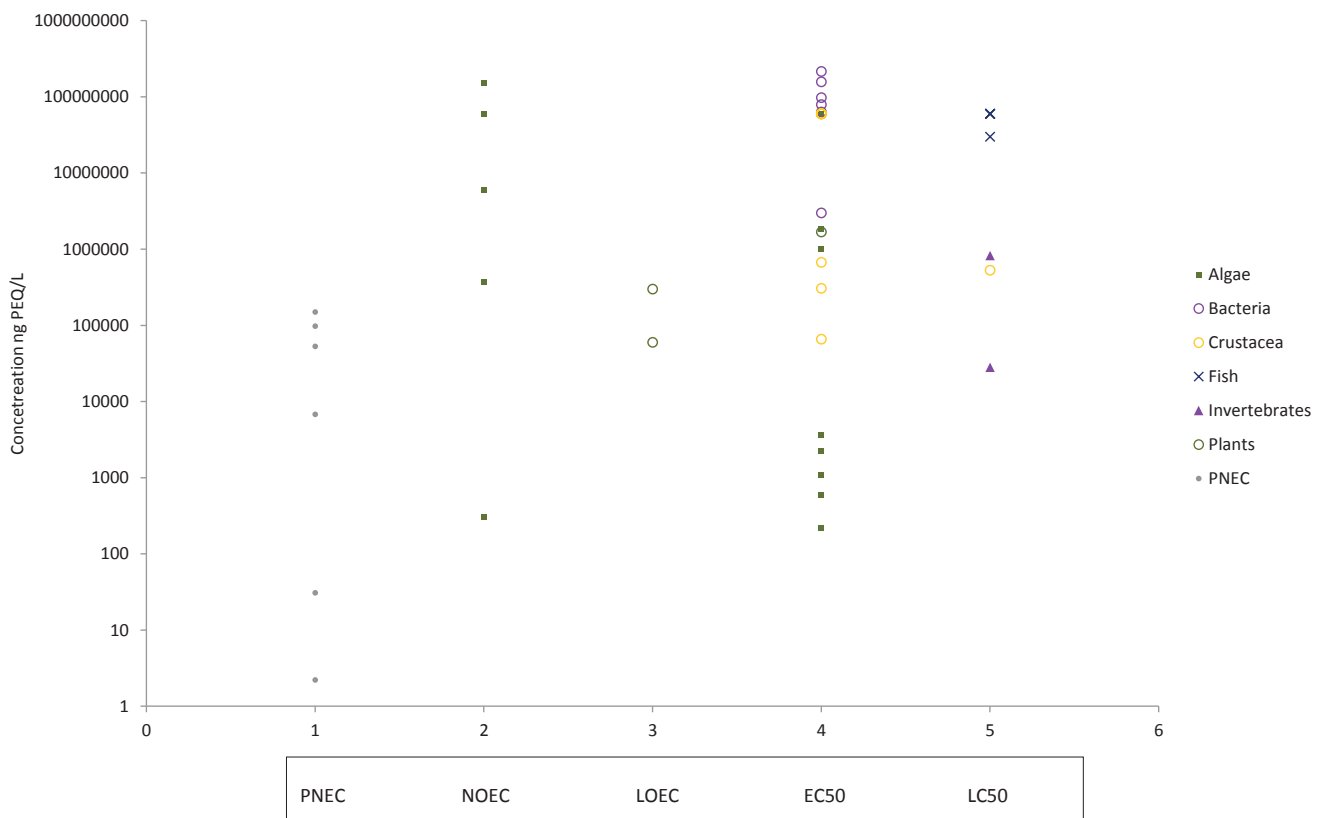
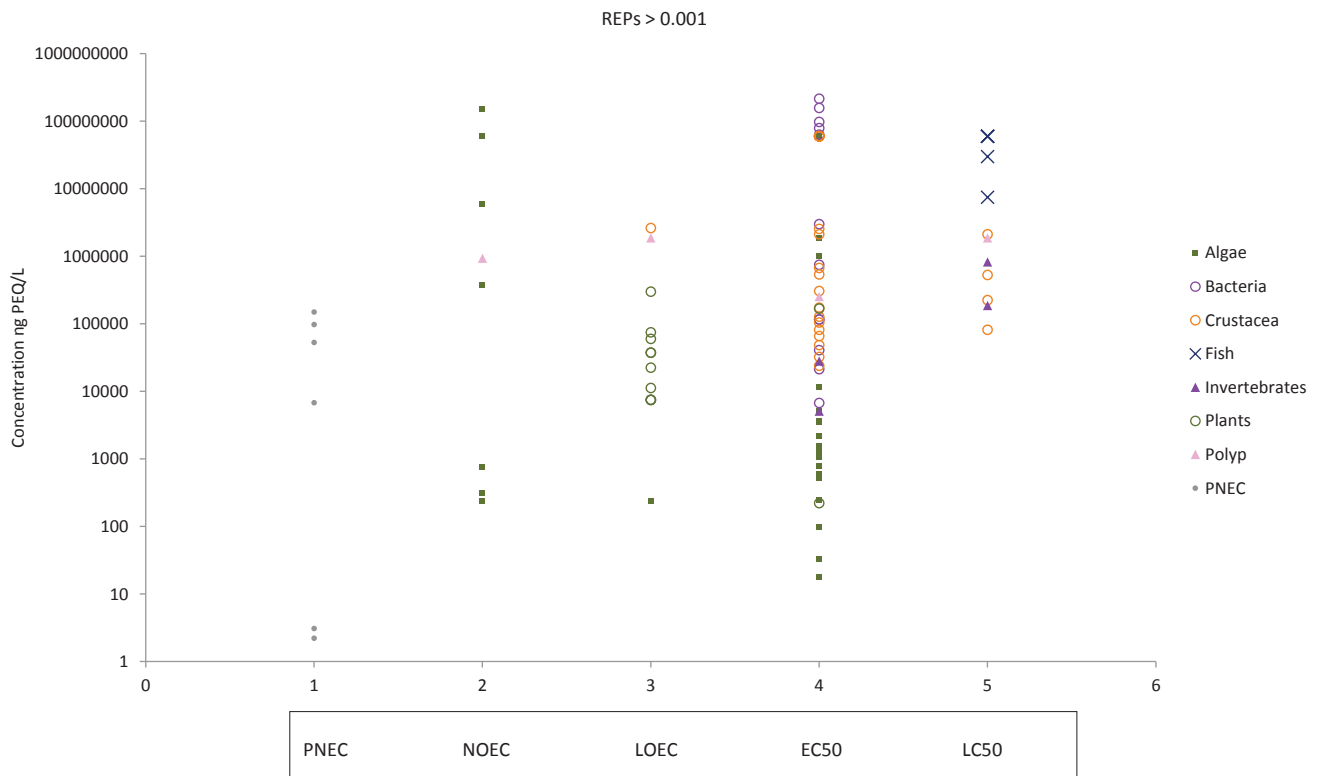
### G: PPARG CALUX



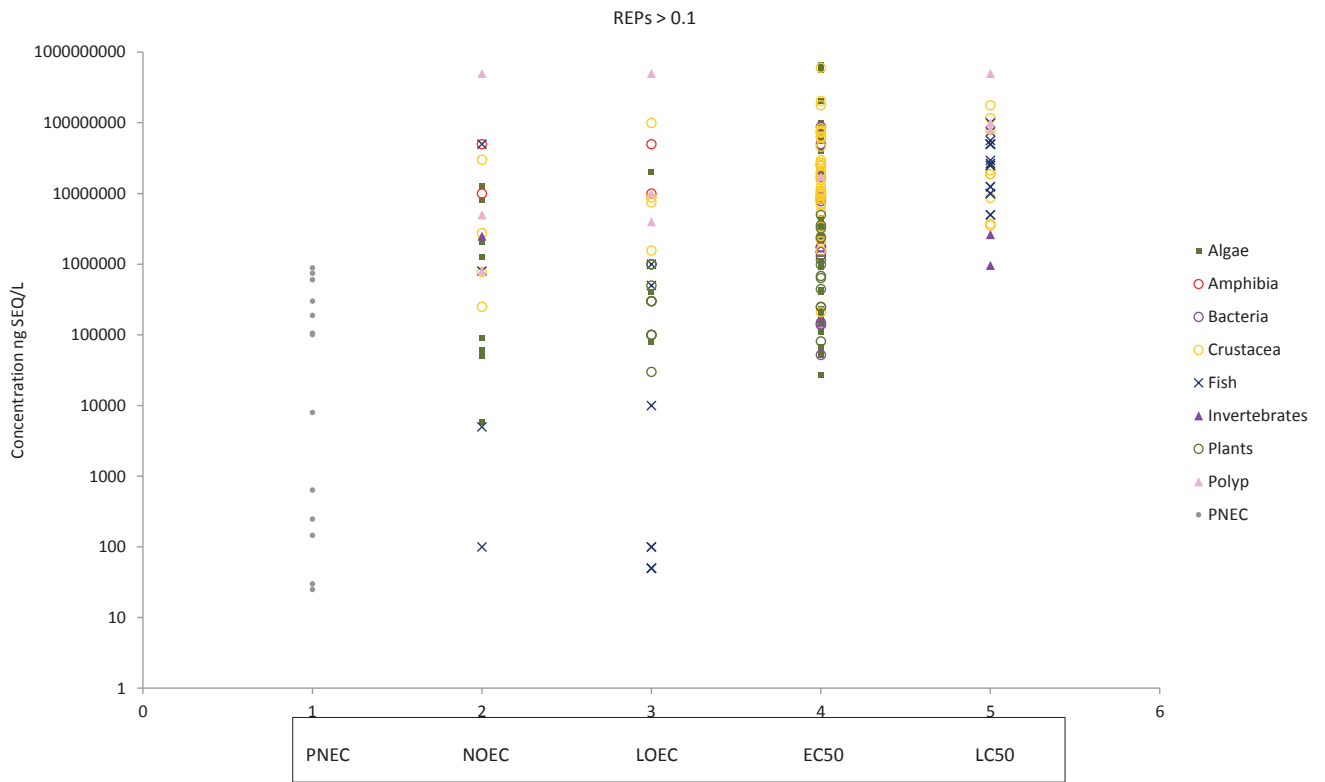
## H: PXR CALUX



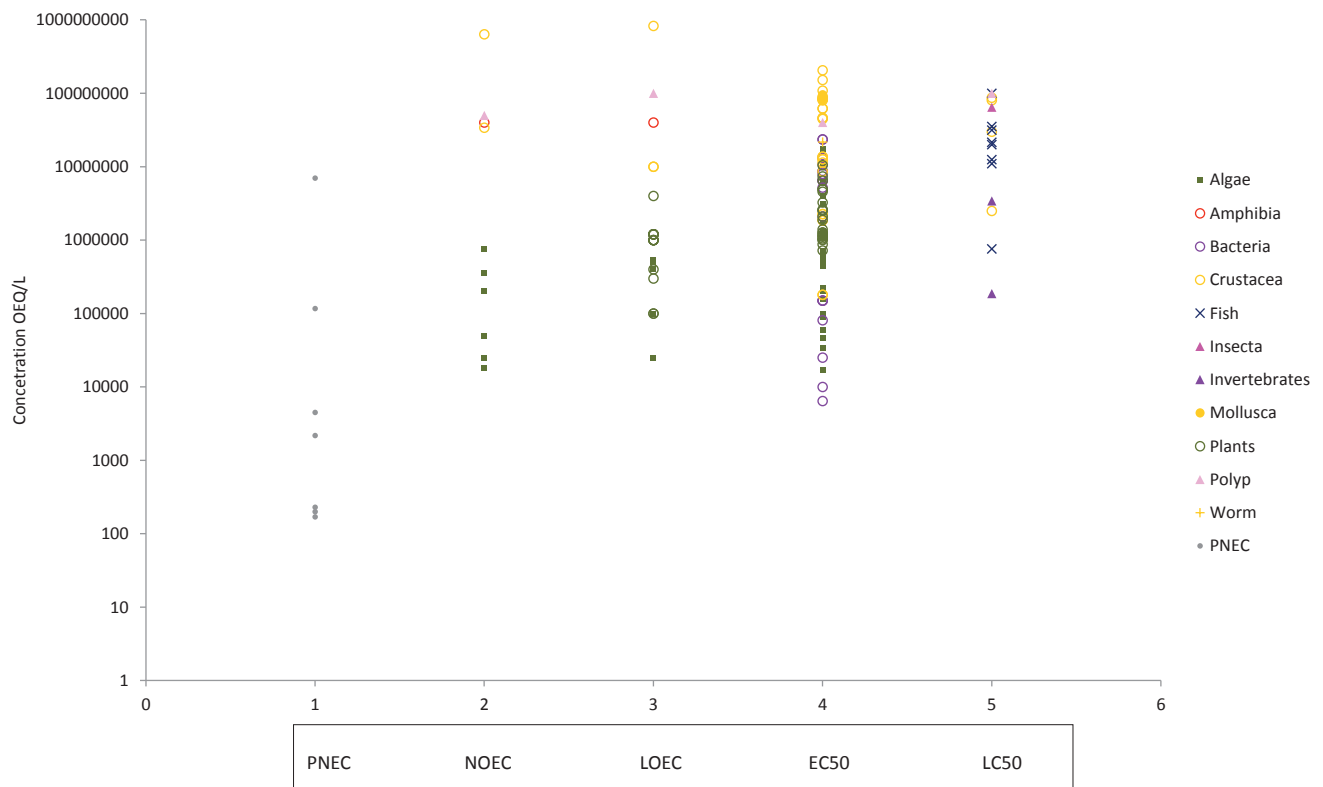
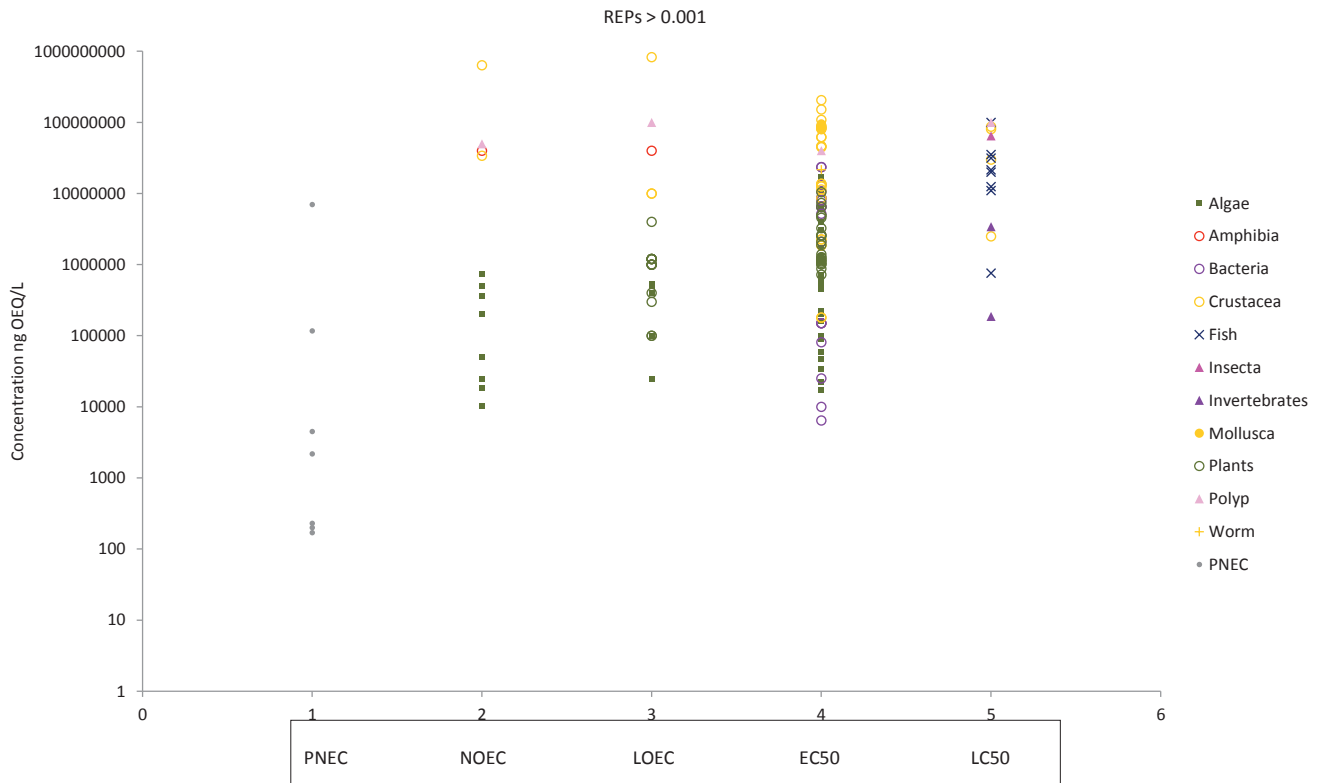
## I.1 AMINOGLYCOSIDES



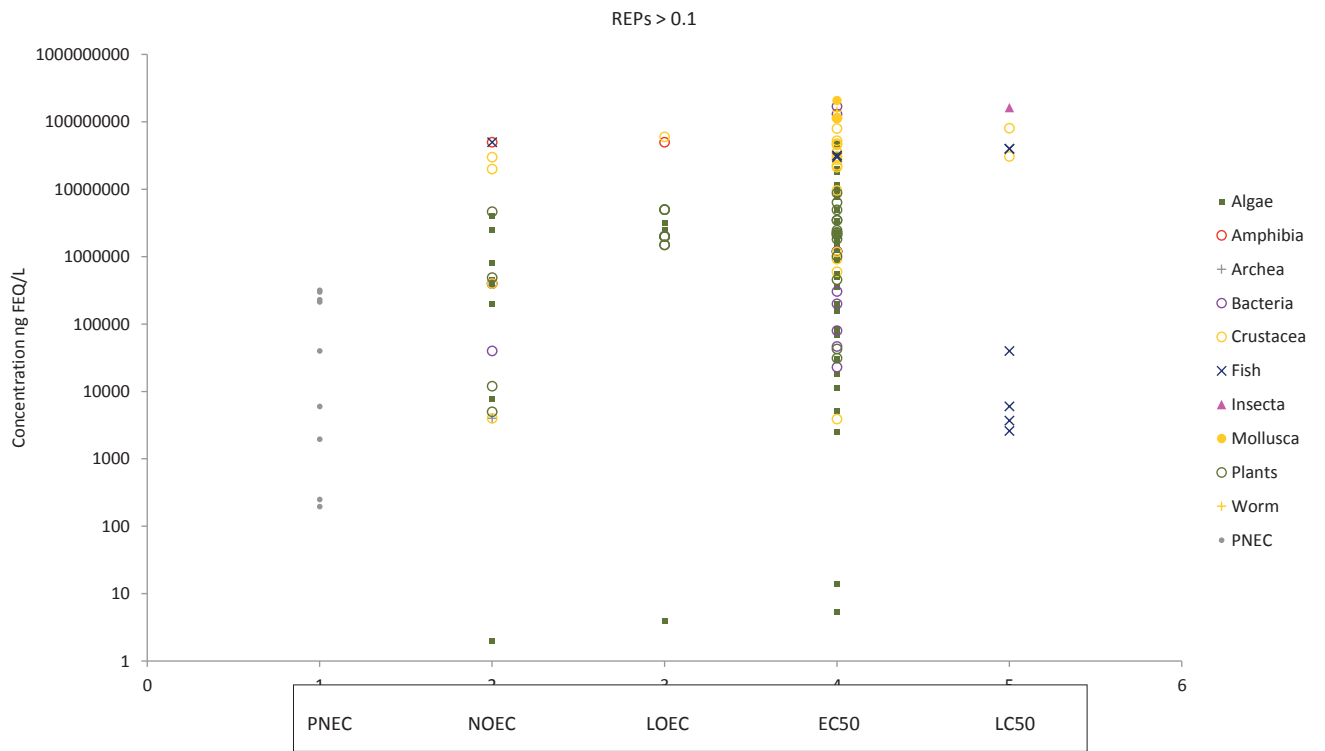
## I.2 MACROLIDES & B LACTAMS



### I.3 SULFONAMIDES

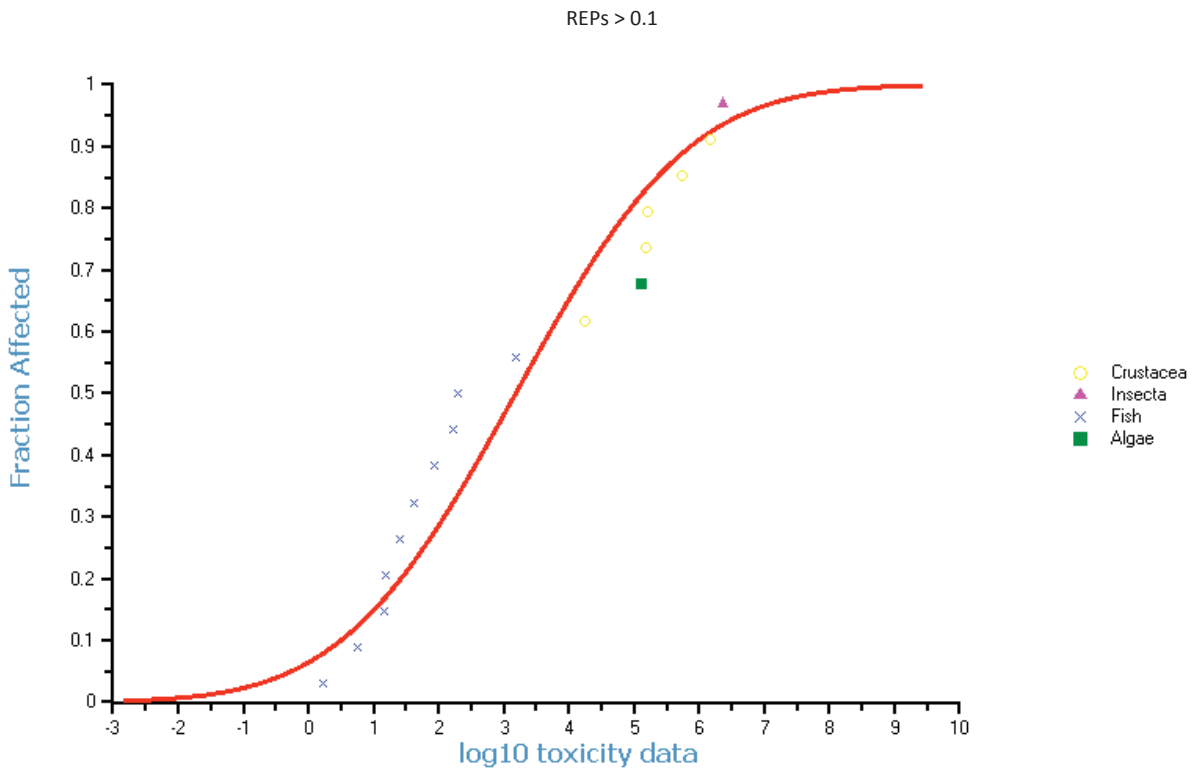
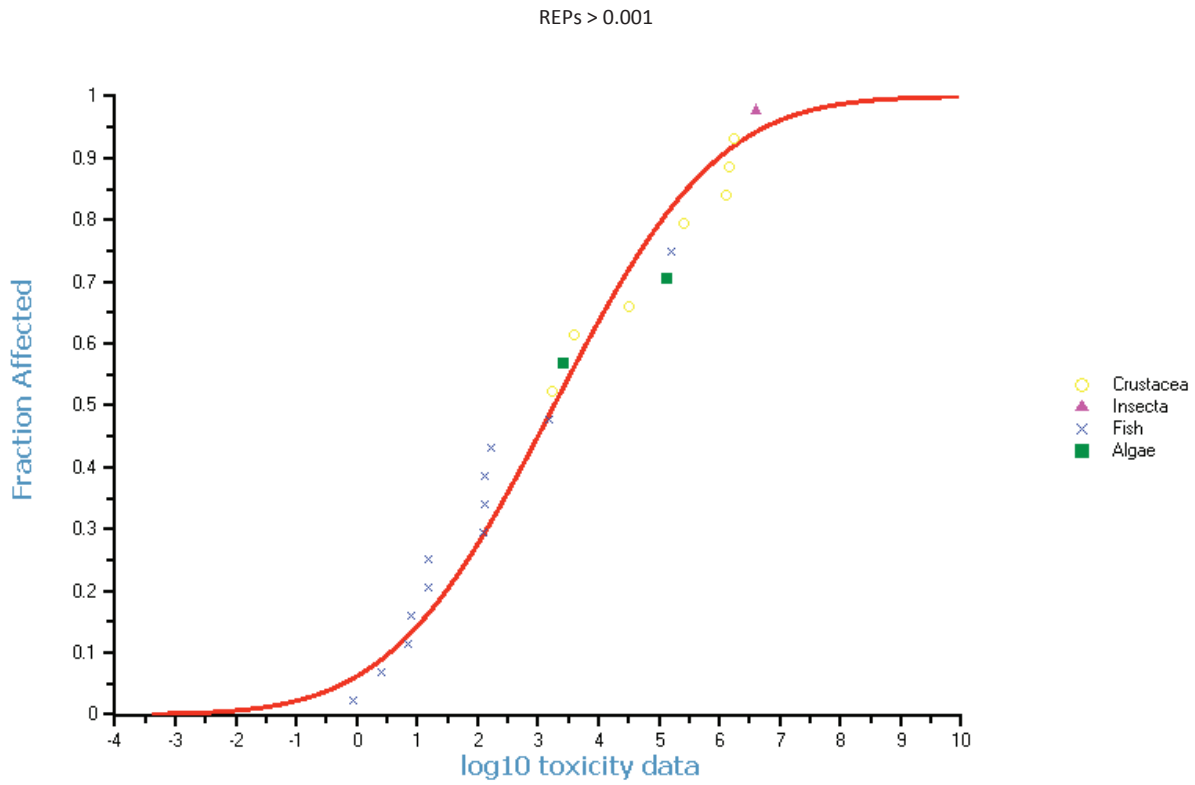


## I.4 TETRACYCLINES





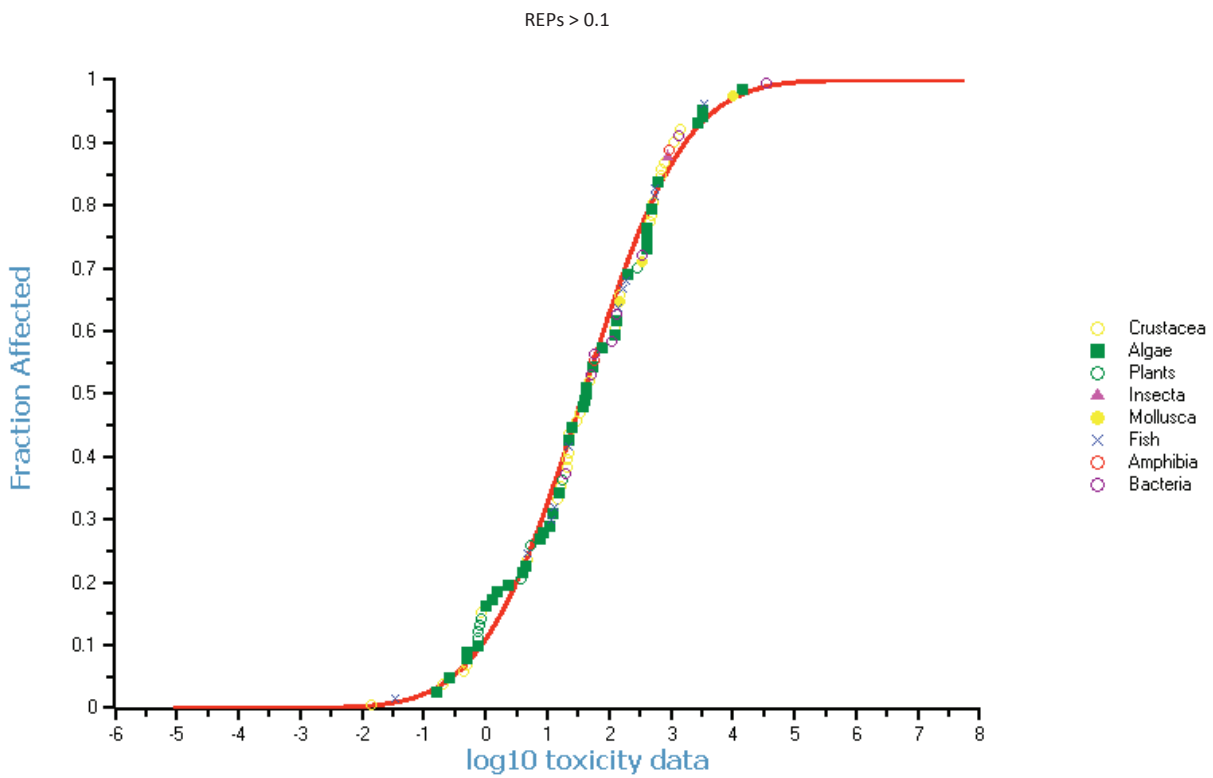
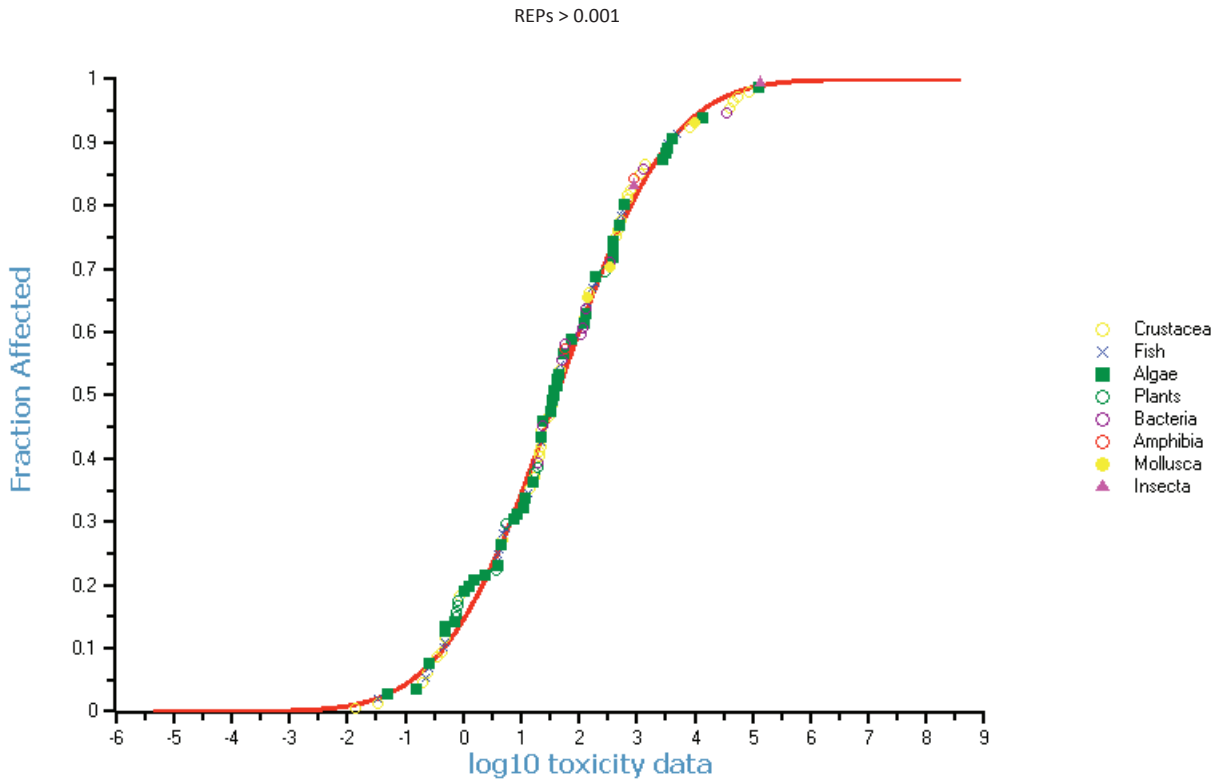
## I.5 QUINOLONES



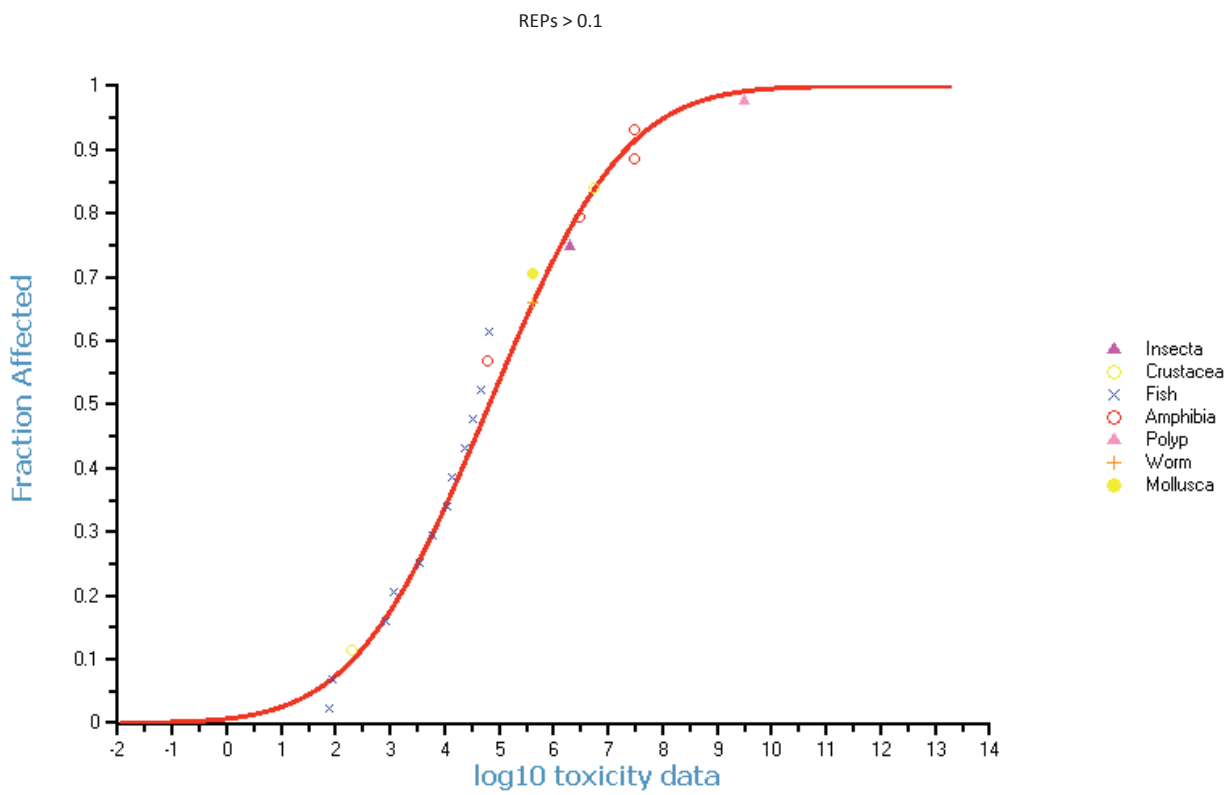
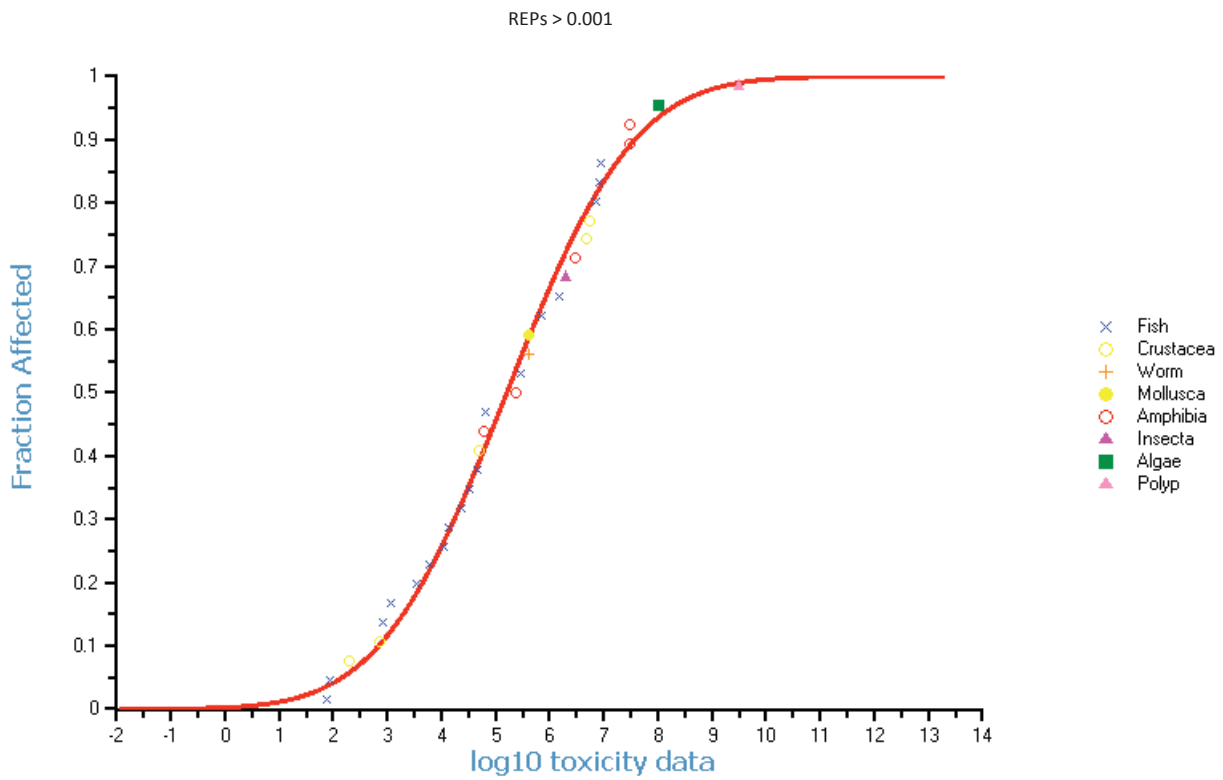
# APPENDIX SI-V

SSD analyses of all bioassays for REP1 and REP2 groups

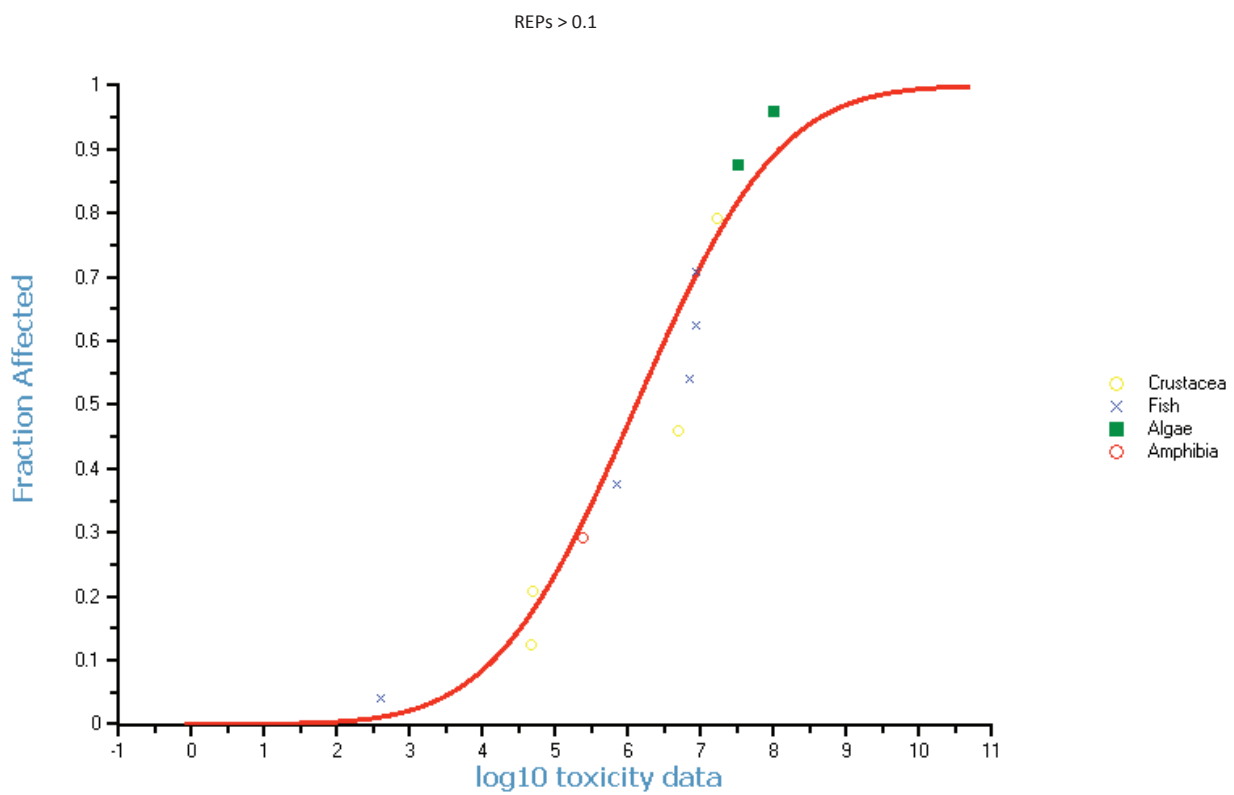
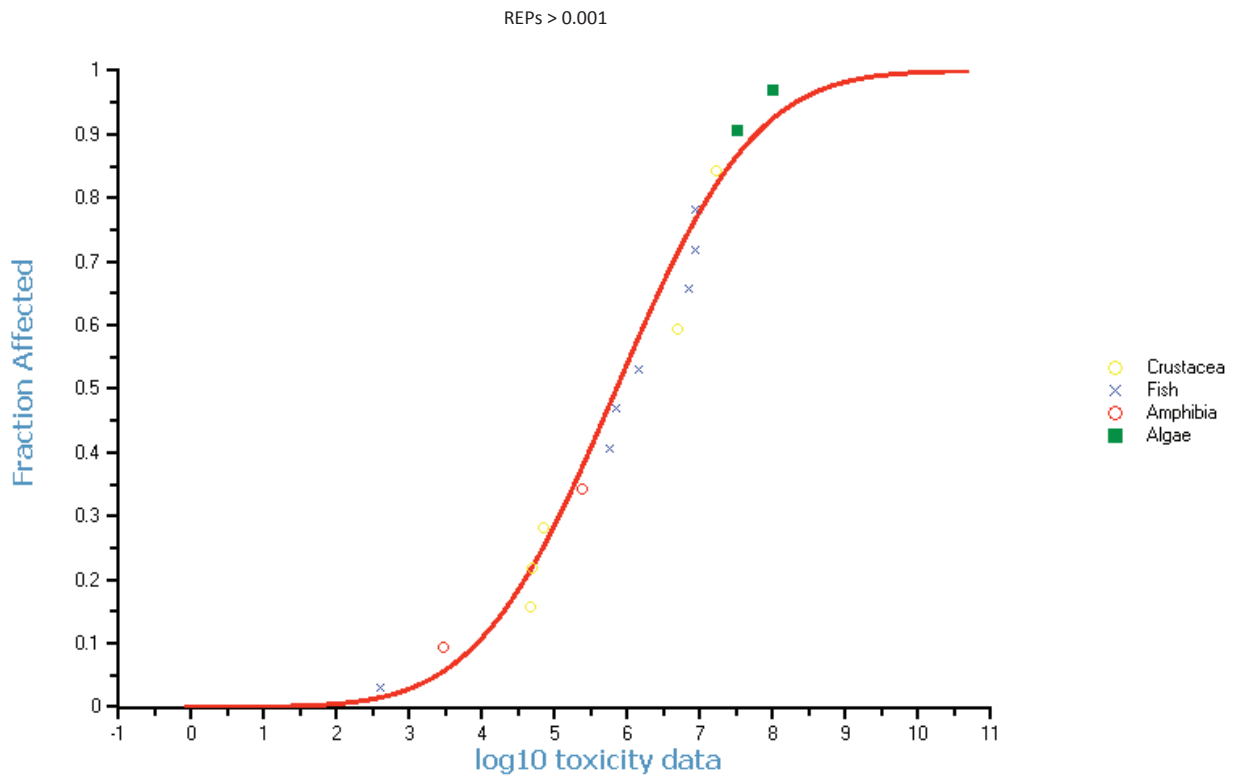
## A: ER CALUX (NG EEQ/L)



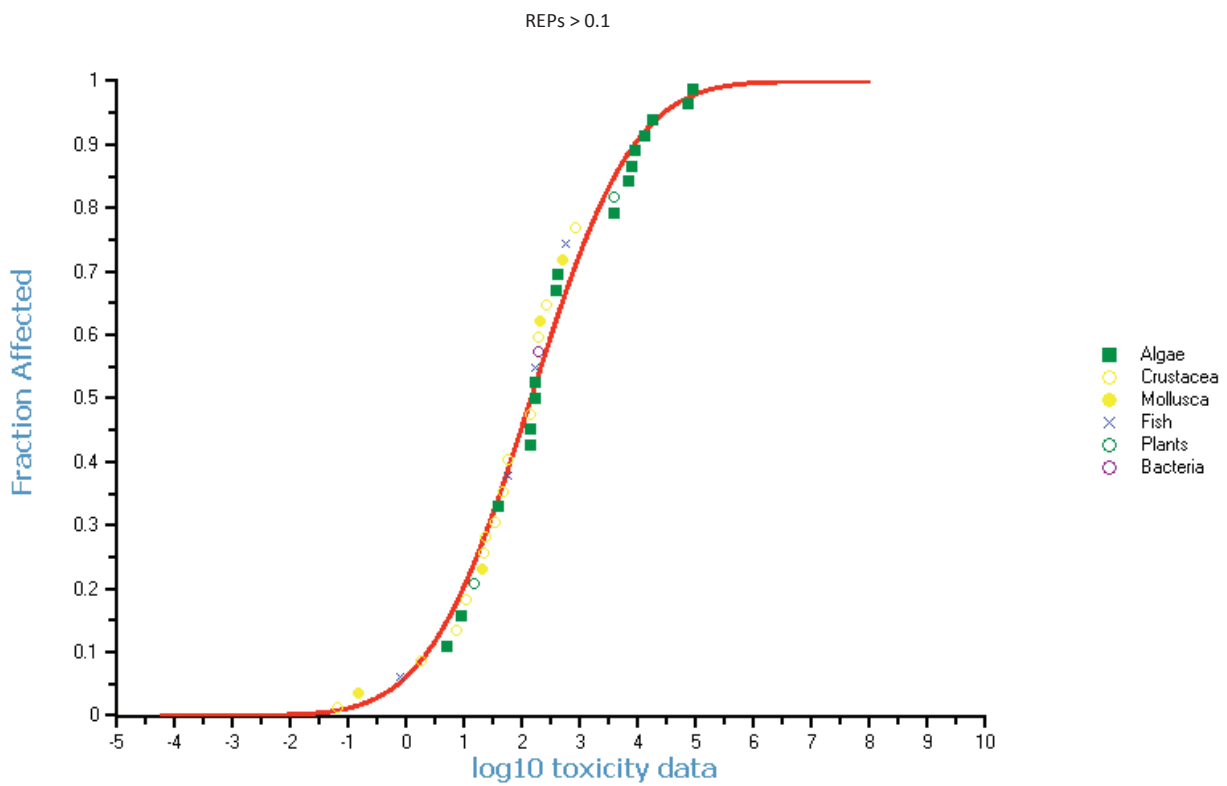
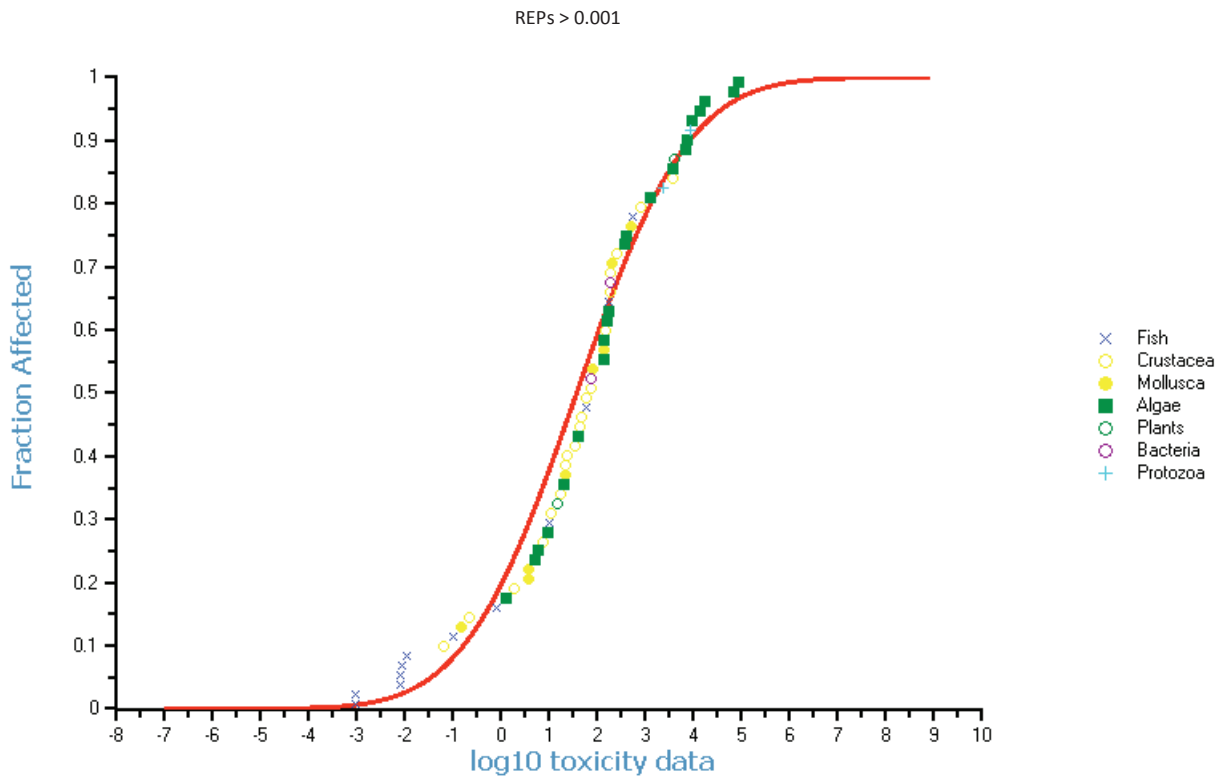
### B: ANTI-AR CALUX (MG FEQ/L)



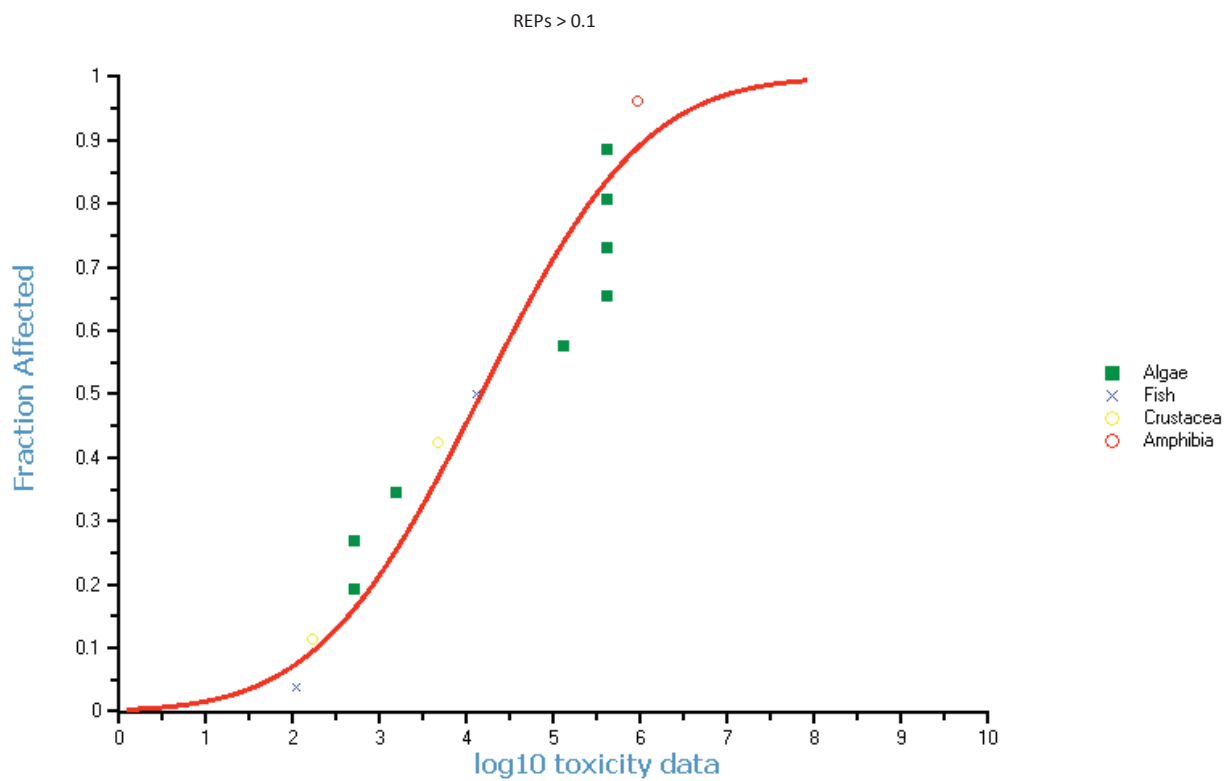
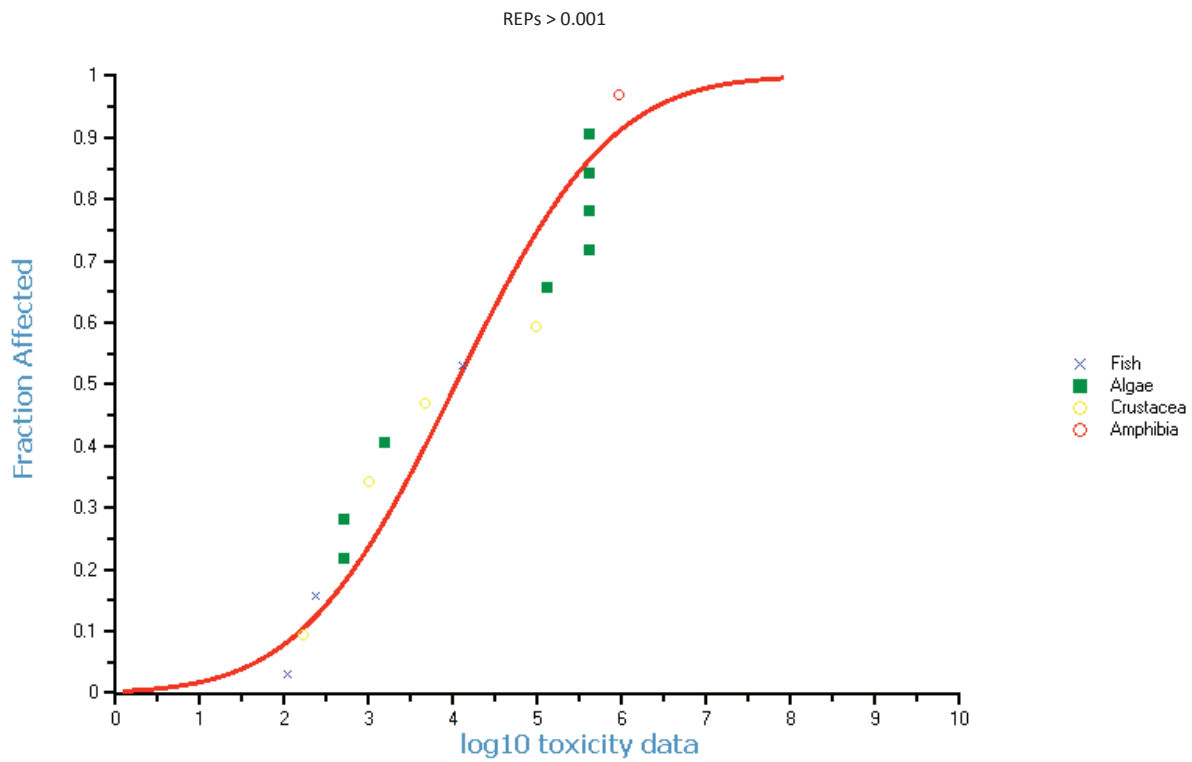
### C: DR CALUX (PG TEQ/L)



### D: GR CALUX (NG DEQ/L)

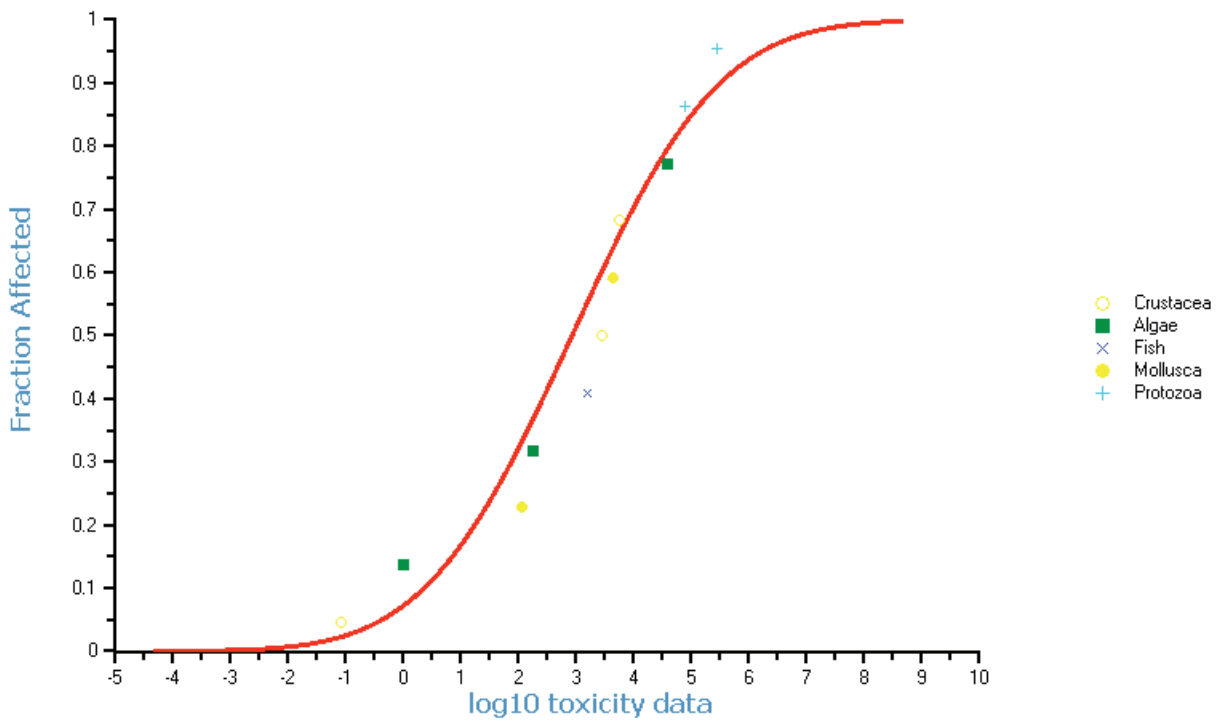


### E: NRF2 CALUX (MG CEQ/L)

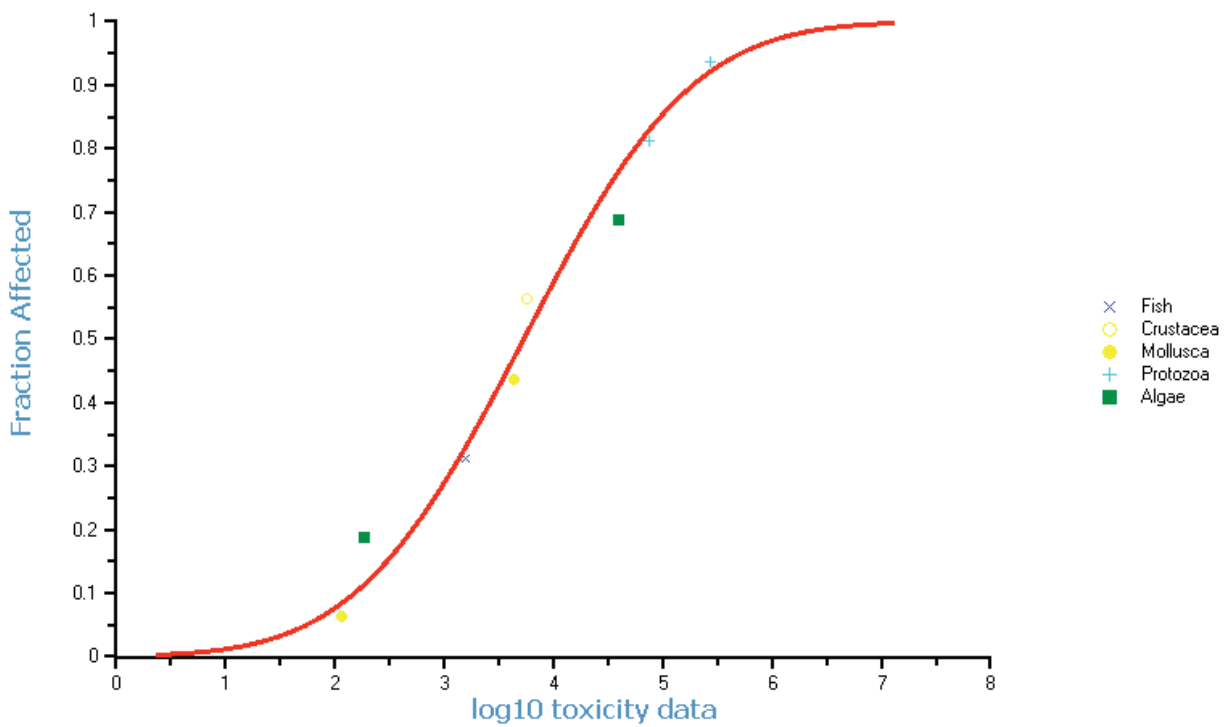


### F: PAH CALUX (NG BEQ/L)

REPs > 0.001

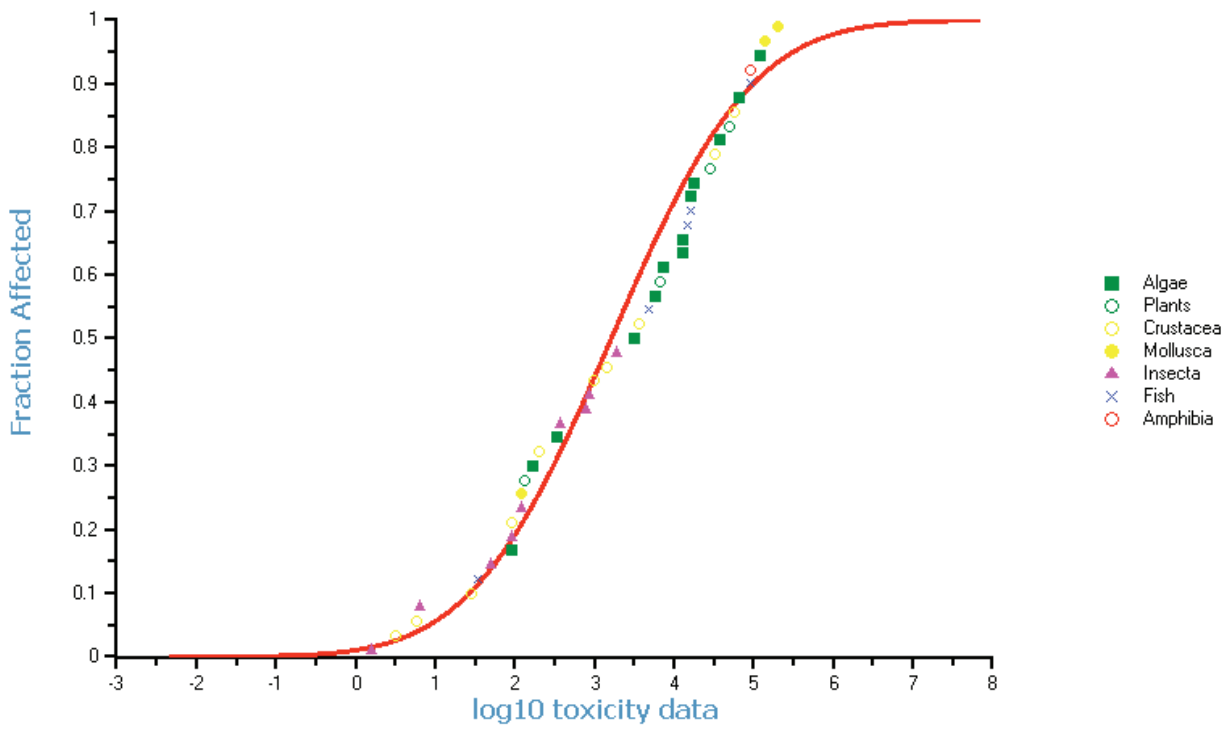


REPs > 0.1

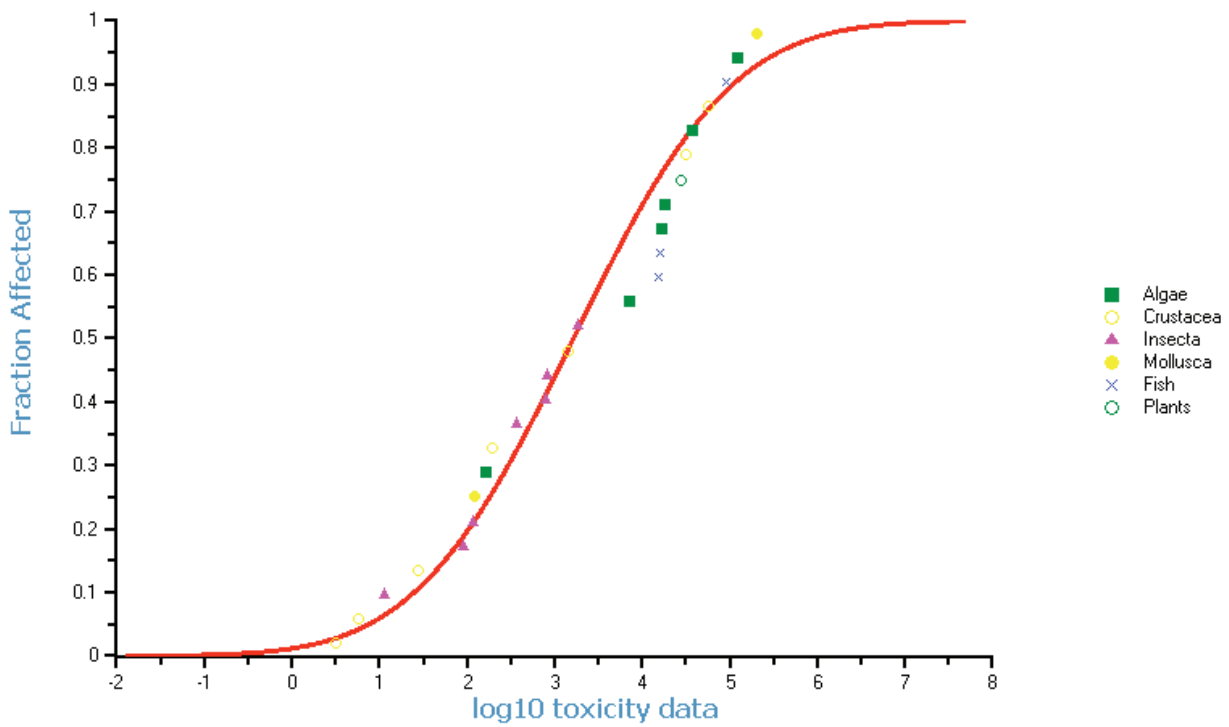


### G: PPARG CALUX (NG REQ/L)

REPs > 0.001

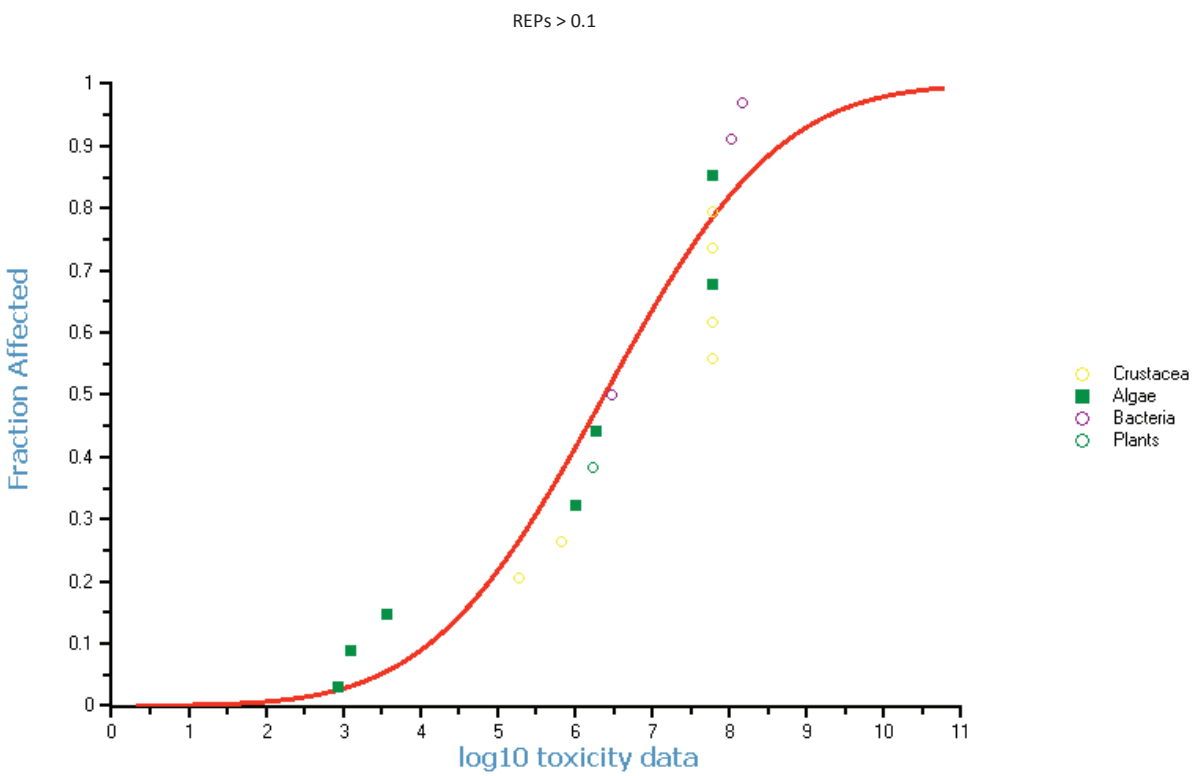
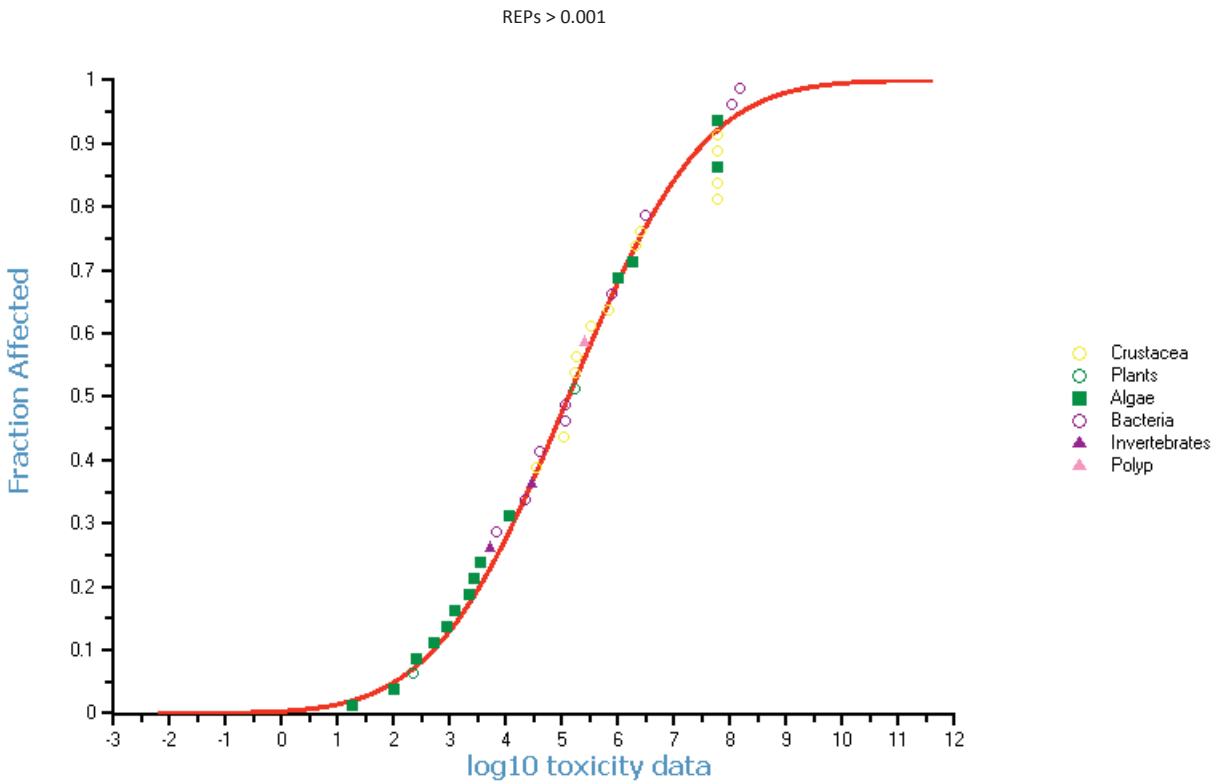


REPs > 0.1

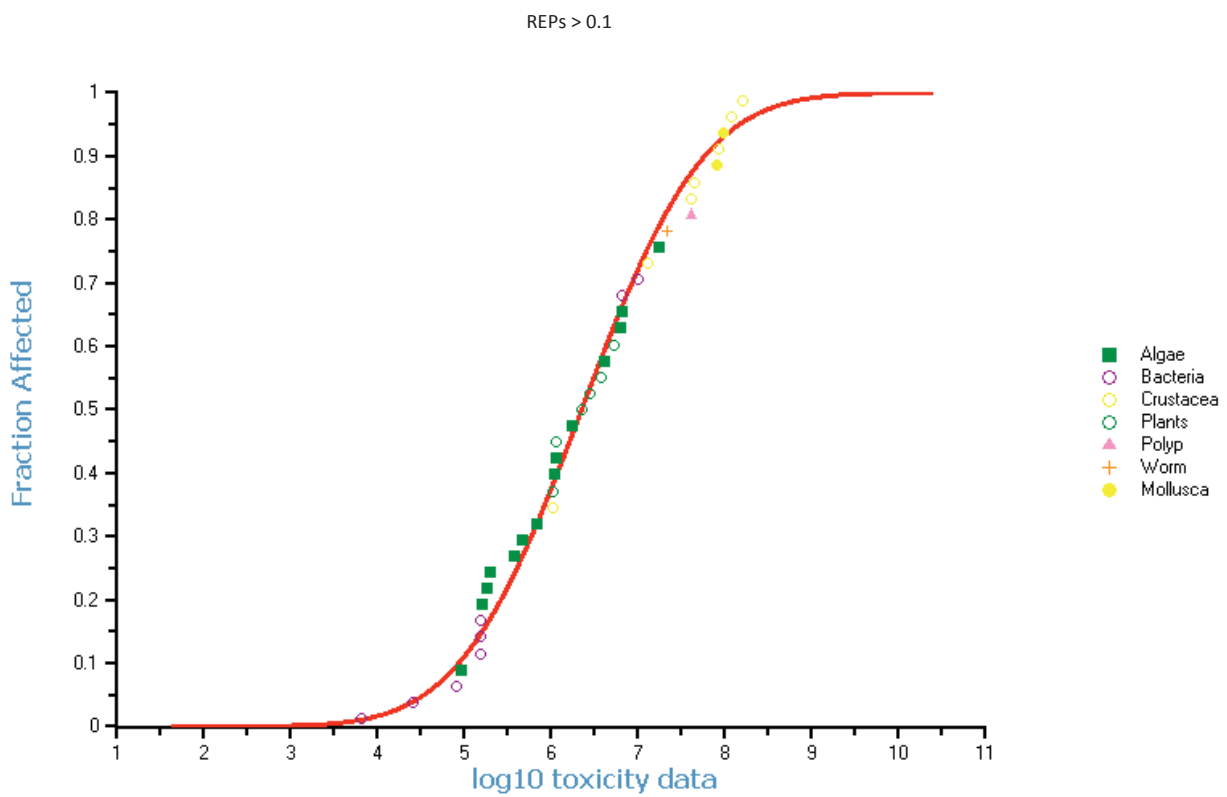
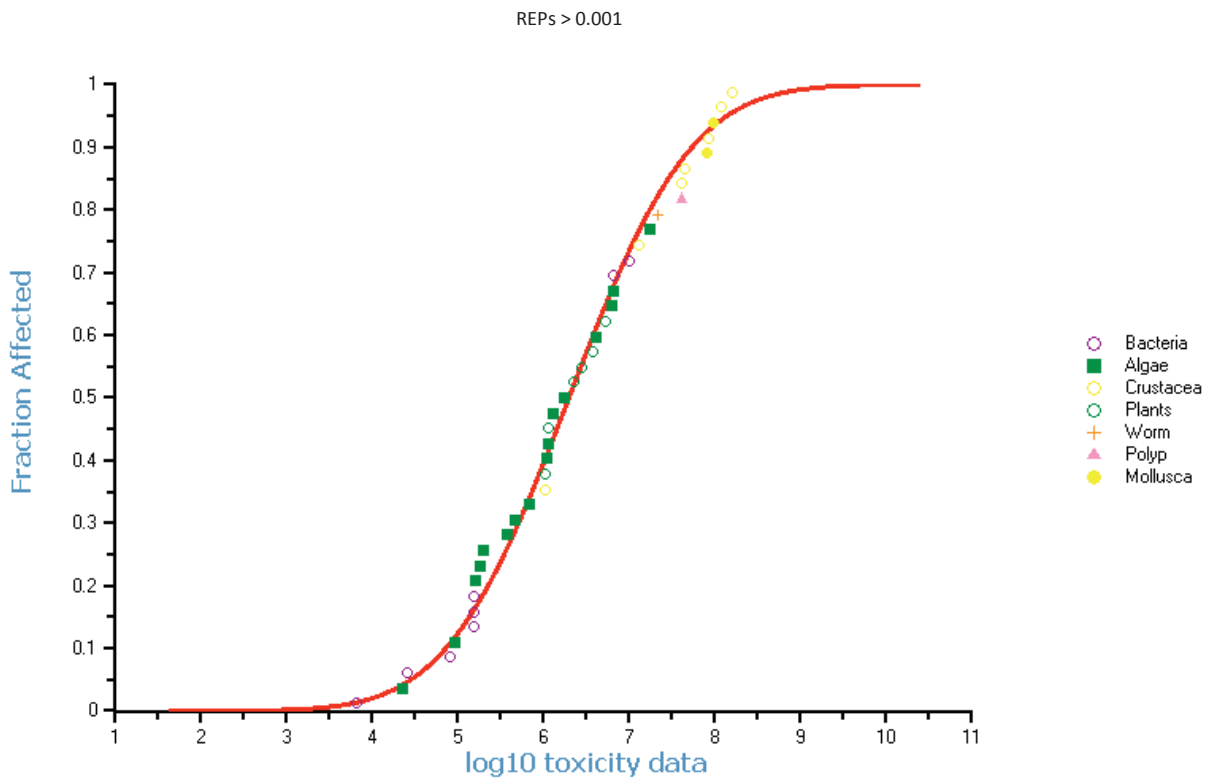




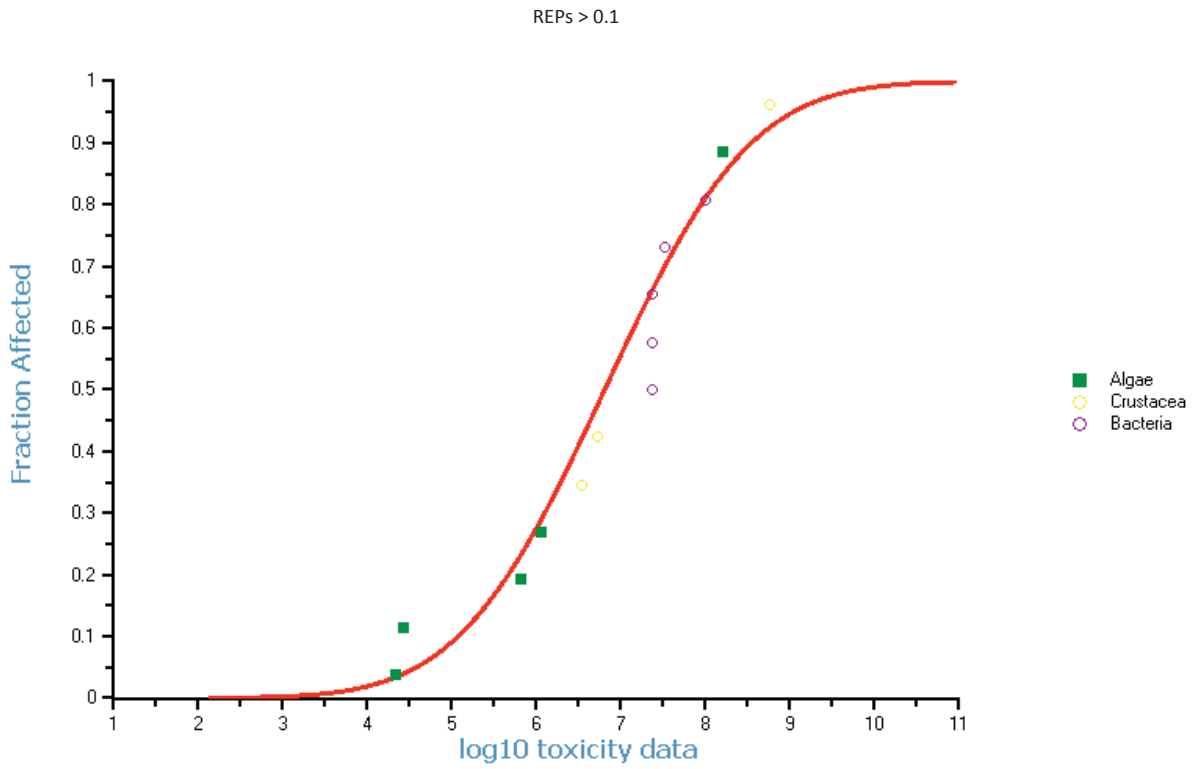
### H: PXR CALUX (NG NEQ/L)



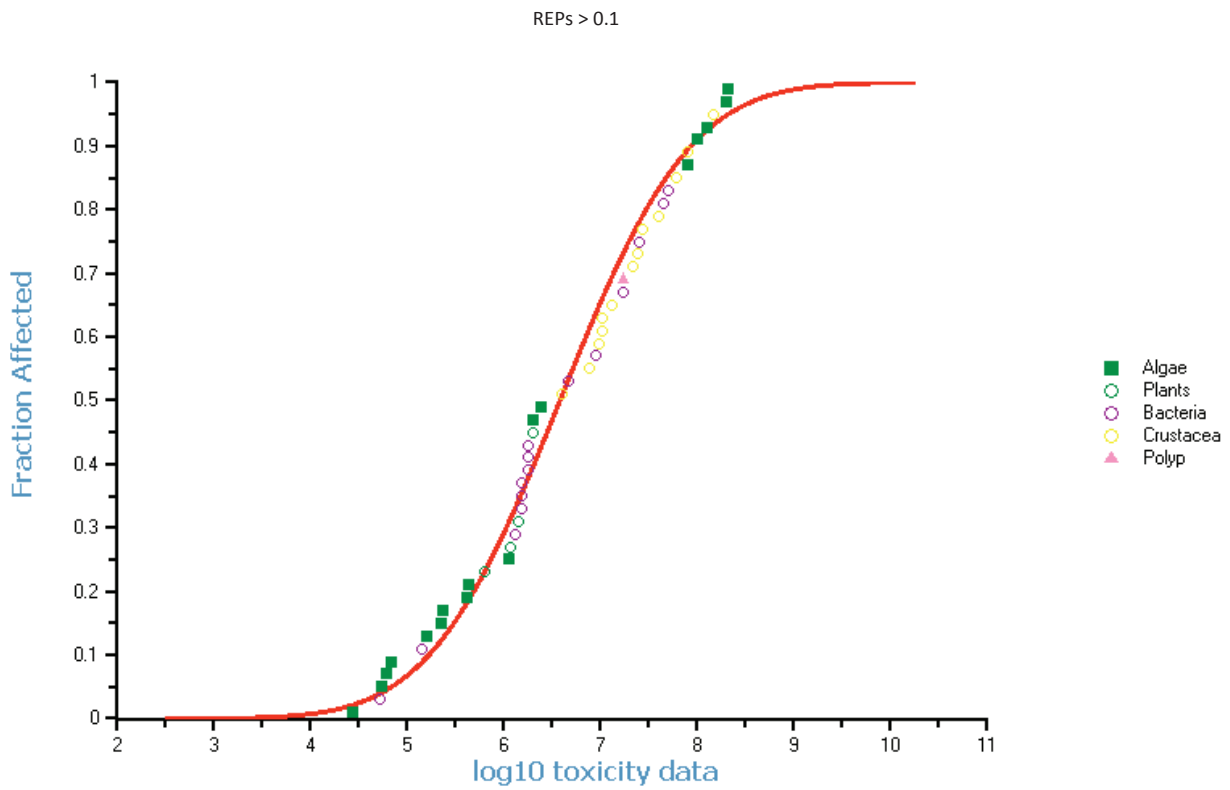
## I.1 MACROLIDES & B LACTAMS (NG PEQ/L)



## I.2 TETRACYCLINES (NG OEQ/L)

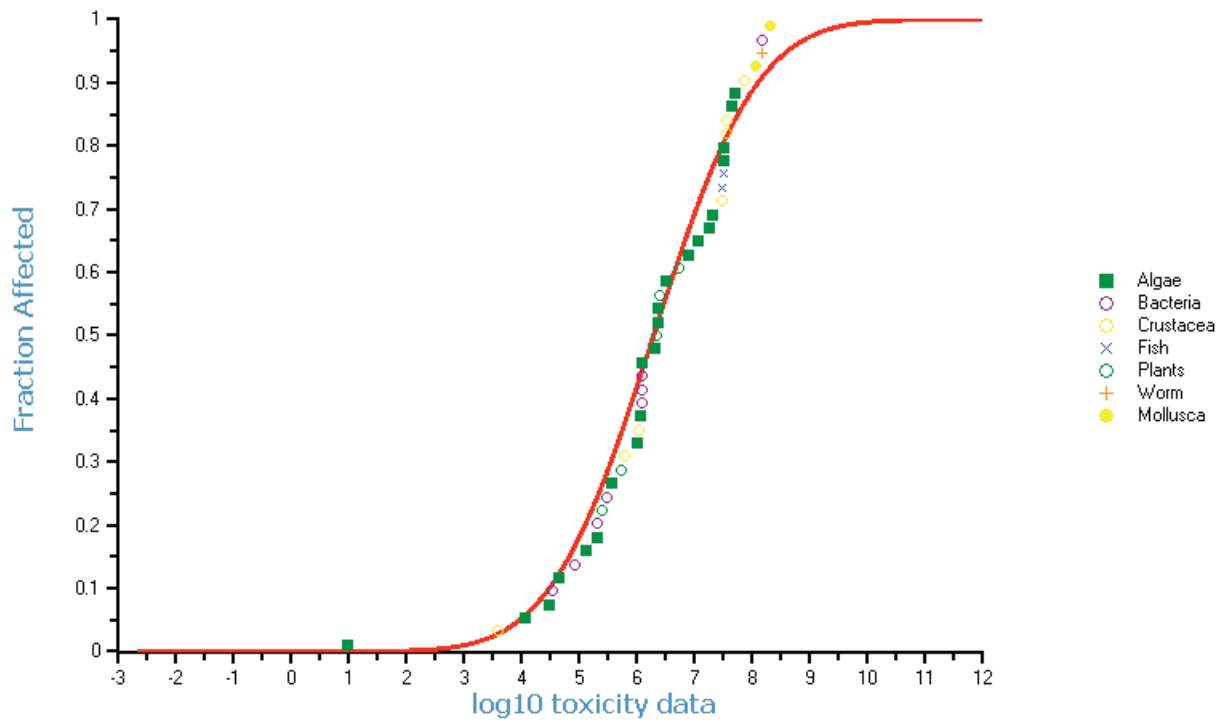


### I.3 AMINOGLYCOSIDES (NG NEQ/L)



### I.4 SULFONAMIDES (NG SEQ/L)

REPs > 0.1



## FOUNDATION FOR APPLIED WATER RESEARCH STOWA

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STOWA (Acronym for Foundation for Applied Water Research) is the knowledge centre of the regional water managers (mostly the Dutch Water Authorities) in the Netherlands. Its mission is to develop, collect, distribute and implement applied knowledge, which the water managers need in order to adequately carry out the tasks that their work supports. This expertise can cover applied technical, scientific, administrative-legal or social science fields.

STOWA is a highly demand-driven operation. We carefully take stock of the knowledge requirements of the Water Authorities and ensure that these are placed with the correct knowledge providers. The initiative for this mainly lies with the users of this knowledge, the water managers, but sometimes also with knowledge institutes and business and industry. This two-way flow of knowledge promotes modernisation and innovation.

Demand-driven operation also means that we are constantly looking for the 'knowledge requirements of tomorrow' – requirements that we dearly want to put on the agenda before they become an issue – in order to ensure that we are optimally prepared for the future.

We ease the burden of the water managers by assuming the tasks of placing the invitation to tender and supervising the joint knowledge projects. STOWA ensures that water managers remain linked to these projects and also retain 'ownership' of them. In this way, we make sure that the correct knowledge requirements are met. The projects are supervised by committees, which also comprise regional water managers. The broad research lines are spread out per field of practice and accounted for by special programme committees. The water managers also have representatives on these committees.

STOWA is not only a link between the users of knowledge and knowledge providers, but also between the regional water managers. The collaboration of the water managers within STOWA ensures they are jointly responsible for the programming, that they set the course, that several Water Authorities are involved with one and the same project and that the results quickly benefit all Water Boards.

### MISSION STATEMENT:

STOWA's fundamental principles are set out in our mission: Defining the knowledge needs in the field of water management and developing, collecting, making available, sharing, strengthening and implementing the required knowledge or arranging for this together with regional water managers.

stowa